

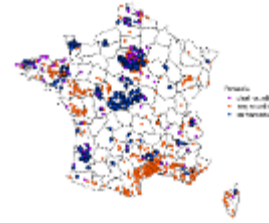
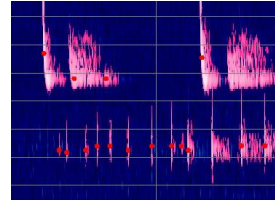
CASPA 2019



Session 1 : Participation

VIGIE-CHIRO : UN SUIVI ACQUSTIQUE STANDARDISÉ DE LA BIODIVERSITÉ TERRESTRE

YVES BAS



Vigie-Chiro : un suivi acoustique standardisé de la biodiversité terrestre



Vigie-Chiro
Suivi des chauves-souris



SON
Suivi des orthoptères nocturnes



Yves Bas, Christian Kerbiriou, Isabelle Le Viol, Stuart Newson, Kevin Barré, Grégoire Lois, Jean-François Julien

VIGIENATURE

Un réseau de citoyens qui fait avancer la science

Naturalistes



SON
Suivi des orthoptères nocturnes



Vigie-Chiro
Suivi des chauves-souris



Grand public



SPIPOLL



BIRDLAB

Gestionnaires, acteurs



Vigie-Chiro
Suivi des chauves-souris



Vigie-Chiro

- 1) Introduction aux chiroptères (chauves-souris) et leur étude par l'acoustique
- 2) Les protocoles standardisés et participatifs
- 3) L'activité acoustique comme mesure d'abondance relative
- 4) L'identification automatique
- 5) La gestion des erreurs et la qualification des données

Vigie-Chiro

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Order Chiroptera (= bats)

The only mammals performing active flight

- Chiroptères~ “Main ailée”
- > 1000 species in the world, 35 in France
- Mostly insect-eating in Europe, 3 species occasionally fishing, and 1 hunting small birds on migration.
- More diverse ecology in the tropics: foraging fruits, nectars, amphibians, other bats, etc
- Echolocation is essential for most species, for orientation and prey detection. No terrestrial equivalent. Roughly equivalent to cetacean echolocation (but much more diverse)
- Exceptional longevity for their size (41 years and 7 g)
- Ecosystem services: pest control, pollination, seed dispersal...



3 main field methods

1) Day Roost search

- 3 main roost types: tree, buildings, cavities
- Time-consuming, especially if search is systematic...



Thermal camera



Endoscope

Diplomacy...



...and safety issues!

3 main field methods

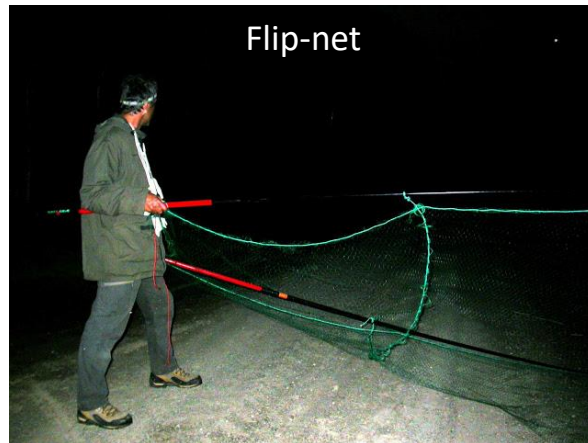


2) Capture

- With mist-nets, over water, at roost entrance or on busy flight corridors
- Low efficiency compared to birds
- Other techniques:
 - Harp-trap
 - Flip-net



Permit necessary
Long apprenticeship

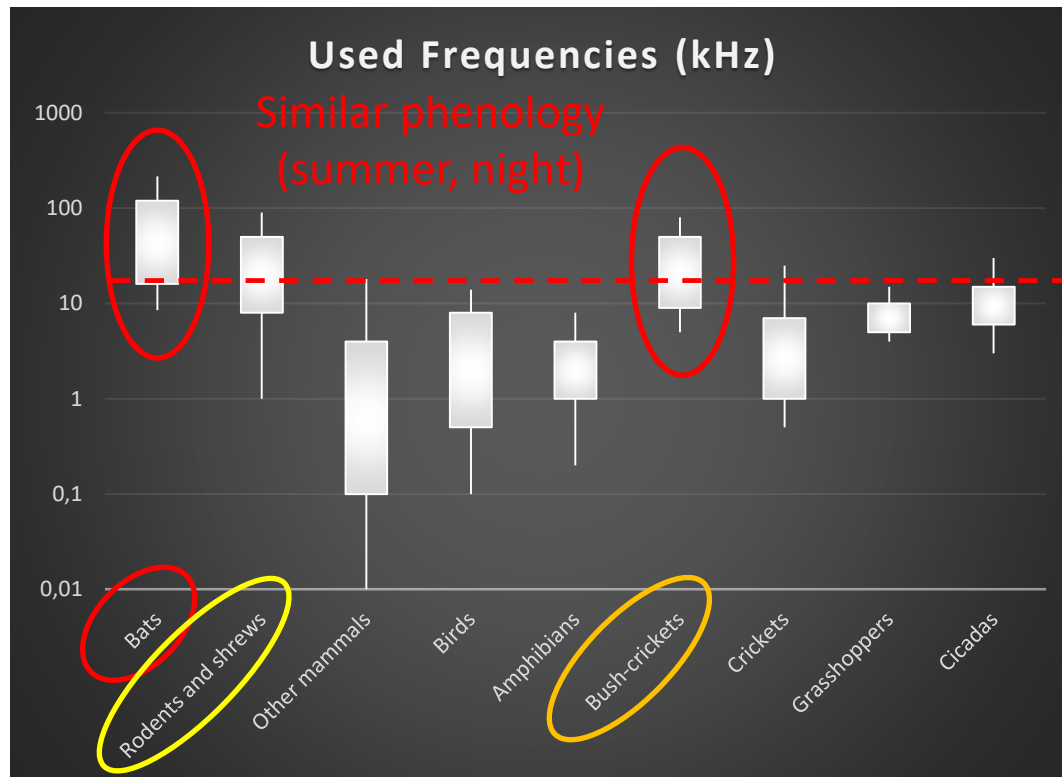


Bat survey: the sound approach

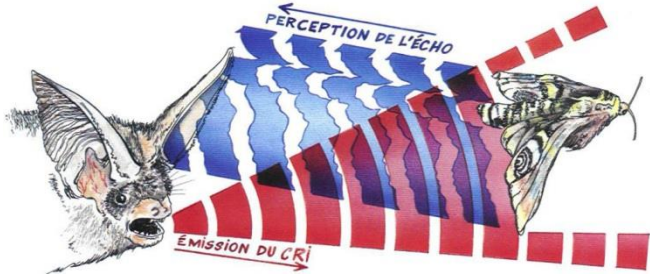
- 1) Introduction to bats and their study
- 2) Acoustic field methods**
- 3) Sampling strategies
- 4) Monitoring protocols
- 5) Acoustic activity as a measure of relative abundance
- 6) Automatic identification: why and how?
- 7) How to deal with errors?
- 8) Acoustic localisation and bat flight path reconstruction

3) Acoustics

Main acoustically active groups (long range)



3 main field methods

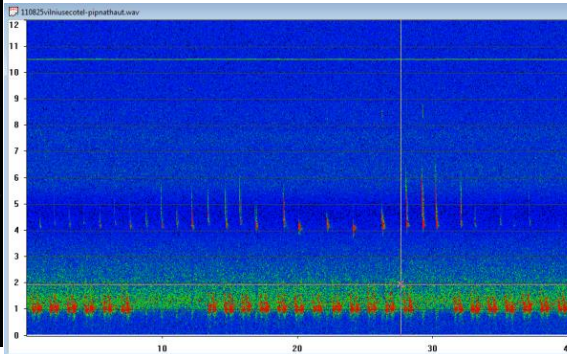


3) Acoustics

- Detectability: bats emit calls continuously during flight
 - 1 call every 1-3 wingbeats (= 2-15 calls/sec)
 - To detect preys, obstacles, water, other individuals, etc
- Limits: most of these calls are beyond ear limits > specific detectors or recorders
- 2 types of id process: (1) by ear in the field or (2) a posteriori spectrogram analysis



2000s

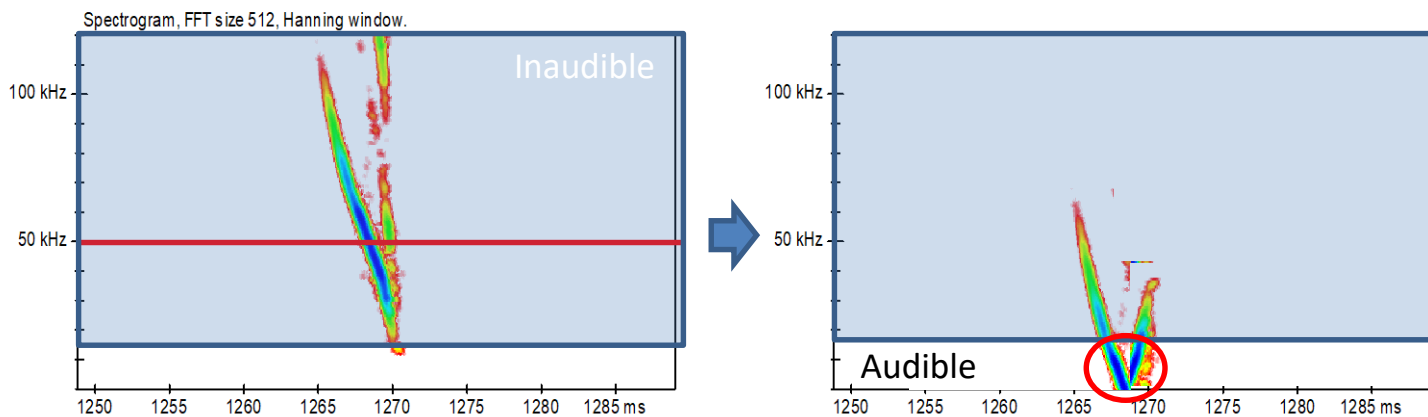




Bat identification: the sound approach

1990s: active detection

- Hearing ultrasounds > 3 techniques:
 - Frequency division: no limit in time and frequency but low taxonomic resolution (poor information)
 - Heterodyne: shifting frequencies according to a certain threshold





Bat identification: the sound approach

2000s: active detection

- Hearing ultrasounds > 3 techniques:
 - Frequency division: no limit in time and frequency but low taxonomic resolution (poor information)
 - Heterodyne: shifting frequencies according to a certain threshold - Moderate taxonomic resolution
 - Time Expansion: sound is recorded during a limited time (0.1-10s) then played back 10x slower to make frequency 10x lower. Limit: sampling time < 10%

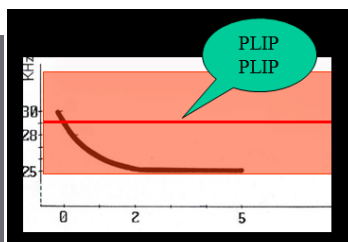




Bat identification: the sound approach

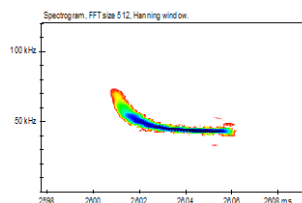
1990s: active detection

La méthode "Barataud" (traditionnelle en France) :



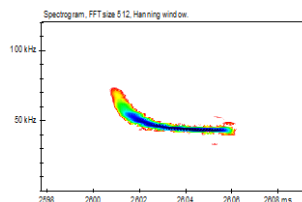
Ecoute en Hétérodyne sur le terrain

puis si ambiguïté...



Ecoute en Expansion de temps sur le terrain et/ou sur ordinateur

puis si ambiguïté...



Determination sur critères visuels (mesures)



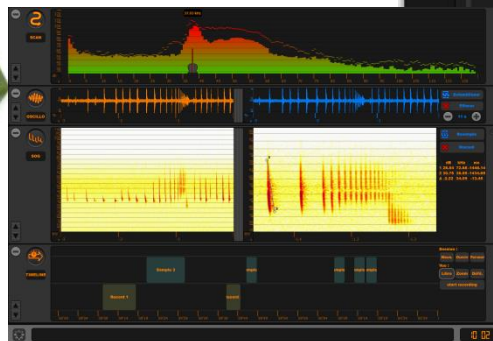


Bat identification: the sound approach

2010s: active detection

Revolution around 2010 : direct high frequency recording affordable!

➤ **Spectrogram analysis in the field!**





Bat identification: the sound approach

2010s - a new method: passive detection

Revolution around 2010 : direct high frequency recording affordable!

➤ **Autonomous long duration recordings!!**

• Features:

- Automatic trigger
- Need for fast spectrogram analysis (new id methods)
- Unattended!





Bat identification: the sound approach

2010s - a new method: passive detection

- Features:

- Automatic trigger
- Need for fast spectrogram analysis (new id methods)
- Unattended!

- Advantages :

- Sampling compounded (more sites, longer sampling)

- Disadvantages :

- A posteriori analysis is time-consuming
- Constant need for automation (softwares)

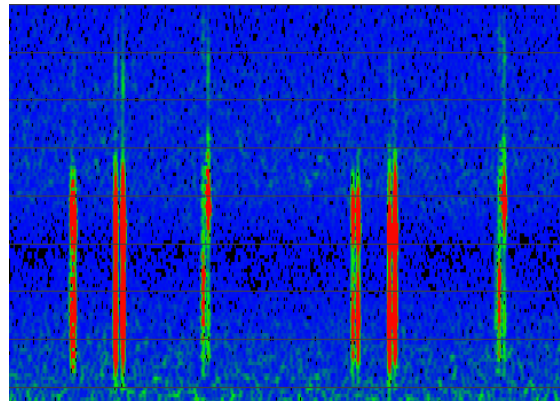




Bat identification: the sound approach

2010s - a new method: passive detection

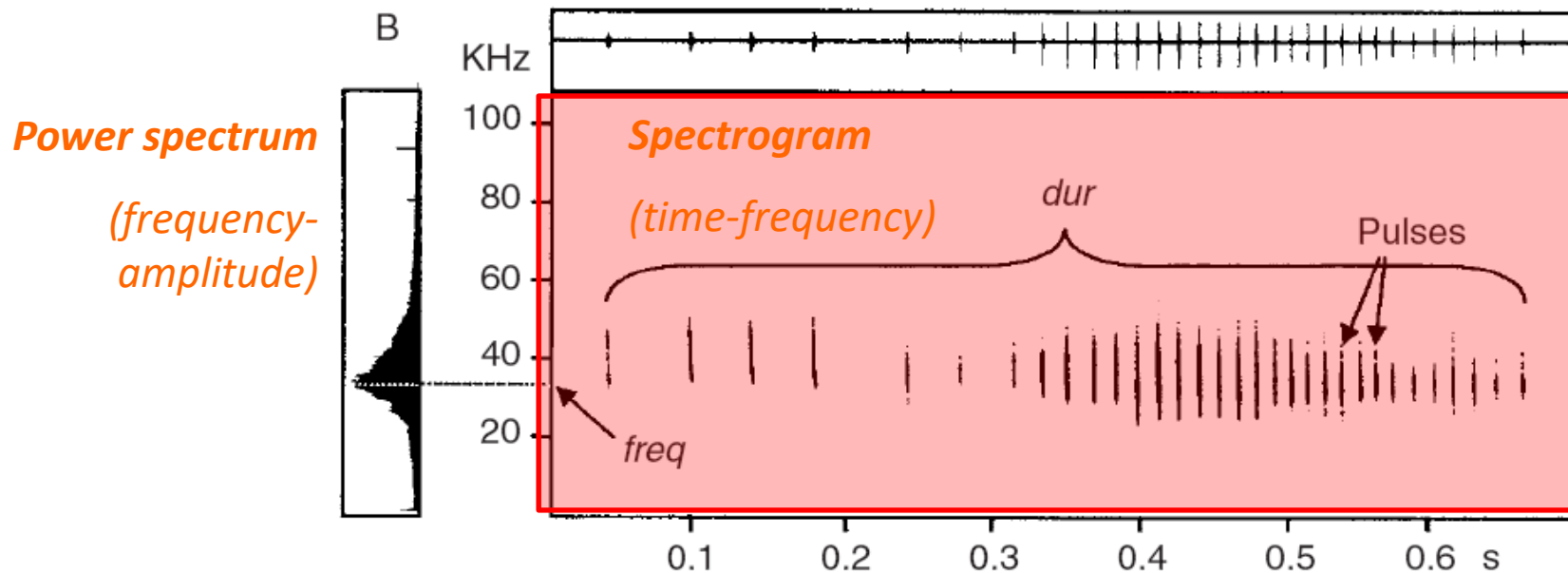
- Automatic trigger principles:
 - Recording is triggered on signal-to-noise ratio
 - Sensitive but false positives due to other "noise" (bush-crickets, rain, wind, etc) – 85 % on average
 - Need for smarter post-processing

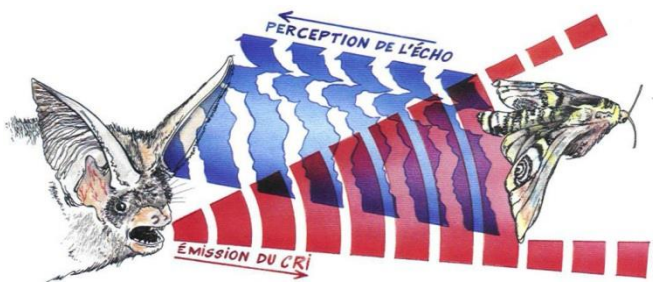


Bat identification: the sound approach

Sound visualisation

Oscillogram (time-amplitude)



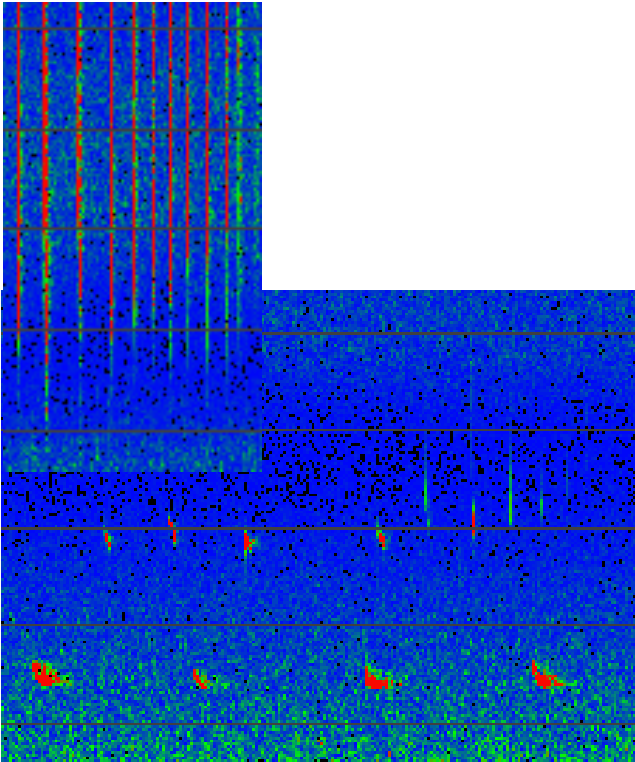


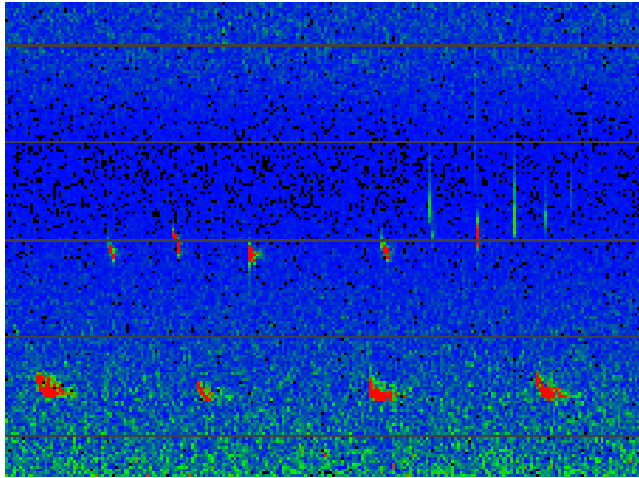
Bat identification: the sound approach

Ecology and echolocation

- Rythm:

- 1 call every 1-3 wingbeats = **2-15 calls/sec**
- **15 calls / seconde** = 15 “frames” per seconde, smooth 😊 but detection radius limited to 10 m ☹️
 - Gleaning or hawking of small abundant preys
- **2 calls / seconde** = “stroboscopic” ☹️ but detection radius of 80 m 😊
 - Large species foraging less abundant preys in open areas



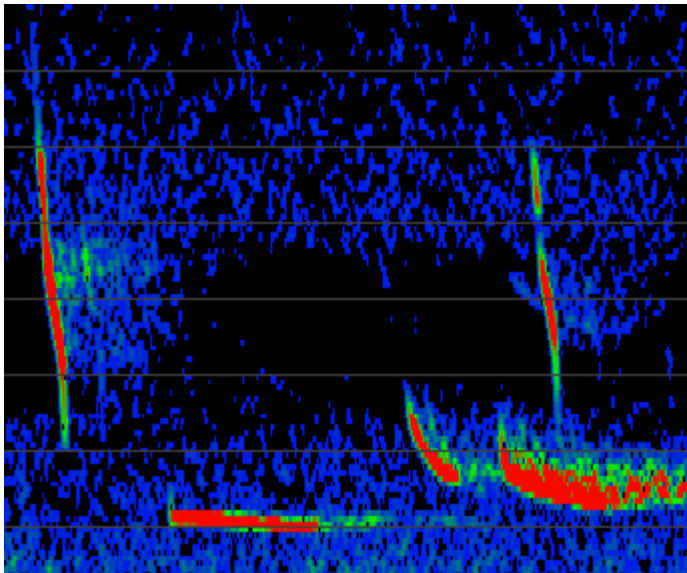


Bat identification: the sound approach

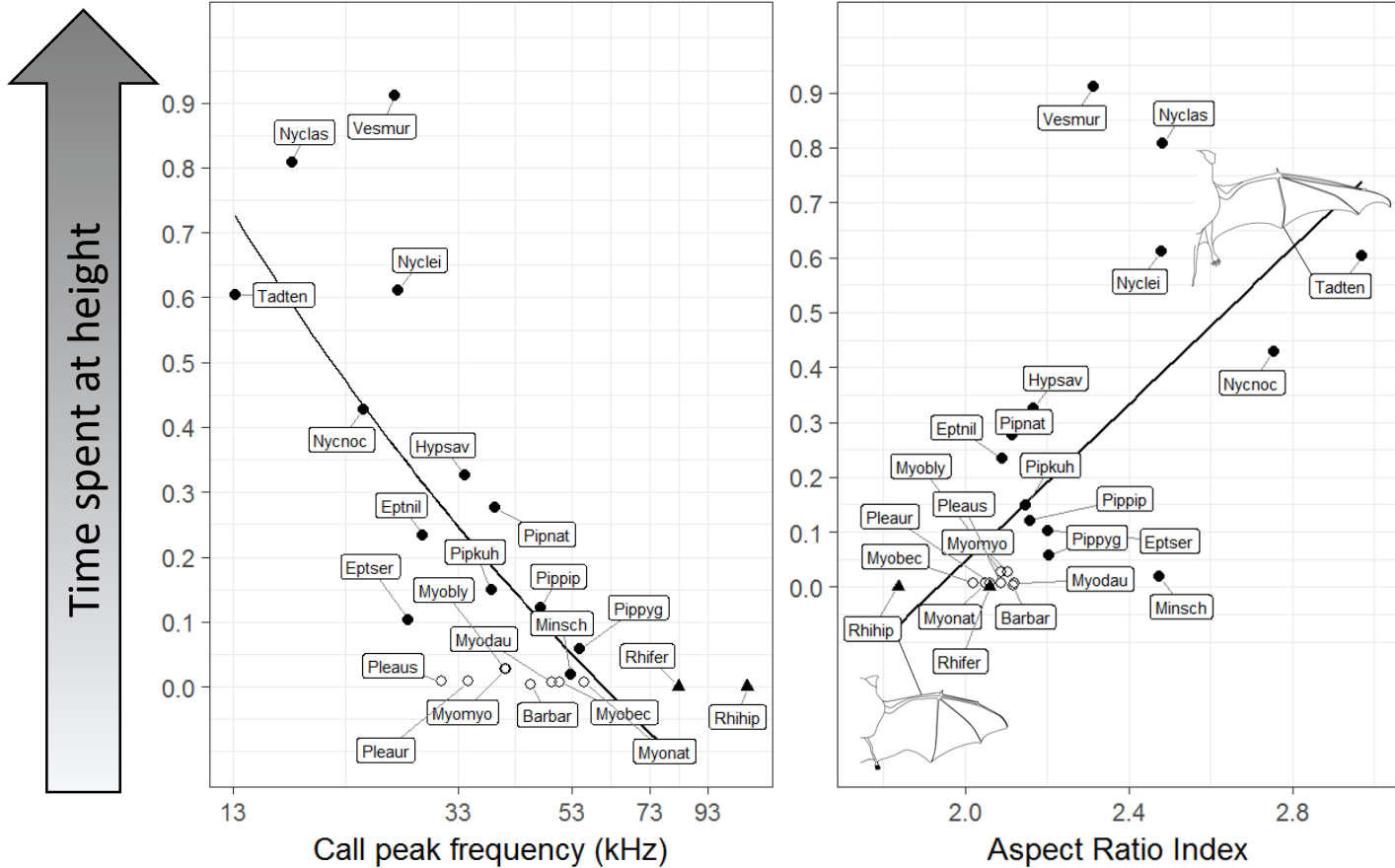
Ecology and echolocation

- **Frequency:**

- The higher the frequency, the smaller preys will return an echo (ex : small diptera or moth) 😊, but stronger attenuation: small detection radius ☹️
 - Example : Soprano Pipistrelle, specialized on mosquitoes
- Inversely, low frequency echolocation will detect only large preys (specialisation) but will have larger detection radius
 - Example : Free-tailed bats fly high to detect preys in large volumes.



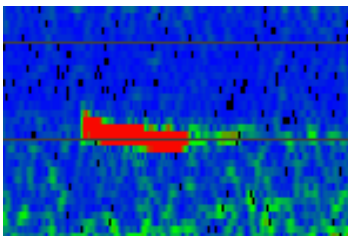
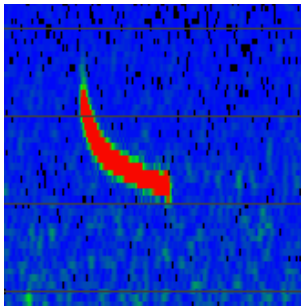
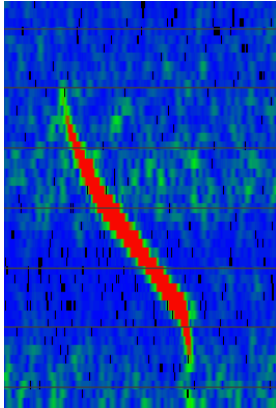
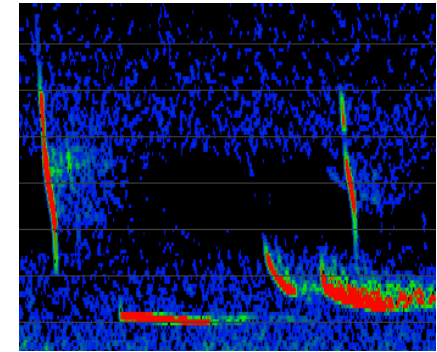
Bats: a unique sensory-motor system



- Bat traits co-evolved with species niche

Roemer et al. submitted

Bat identification: the sound approach



Ecology and echolocation

• Frequency modulation:

- Large bandwidth:
 - Accurate range resolution (=distance to prey)
 - Able to detect preys on vegetation (= gleaning)
 - Long-eared bats and Myotis in Europe
 - Disadvantage : small detection radius ☹
- Inversely, open area hawkers tend to concentrate most energy in a narrow bandwidth
 - Advantage: maximise detection radius ☺
 - Increase modulation in approach phase
 - Pipistrelles, Serotines, Noctules, etc

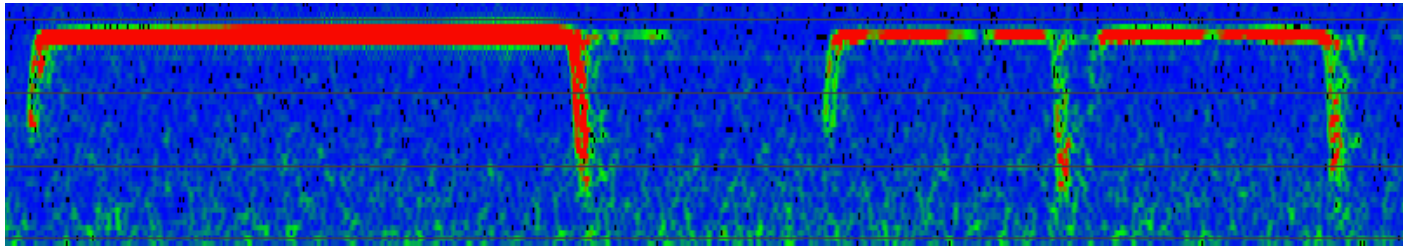


Bat identification: the sound approach

The doppler technique: Horseshoe bats

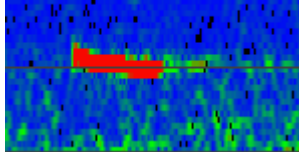
- **Drastically different techniques:**

- Long constant frequency calls to exploit Doppler shifts information:
 - Able to detect moving preys, even in cluttered environment
 - Able to forage from a branch
 - Anticipating prey movements (moth)
- Angular resolution thanks to constant ear movements



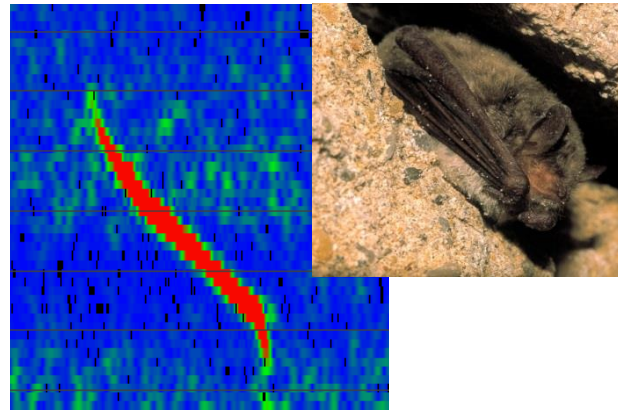
Bat identification: the sound approach

European acoustic types



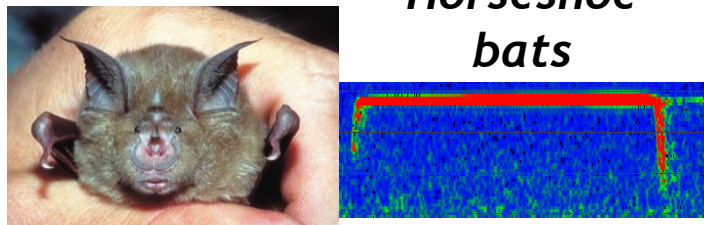
High flight
(largest group)

Myotis (large bandwidth)

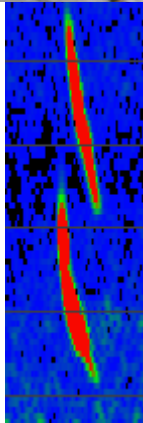


Barbastelle Bat

Horseshoe bats



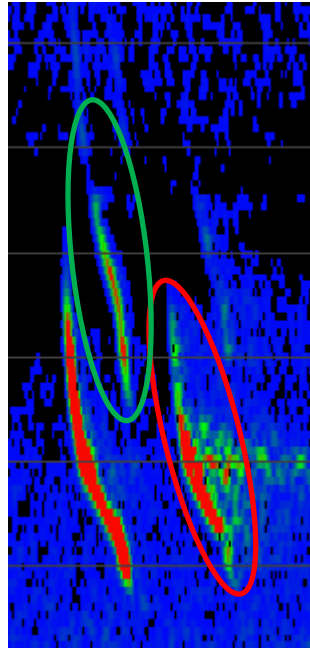
Long-eared bats



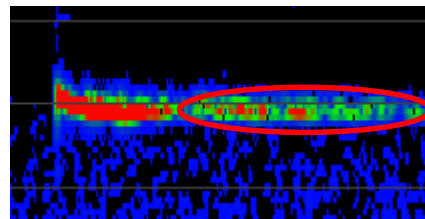
Bat identification: the sound approach

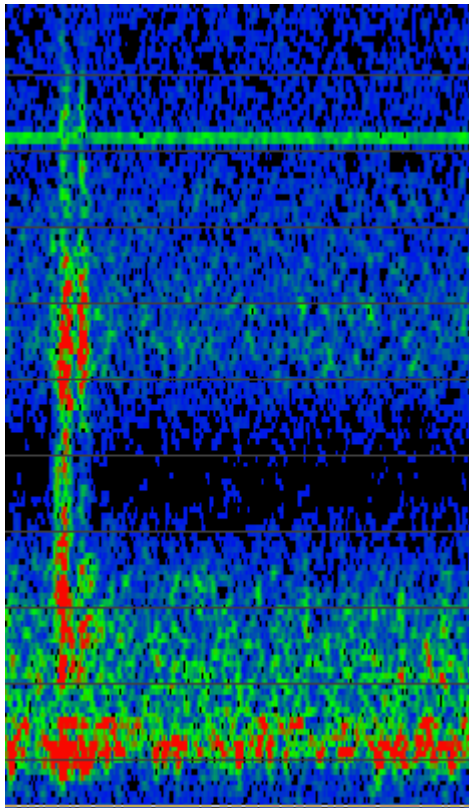
Some traps in spectrogram analysis

*Harmonics (frequency
x2, x3, ...)*



Echoes



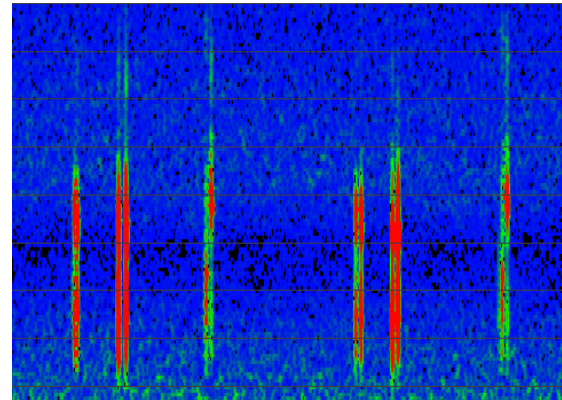
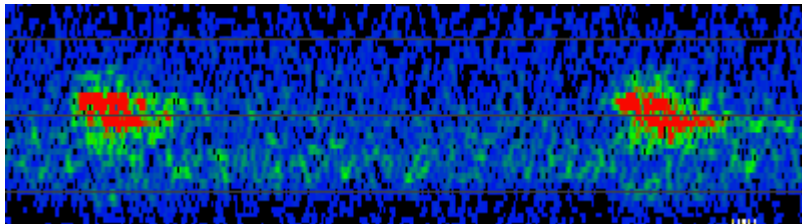


Bat identification: the sound approach

Some traps in spectrogram analysis

- **Recognizing non bat ultrasounds**

- Usually non pure tones
- Irregular rhythms
- Usually extremely short (wind, rain, bush-crickets)
- Or very long (metallic sounds, electronic parasites, birds, rodents, ...)

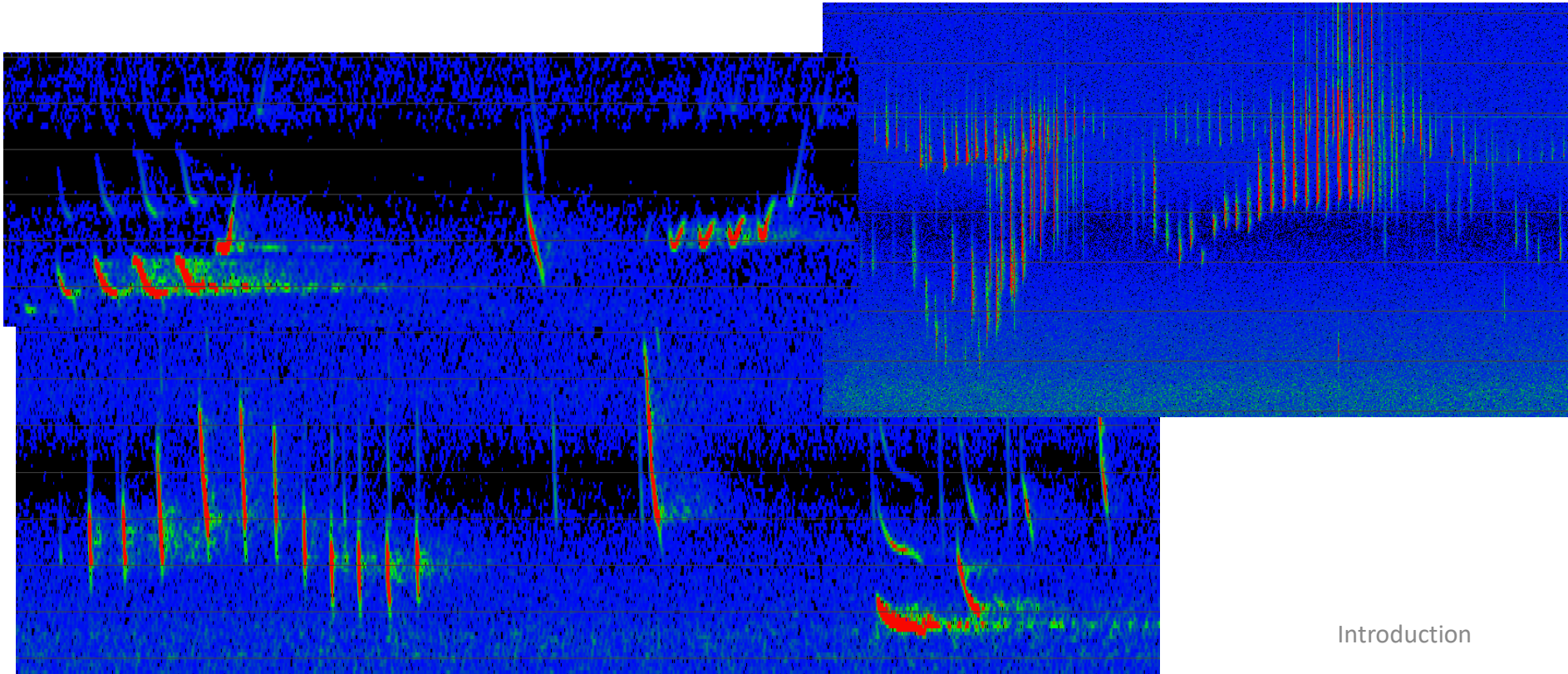


Bat identification: the sound approach

Lack of knowledge to be identified in many cases

Some traps in spectrogram analysis

- **Recognising social calls**
 - Chaotic frequency shifts
 - Structurally more complex

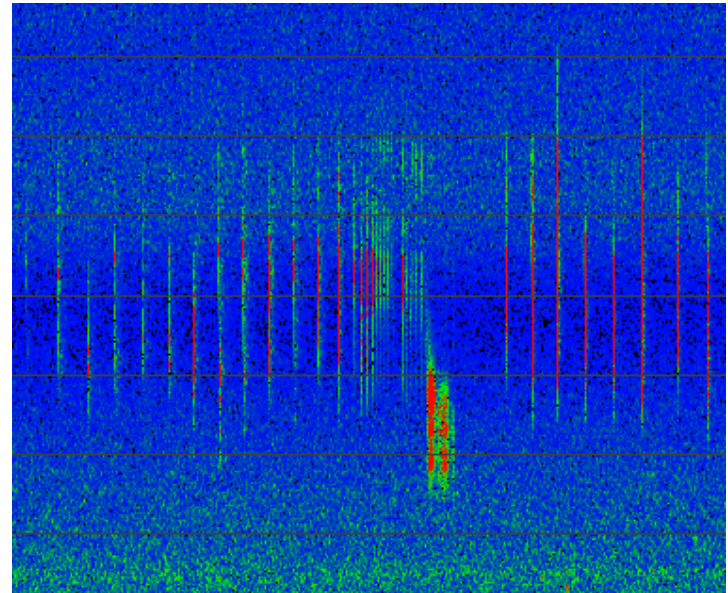
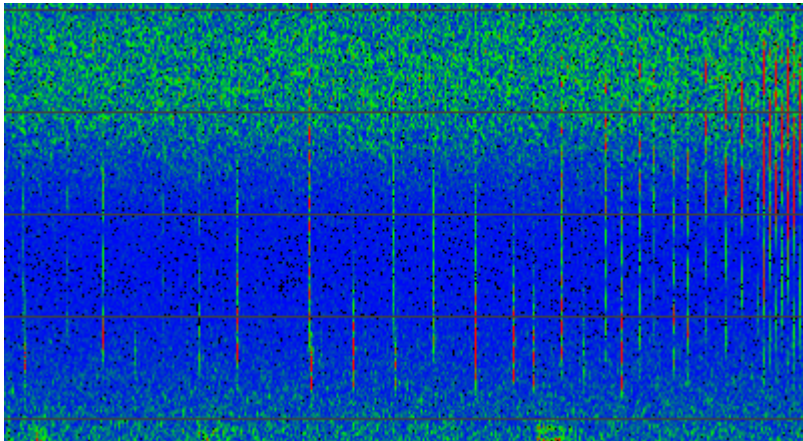
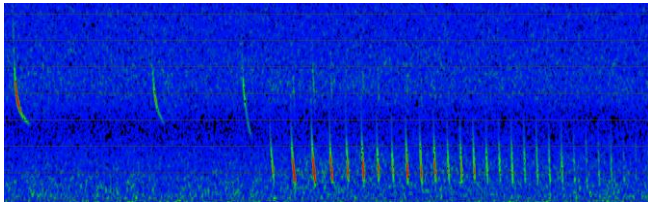


Bat identification: the sound approach

Some traps in spectrogram analysis

- **Recognising « buzzes »**

= progressive acceleration until inter-pulse intervals < 10 ms :
Typical of prey capture sequences, but also when drinking,
landing, etc.

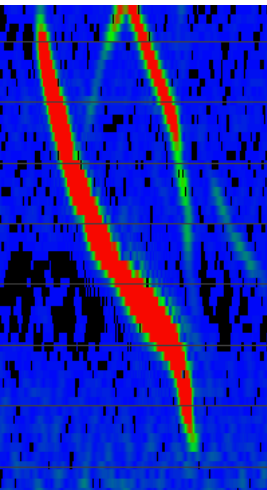


Bat identification: the sound approach

Some traps in spectrogram analysis

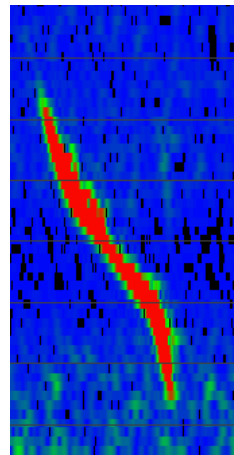
- **Strong and heterogeneous environmental attenuation**

Exemple of the very same call recorded by 4 microphones



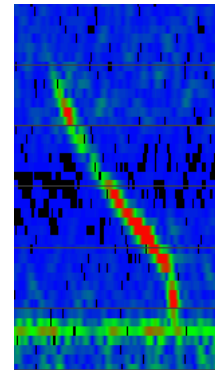
EF = 22 kHz
BW = 73 kHz
Dur = 5.7 ms
FME = 45 kHz

Distance = 6 m
Angle = 5°



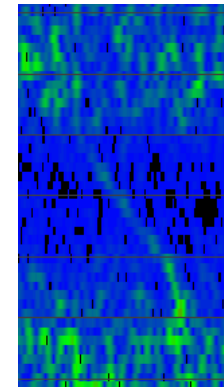
EF = 23 kHz
BW = 53 kHz
Dur = 5.0 ms
FME = 58 kHz

Distance = 9 m
Angle = 60°



EF = 28 kHz
BW = 41 kHz
Dur = 4.5 ms
FME = 42 kHz

Distance = 8 m
Angle = 170°



EF = 30 kHz
BW = 26 kHz
Dur = 3.2 ms
FME = 38 kHz

Distance = 10 m
Angle = 120°

Increasing distance and angle

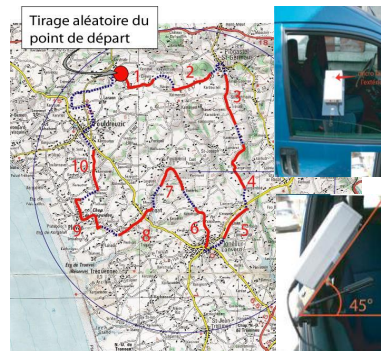
Vigie-Chiro

- 1) Introduction aux chiroptères (chauves-souris) et leur étude par l'acoustique
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Bat acoustic monitoring: principles

- Standardized recordings (=repeatable measurement)
 - Same locations
 - Same periods
 - Same detectability
 - Trigger sensitivity
 - Microphone type
- Different methods depending on public and material

Walk transects



Car transects

Whole-night recordings



Vigie-Chiro: 3 different data collection protocols

Starting 2006

Car transects

10 x 2 km / 25 km/h

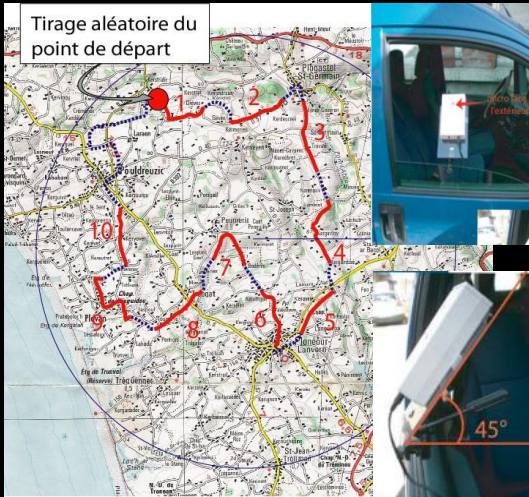
Point counts

10 points / 2x2 km squares

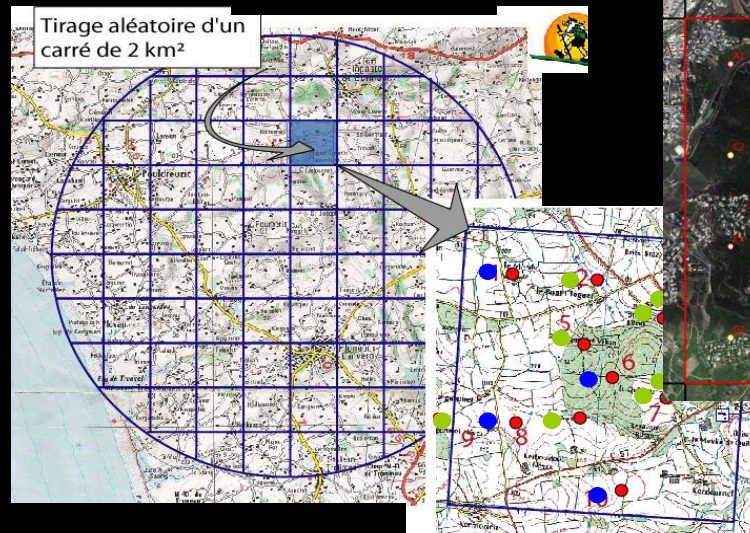
NEW
2014

Stations

1 - 10 points
/ carrés 2x2 km



Following iBat
international scheme



6 minutes
recording



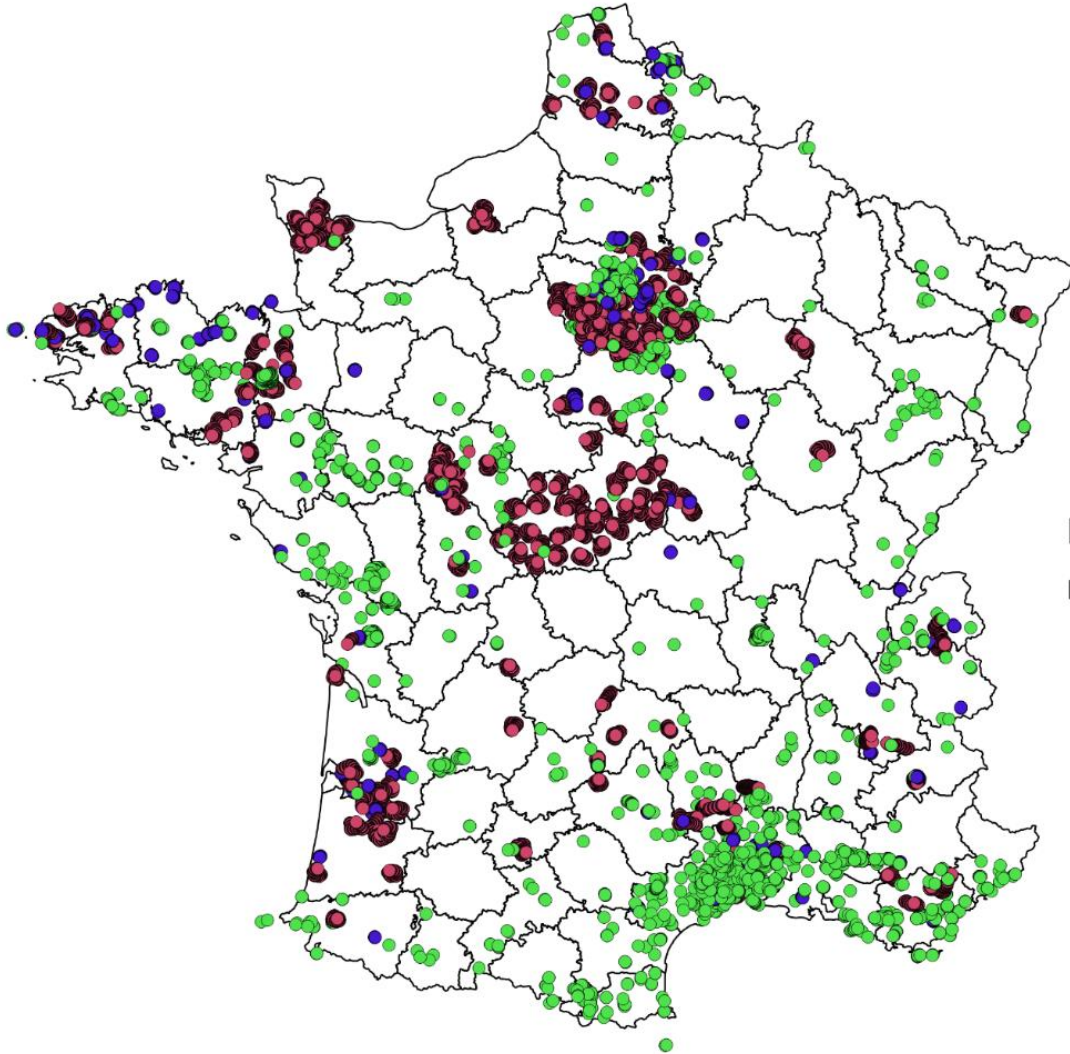
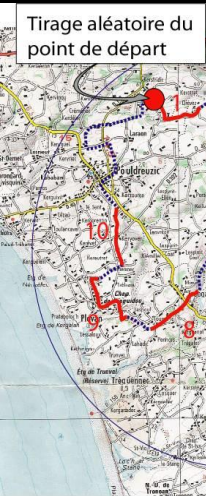
Whole night
recordings

What is the relative contribution of each protocols to monitor bats and bush-crickets?
Is there some complementarity?

Vigie-Chiro: 3 different data collection protocols

Car
10 x 2

tions
points
2x2 km



Légende

Protocoles

- PEDESTRE
- POINT_FIXE
- ROUTIER



night
dings



Foll
intern

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Monitoring protocols comparison

Testing trend detection power:

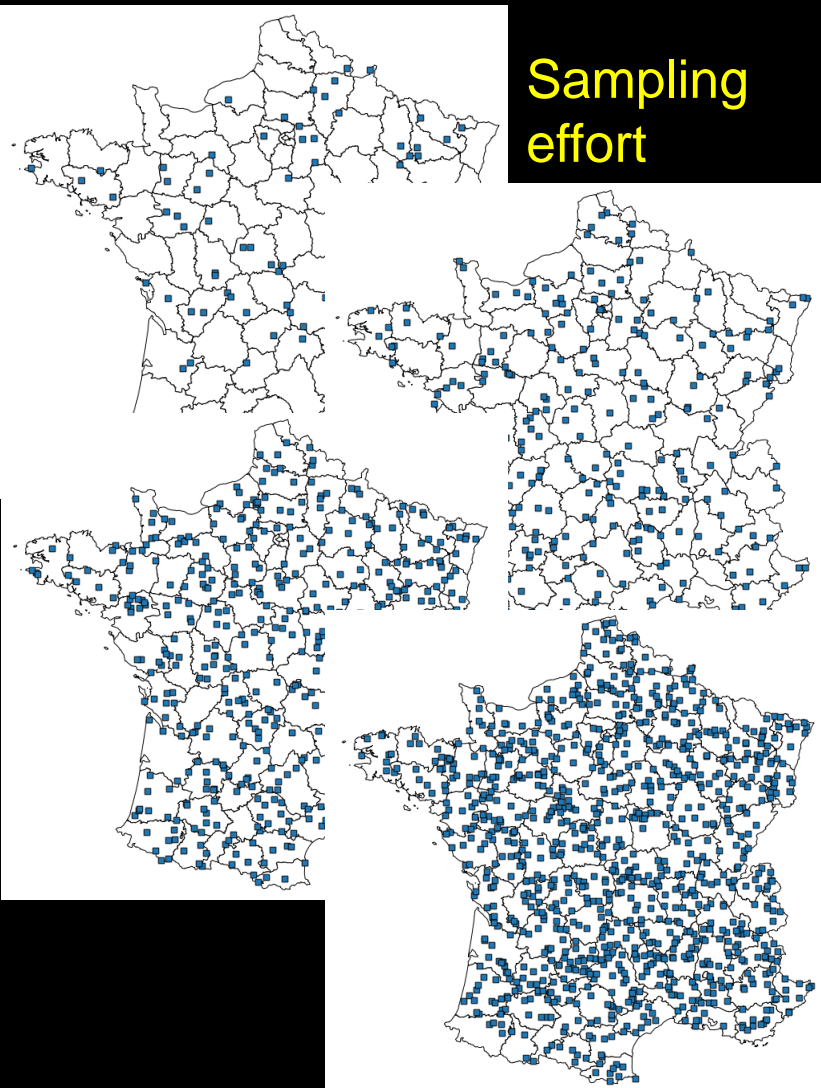
Scenarios:	Car transects	Point counts	Stations
Pessimistic	50	300	100
Current	100	1000	250
Optimistic	400	3000	1000

Simulations fed with Vigie-Chiro data input:

- Average measured activity (*corrected by latitude/longitude*)
- Spatial variability (*among sites variance*)
- Temporal variability (*among same-year visits variance*)

Monitoring protocols comparison

Testing trend detection power:



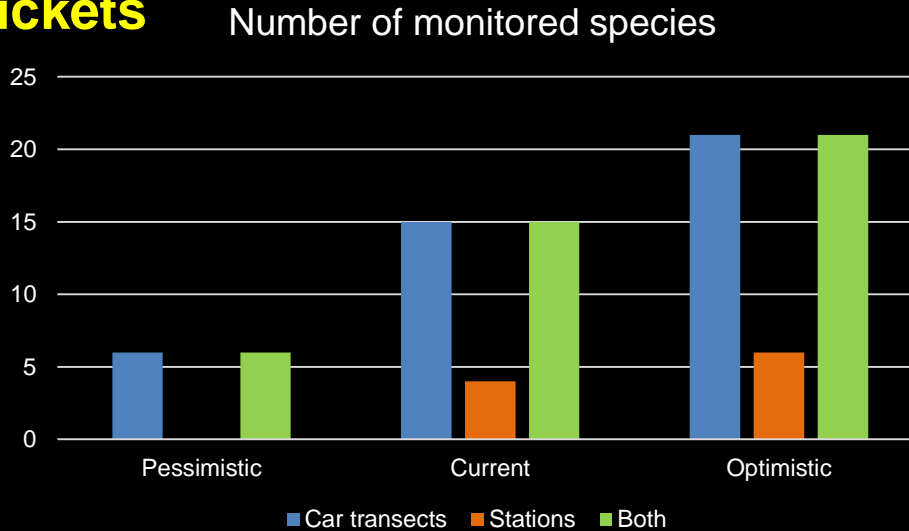
Simulations:

- 11 successive years sampling
- Negative binomial distributions (both spatially and temporally)
- **30% decreasing trends** (= smallest decreasing trends that could justify listing VU (IUCN))
- 2 nights recorded / year / points or transects
- 500 data sets for n points
- GLM: Bat passes \sim Time + Site

=> Target: getting significant effect of time (z-test, $p < 0.05$) on > 50% simulated datasets

Monitoring protocols comparison

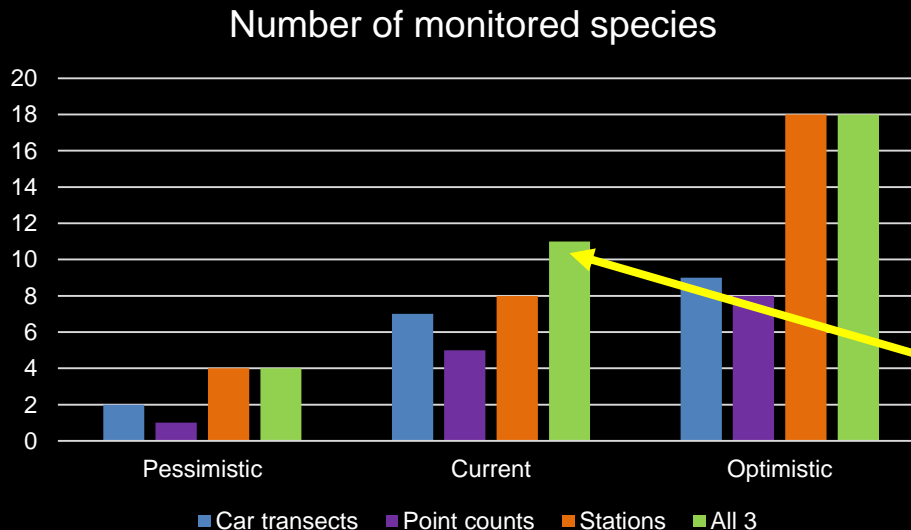
Bush-crickets



Car transects more efficient (immobility of singing individuals)

...but representativity bias (road edges) to be control by other protocols

Bats



Stations protocol has great potential to increase number of monitored species

...but good complementarity of the 2 other schemes

Monitoring protocols comparison

Bats under current scenario

Species monitored	Car transects	Point counts	Stations
<i>Barbastella barbastellus</i>	○	○	×
<i>Eptesicus serotinus</i>	×	○	○
<i>Miniopterus schreibersii</i>	○	○	×
<i>Myotis daubentonii</i>	○	×	×
<i>Nyctalus leisleri</i>	×	×	○
<i>Nyctalus noctula</i>	×	○	○
<i>Pipistrellus kuhlii</i>	×	×	×
<i>Pipistrellus nathusii</i>	×	×	×
<i>Pipistrellus pipistrellus</i>	×	×	×
<i>Pipistrellus pygmaeus</i>	×	○	×
<i>Plecotus</i> spp.	○	○	×

Monitoring protocols comparison

Bats under current scenario

Species monitored	Car transects	Point counts	Stations
<i>Barbastella barbastellus</i>	○	○	×
<i>Eptesicus serotinus</i>	×	○	○
<i>Miniopterus schreibersii</i>	○	○	×
<i>Myotis daubentonii</i>	○	×	×
<i>Nyctalus leisleri</i>	×	×	○
<i>Nyctalus noctula</i>	×	○	○
<i>Pipistrellus kuhlii</i>	×	×	×
<i>Pipistrellus nathusii</i>	×	×	×
<i>Pipistrellus pipistrellus</i>	×	×	×
<i>Pipistrellus pygmaeus</i>	×	○	×
<i>Plecotus spp.</i>	○	○	×

Large, detectable, crepuscular species are best monitored by car transects

Monitoring protocols comparison

Bats under current scenario

Species monitored	Car transects	Point counts	Stations
<i>Barbastella barbastellus</i>	○	○	✕
<i>Eptesicus serotinus</i>	✕	○	○
<i>Miniopterus schreibersii</i>	○	○	✕
<i>Myotis daubentonii</i>	○	✕	✕
<i>Nyctalus leisleri</i>	✕	✕	○
<i>Nyctalus noctula</i>	✕	○	○
<i>Pipistrellus kuhlii</i>	✕	✕	✕
<i>Pipistrellus nathusii</i>	✕	✕	✕
<i>Pipistrellus pipistrellus</i>	✕	✕	✕
<i>Pipistrellus pygmaeus</i>	✕	○	✕
<i>Plecotus spp.</i>	○	○	✕

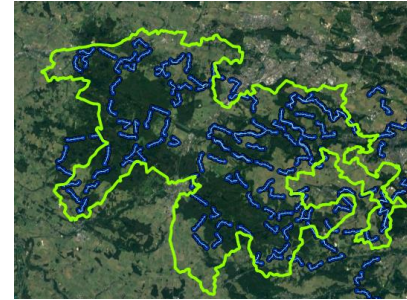
Species with constant late night activity are best monitored by stationary recordings

Vigie-Chiro

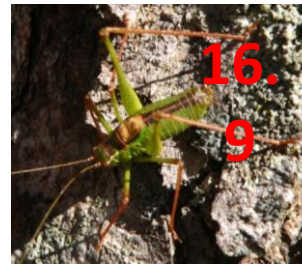
- 1) Introduction aux chiroptères (chauves-souris) et leur étude par l'acoustique
- 2) Les protocoles standardisés et participatifs
- 3) **L'activité acoustique comme mesure d'abondance relative**
- 4) L'identification automatique
- 5) La gestion des erreurs et la qualification des données

Acoustic activity as a measure of relative abundance

Interpreting data locally



Nb of call
sequences /
20 km
transect

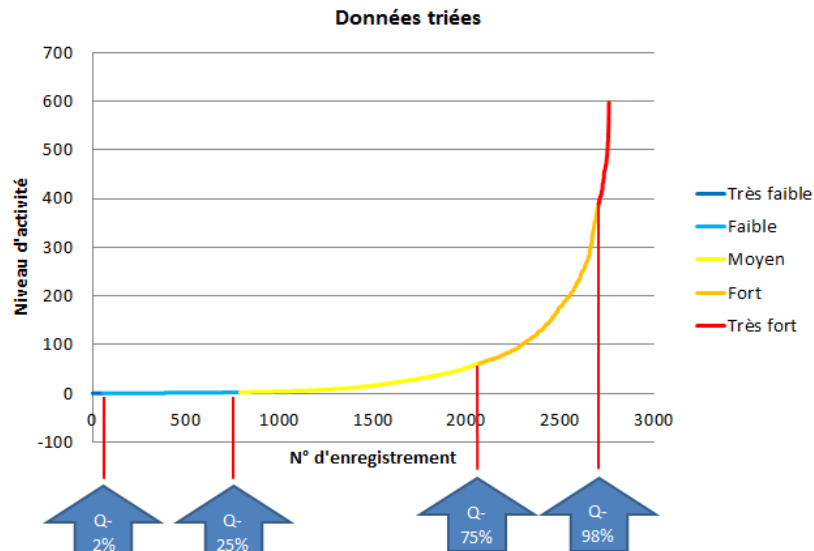


Raw data quite
uninformative



Benchmarking activity

→ Simple principle, powerful tool (cf. Haquart 2013)



Déclinable (*avec suffisamment de données*)

:

- Par région
- Par habitats
- Par saisons
- Par types de Behaviour (buzz, social/swarming...)
- Etc

Benchmarking activity

→ En pratique, c'est une grille de lecture :

Référentiels d'activité des protocoles Vigie-Chiro

Les valeurs données dans le tableau ci-dessous sont des nombres de contacts cumulés sur l'ensemble du circuit routier ou sur l'ensemble du carré pédestre ou encore sur une nuit complète en point fixe, selon le protocole. Elles permettent d'interpréter objectivement l'activité mesurée sur vos sites :

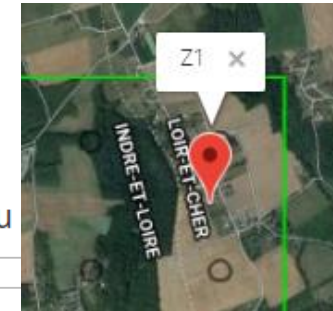
- Si vous mesurez une activité supérieure à la valeur **Q98%**, c'est que vous avez obtenu une activité **très forte**, particulièrement notable pour l'espèce
- Si vous mesurez une activité supérieure à la valeur **Q75%**, c'est que vous avez obtenu une activité **forte**, révélant l'intérêt de la zone pour l'espèce
- Si vous mesurez une activité supérieure à la valeur **Q25%**, c'est que vous avez obtenu une activité **modérée**, donc dans la norme nationale
- Si vous mesurez une activité inférieure à la valeur **Q25%**, vous pouvez considérer l'activité comme **faible** pour l'espèce

Espece	Protocole Routier			Protocole Pédestre			Protocole Point Fixe		
	Q25%	Q75%	Q98%	Q25%	Q75%	Q98%	Q25%	Q75%	Q98%
<i>Barbastella barbastellus</i>	1	2	7	1	7	10	1	15	406
<i>Eptesicus serotinus</i>	1	7	18	1	4	22	2	9	69
<i>Hypsugo savii</i>	3	13	23				3	14	65
<i>Miniopterus schreibersii</i>							2	6	26
<i>Myotis bechsteinii</i>							1	4	9
<i>Myotis daubentonii</i>	1	3	11	2	10	92	1	6	264
<i>Myotis emarginatus</i>							1	3	33
<i>Myotis blythii/myotis</i>							1	2	3
<i>Myotis mystacinus</i>							2	6	100
<i>Myotis cf. nattereri</i>	1	2	4	1	5	8	1	4	77
<i>Nyctalus leisleri</i>	2	7	18	2	7	42	2	14	185
<i>Nyctalus noctula</i>	2	7	18	1	8	25	3	11	174
<i>Pipistrellus kuhlii</i>	2	9	33	3	20	71	17	191	1182
<i>Pipistrellus nathusii</i>	1	10	36	1	4	44	2	13	45
<i>Pipistrellus pipistrellus</i>	35	95	163	13	59	119	24	236	1400

<http://vigienature.mnhn.fr/page/des-referentiels-pour-interpreter-vos-donnees>

Benchmarking activity

→ Exemple (1/2) :



Participation du 2 sept. 2016 20:03:00 effectuée par Yohan Douvneau

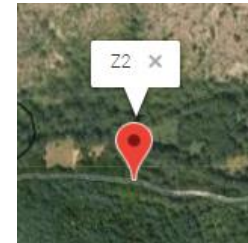
Search:

Nom français	Nom scientifique	Risque d'erreur (%)	Nb Validations	Effort de validation	Nb de Contacts par Nuit	Niveau d'Activite
Pipistrelle commune	Pipistrellus pipistrellus	1	26	FORT	949	FORTE
Serotine commune	Eptesicus serotinus	1	6	FORT	63	FORTE
Pipistrelle de Kuhl	Pipistrellus kuhlii	1	8	FORT	42	MODEREE
Noctule commune	Nyctalus noctula	1	1	SUFFISANT	38	FORTE
Noctule de Leisler	Nyctalus leisleri	1	2	FORT	26	FORTE
Oreillard gris	Plecotus austriacus	1	2	FORT	21	FORTE

Benchmarking activity

→ Exemple (2/2), consecutive nights:

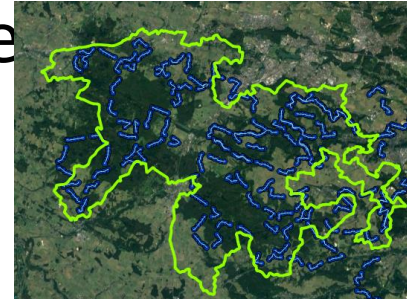
Participation du 11 juil. 2016 21:00:00 effectuée par Fédération des réserves naturelles catalanes
Vigiechiro - Point Fixe-660437



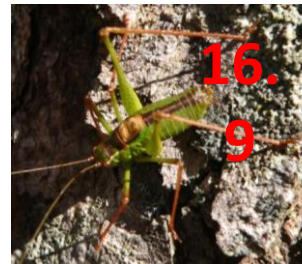
Code	Groupe	Nom français	Nom scientifique	Risque d'erreur (%)	TriGroupe	11	12	13	14	15	16	17
Barbar	Chauve-souris	Barbastelle d'Europe	Barbastella barbastellus	1	3	9	5	1	1	6	8	6
Eptser	Chauve-souris	Serotine commune	Eptesicus serotinus	1	3	16	12	41	28	35	41	29
Hypsav	Chauve-souris	Vespere de Savi	Hypsugo savii	1	3	295	124	22	16	106	65	85
Minsch	Chauve-souris	Minioptere	Miniopterus schreibersii	1	3	3	6	8	5	5	2	1
Myocap	Chauve-souris	Murin de Capaccini	Myotis capaccinii	77	3	0	0	1	0	0	0	0
Myodau	Chauve-souris	Murin de Daubenton	Myotis daubentonii	84	3	1	1	0	0	1	0	1
Myoema	Chauve-souris	Murin a oreilles echancrees	Myotis emarginatus	1	3	7	4	0	0	1	0	0

Acoustic activity as a measure of relative abundance

Interpreting data locally



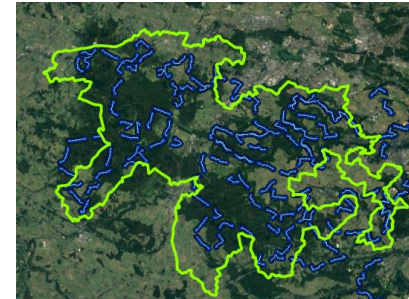
Nb of call
sequences /
20 km
transect



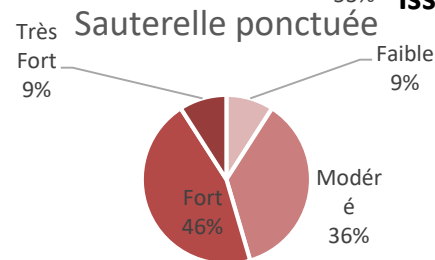
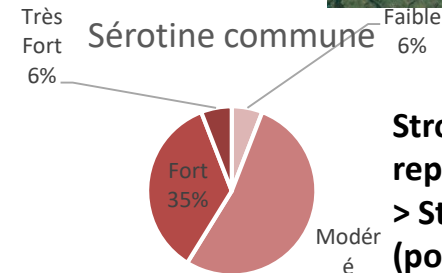
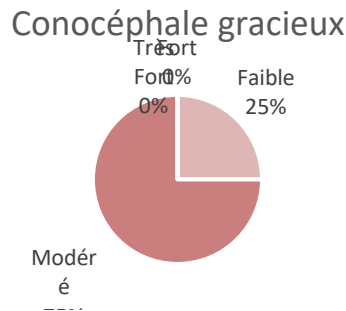
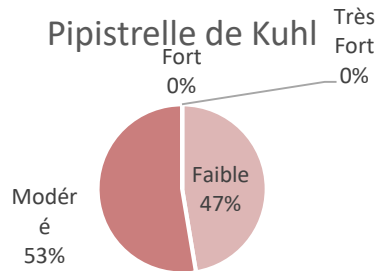
Acoustic activity as a measure of relative abundance

Exemple :

2 espèces dont l'activité est toujours faible à modéré :



Quantile distribution on a whole site



Strong activity over-represented
> Strong responsibility (potential conservation issue)



Acoustic activity as a measure of relative abundance

Robust method: applicable to rare species (little
data necessary)

RNCFS Caroux-Espinouse



Barbastelle



Ehippigrè
carénée



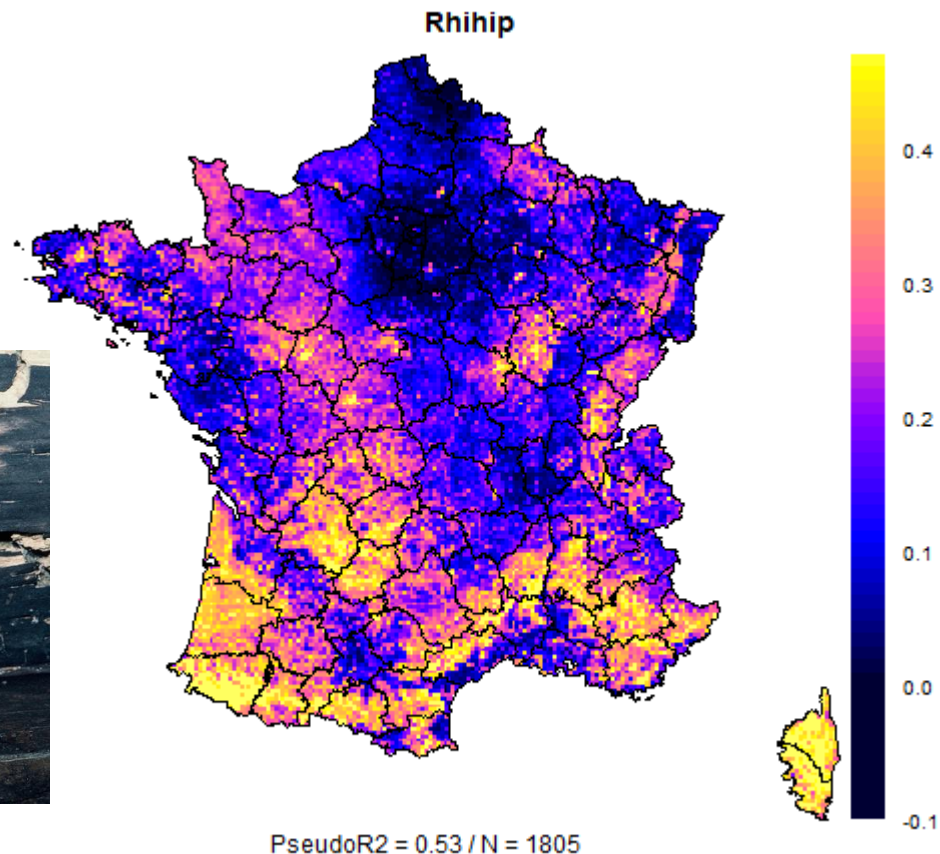
Oreillard gris



Using acoustic activity to map species relative abundance

Identifying stronghold populations

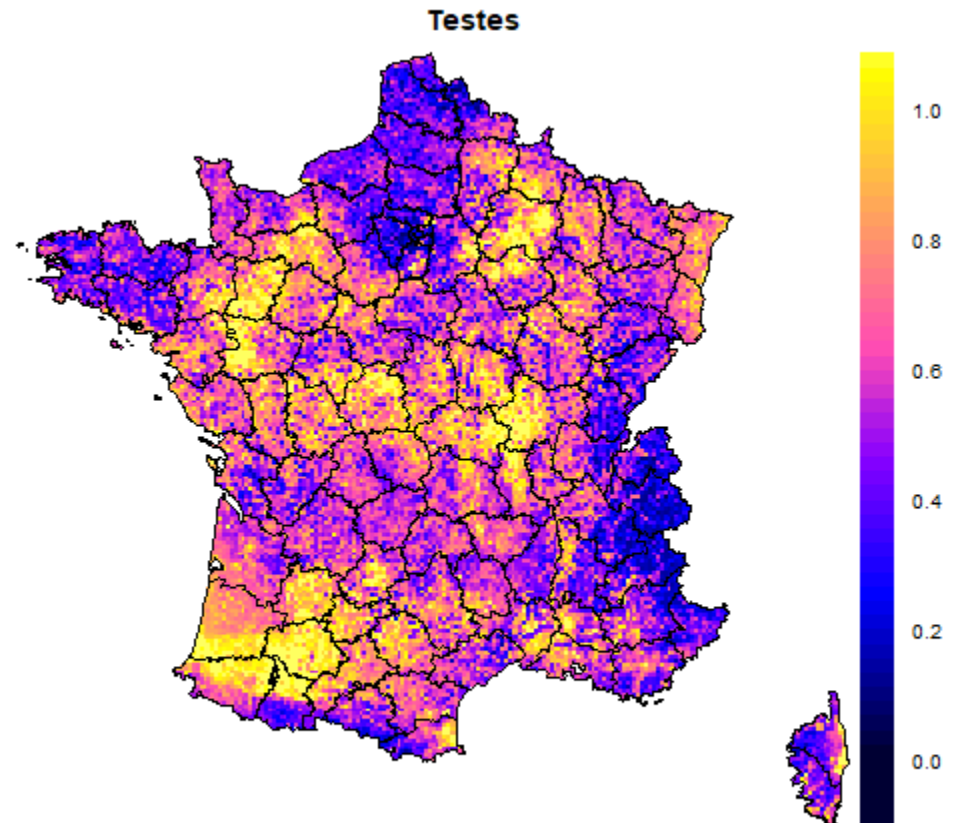
Lesser
Horseshoe Bat



Using acoustic activity to map species relative abundance

Identifying stronghold populations

Slender Bush-Cricket

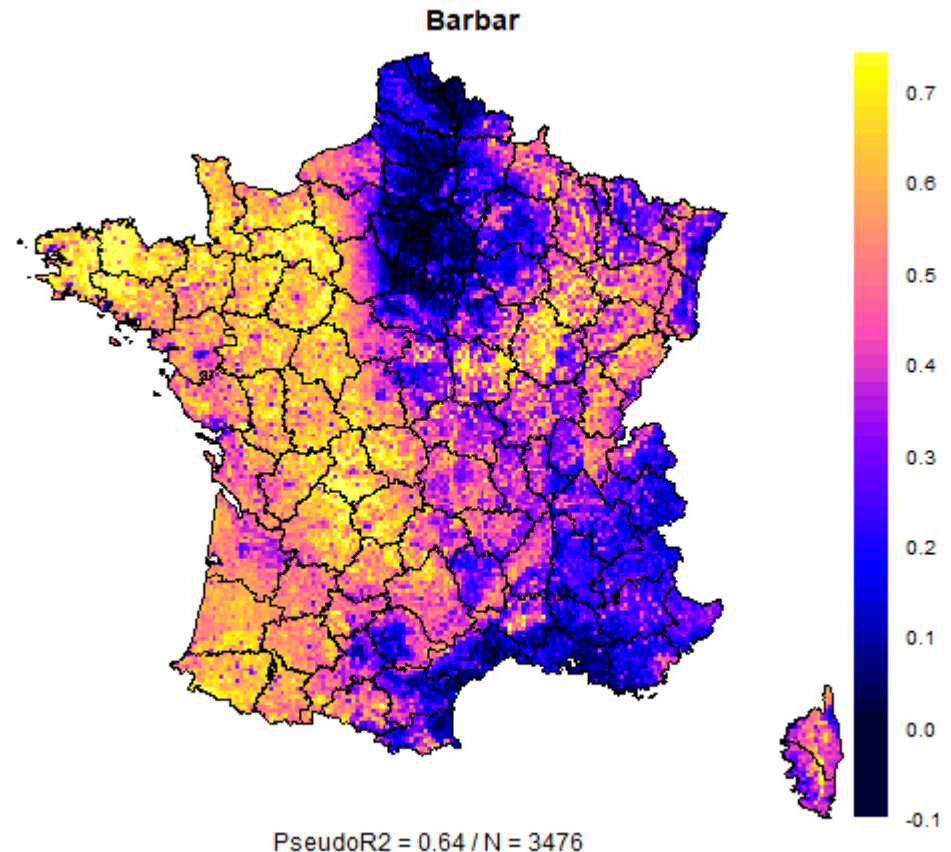


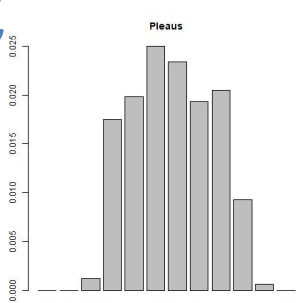
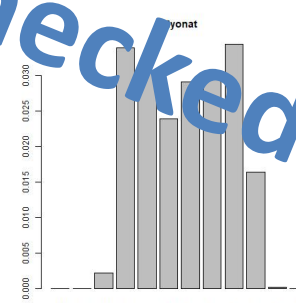
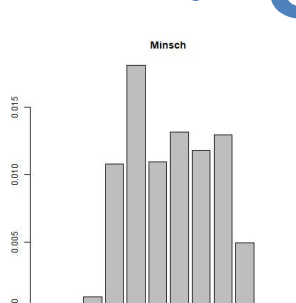
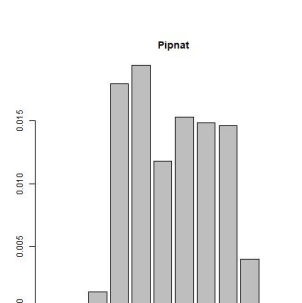
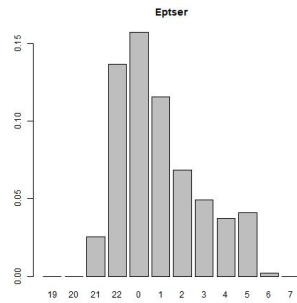
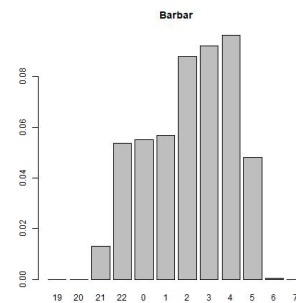
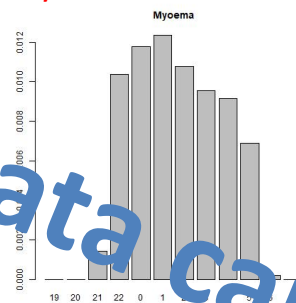
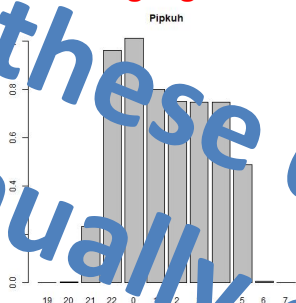
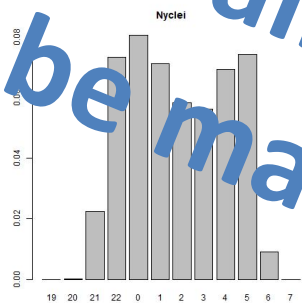
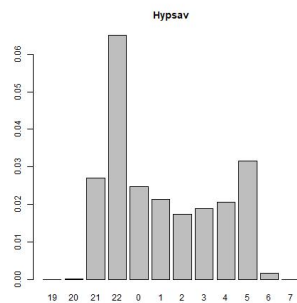
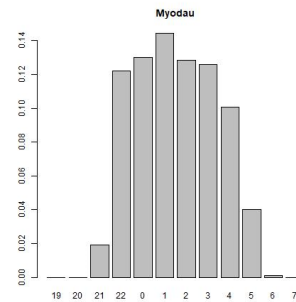
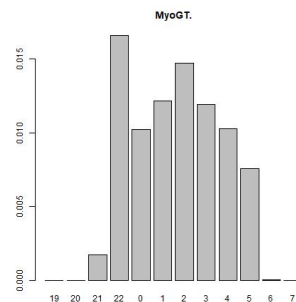
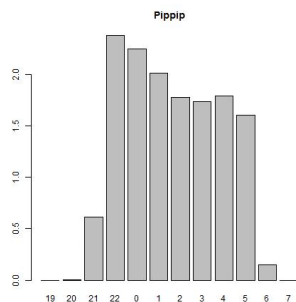
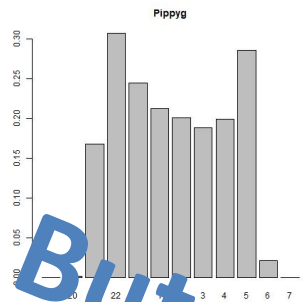
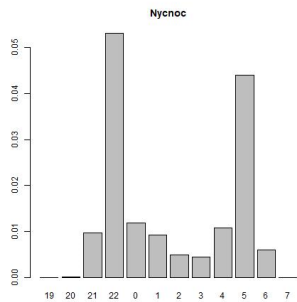
PseudoR2 = 0.5 / N = 2744

Using acoustic activity to map species relative abundance

Identifying stronghold populations

Barbastelle
Bat





But all these data can't
be manually checked

Benchmarking night-time activity



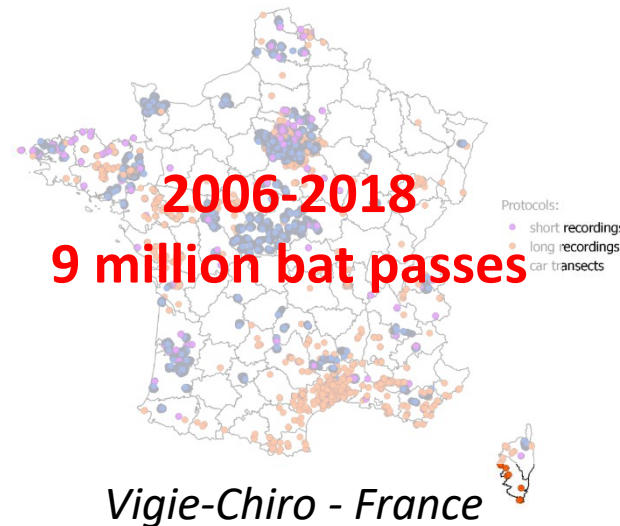
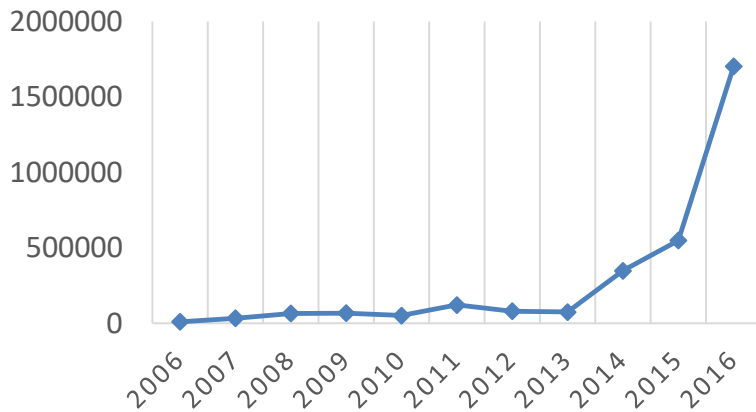
Vigie-Chiro

- 1) Introduction aux chiroptères (chauves-souris) et leur étude par l'acoustique
- 2) Les protocoles standardisés et participatifs
- 3) L'activité acoustique comme mesure d'abondance relative
- 4) L'identification automatique**
- 5) La gestion des erreurs et la qualification des données

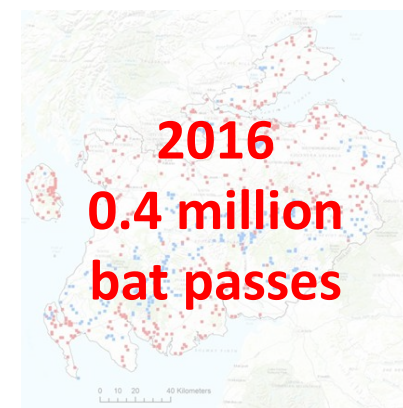
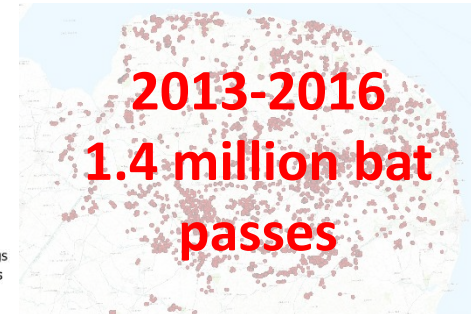
Why using automatic id?

- Reduced costs => exponential data increase

BAT PASSES RECORDED / YEAR (FRANCE)



Norfolk BS (UK)



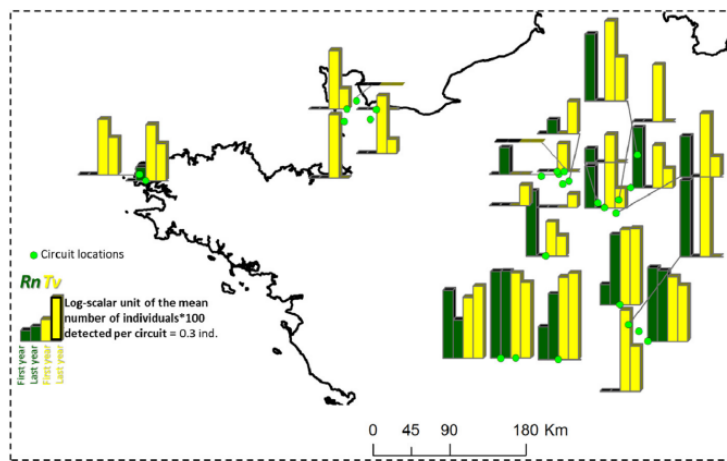
South Scotland BS (UK)

- Complete manual checking impossible...

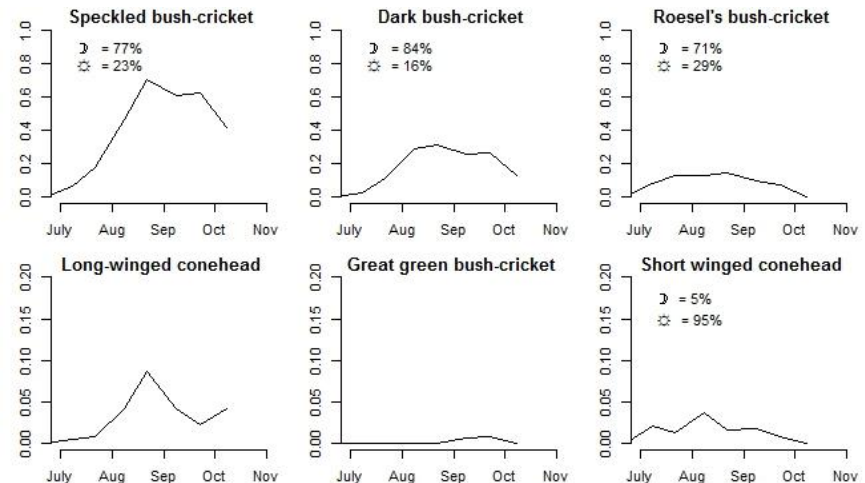
There is just no other way than auto id!

Why using automatic id?

- Other less known good reasons:
 - 1) Manual checking error rate is decreasing over time! but biasing trends estimates... Solution: machines can easily re-analyse historic data and control observer bias
 - 2) You can get very good data on non-targeted taxa such as bush-cricket: spatial and temporal patterns, trends!!



Jeliaskov et al. (2016) GECCO



Newson et al. (2017) MEE

Auto id developments

Appeared recently (2012)

- **Fast development in Europe**

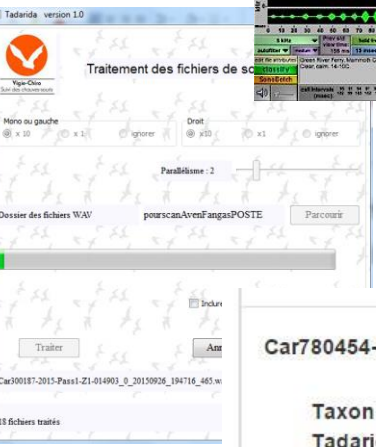
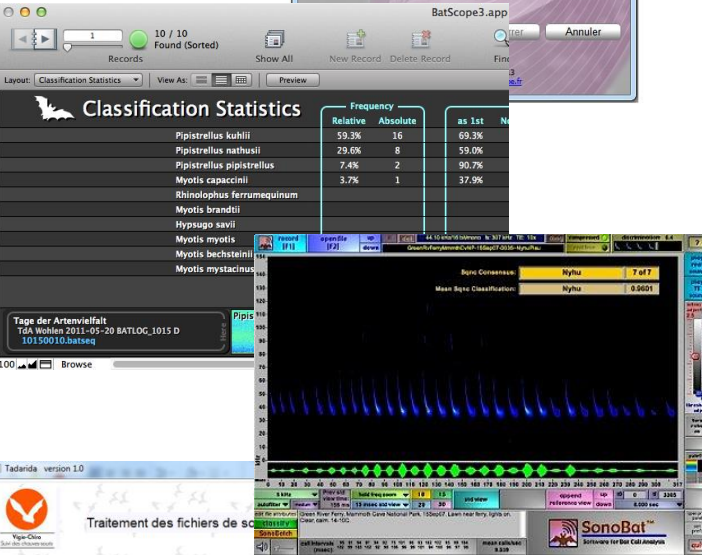
- 7 softwares en Europe, 4 in North America, 3 in Latin America, 1 in South Africa, etc

- Efficiency depends:

- On size of soundreference database
- Recorder type
- The number of species covered (10-114)

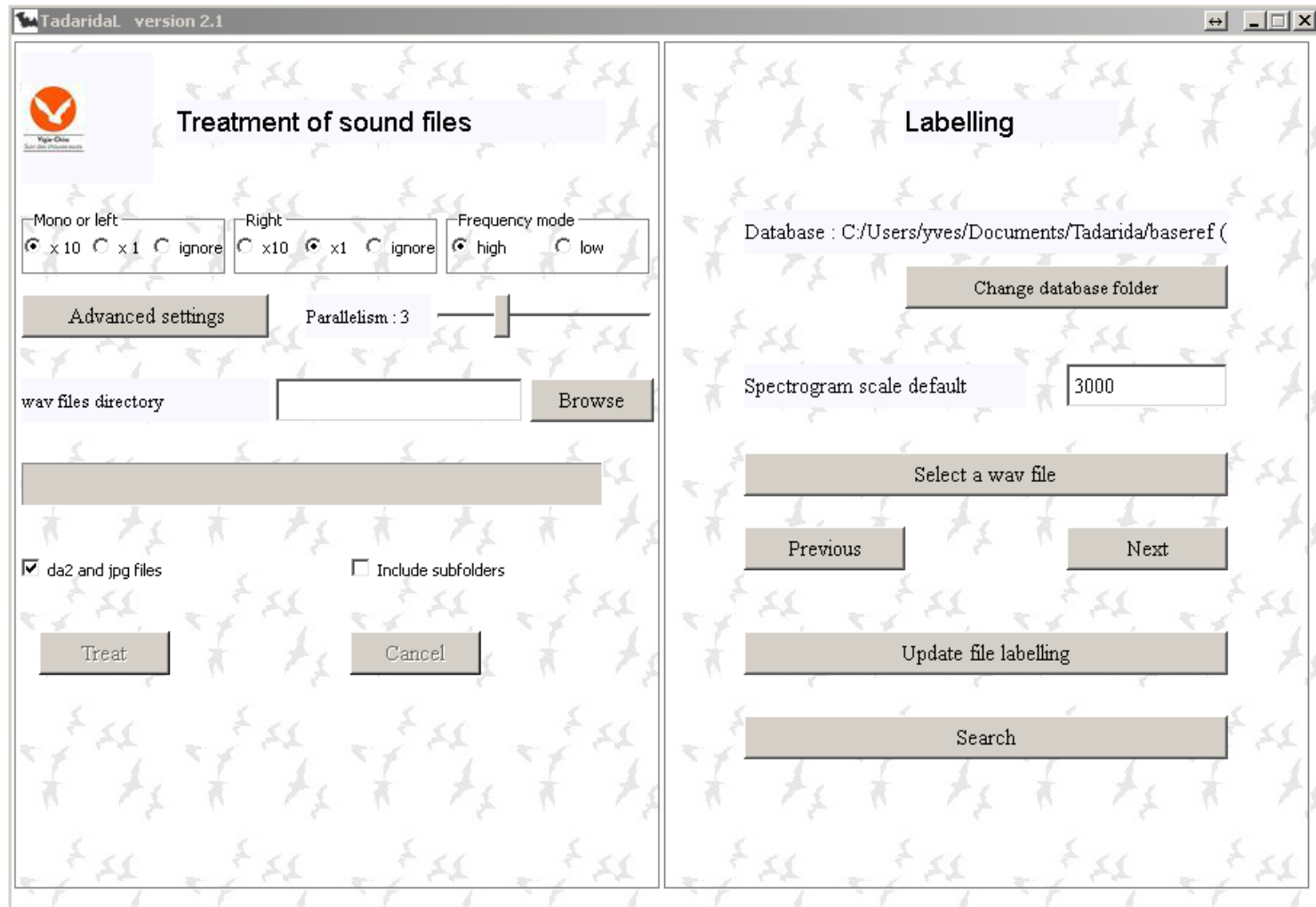
- Principle: **software propose an identification with a confidence score, the user validate a sample...**

- Time gain depends on objectives of analysis



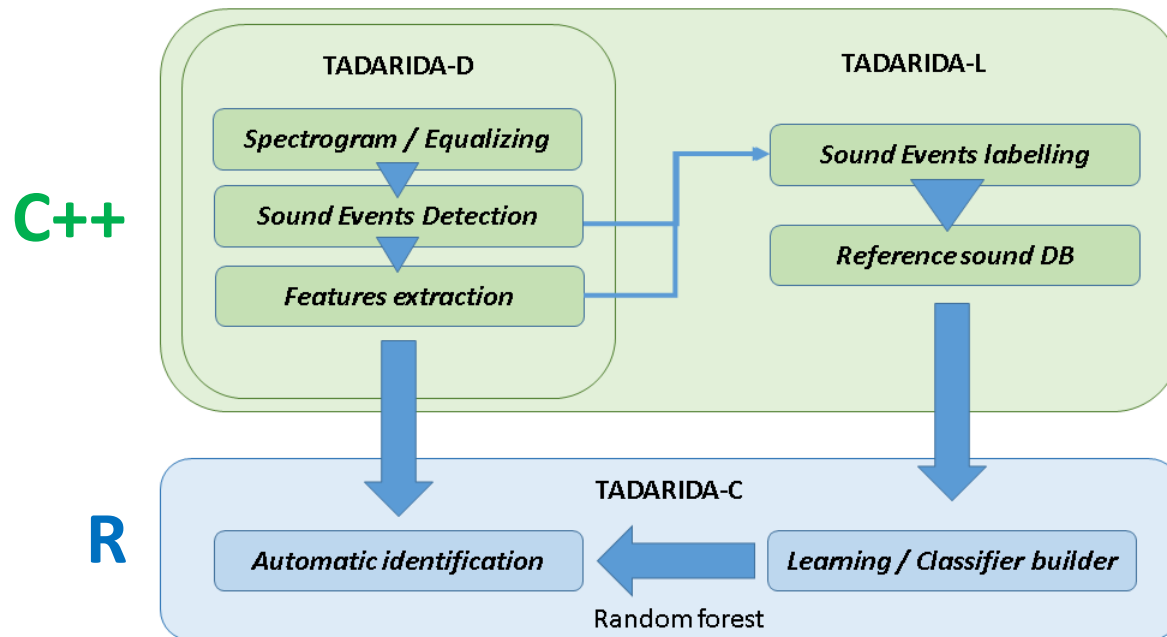
How using auto id?

- The example of Tadarida open software



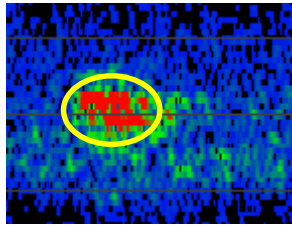
How using auto id?

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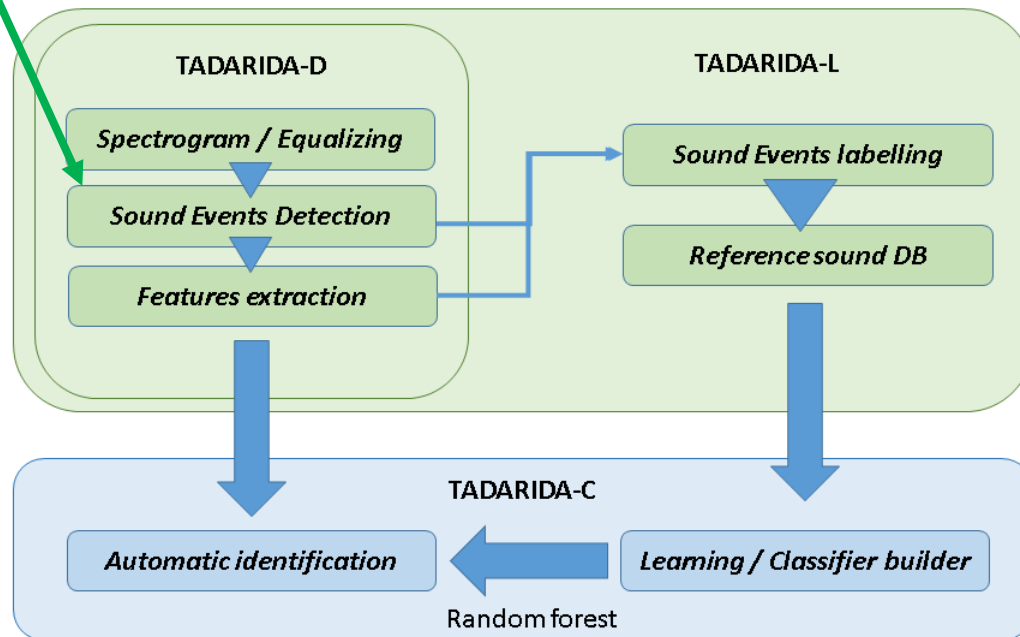
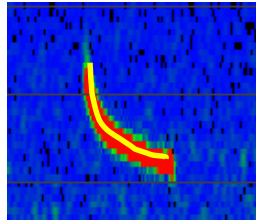


How using auto id?

- The example of Tadarida open software

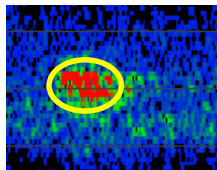


Generic
time/freq.
segmentation

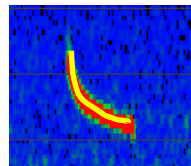


How using auto id?

- The example of Tadarida open software



Generic acoustic event detector



High frequency mode (8-250 kHz)

FFT size : 0.8-1.6 ms

Overlap 75%

➤ Resolution :

➤ 0.2-0.4 ms

➤ 0.6-1.2 kHz

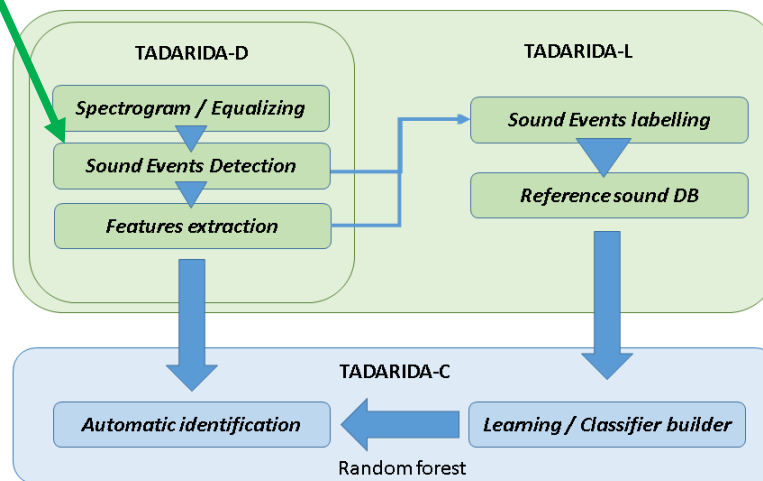
Background noise =

quantile 5% per frequency bands

Detection on start (26 dB SNR) and

stop (20 dB SNR) threshold

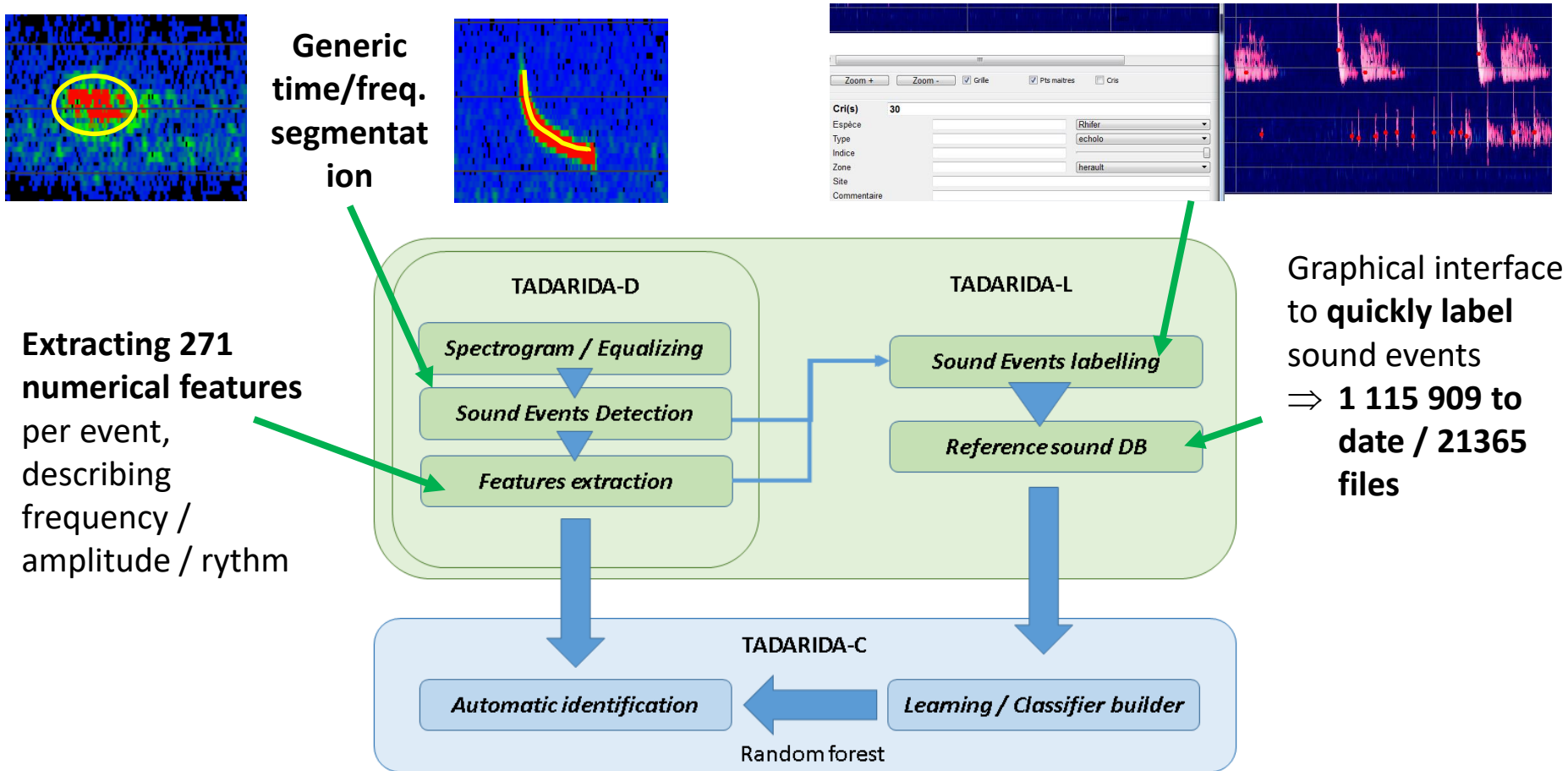
(hysteresis principle)



Bas et al. (2017) Journal of Open Research Software

How using auto id?

- The example of Tadarida open software



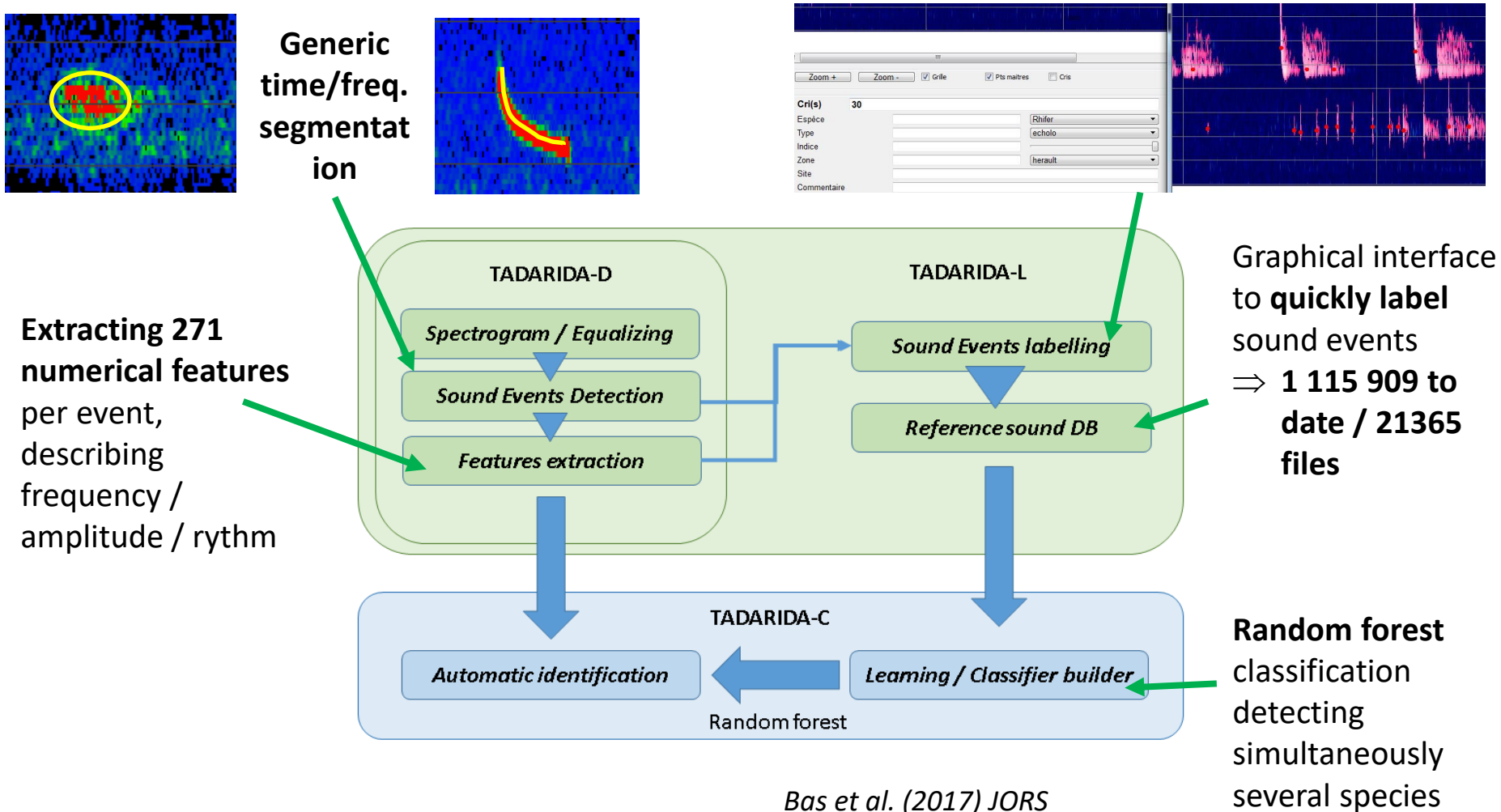
How using auto id?

- The example of Tadarida open software

The screenshot displays the Tadarida - Labelling software interface. The main window shows a spectrogram of sound events with a grid overlay. The y-axis represents frequency in kHz (20, 40, 60, 80, 100) and the x-axis represents time in seconds (1, 2, 3). The spectrogram shows several sound events, with some labeled with green and red dots. The interface includes a toolbar with buttons for Zoom +, Zoom -, Grid, Master pts, and Lines. The status bar shows r=10.08, -51 ms, and 117.00 kHz. The file path is C:/Users/yves/Documents/Tadarida/test/eti. The folder is C:/Users/yves/Documents/Tadarida/test/eti. The file name is Car170517-2014-Pass1-C1-OB-1_20140702_224038_761.eti. The folder contains 19 labels, and the current label is Pippip. The sound event(s) are 3,4,5,7,10,12,14,15,19,21,24,26,27,29,30,33,35,36,37. The metadata form includes fields for Species (Barbar), Type (echolo), Confidence (3), Region (Charente-Maritime), Site (Tonnay-Charente), Recorder (SM2BAT+), Confidential (checkbox), Date, Author (Laurene Trebuca), and Labeller (Yves Bas). The interface also has buttons for Validate the label(s), Save the labels file, and Close.

How using auto id?

- The example of Tadarida open software



How using auto id?

- Tadarida-C

- 3 classification layers:

- 1) Classification per calls

- Random forest with bootstrap of train samples stratified per sites

- Correcting unevenness of sample per classes/species (trade-off quantity / evenness)

- + Ad hoc Algorithm clustering clustering calls in sequences

- 2) Classification of sequences (= different species or not)

- Random forest on train files from which species list are exhaustives

- 3) Error correction according to context

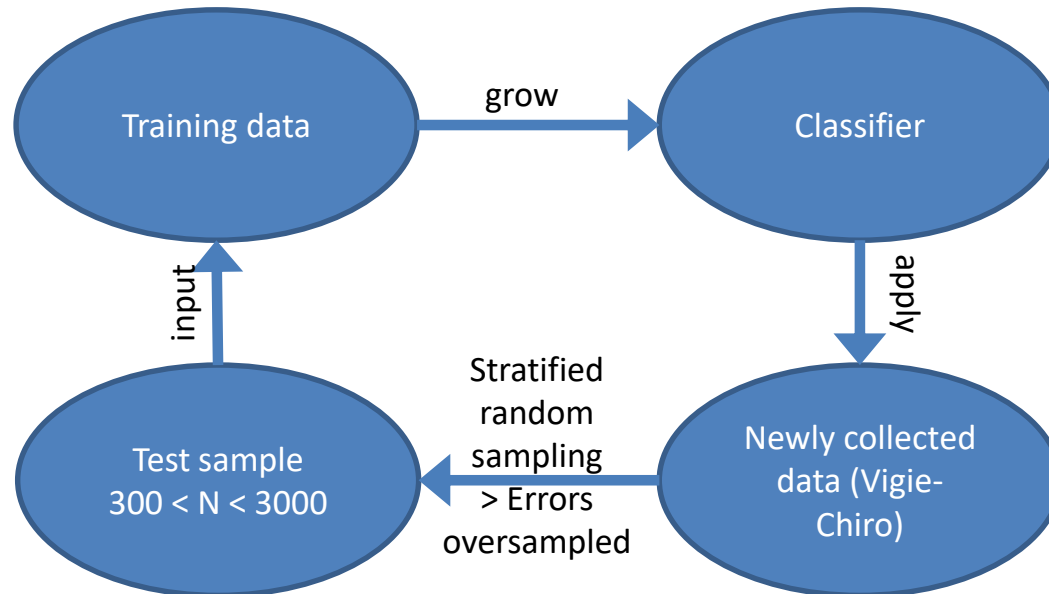
- Random forest of sequences according to whole-night sampling information (scores of other occurrences, ratio of species occurrences)

AUC = 0.885

AUC = 0.967

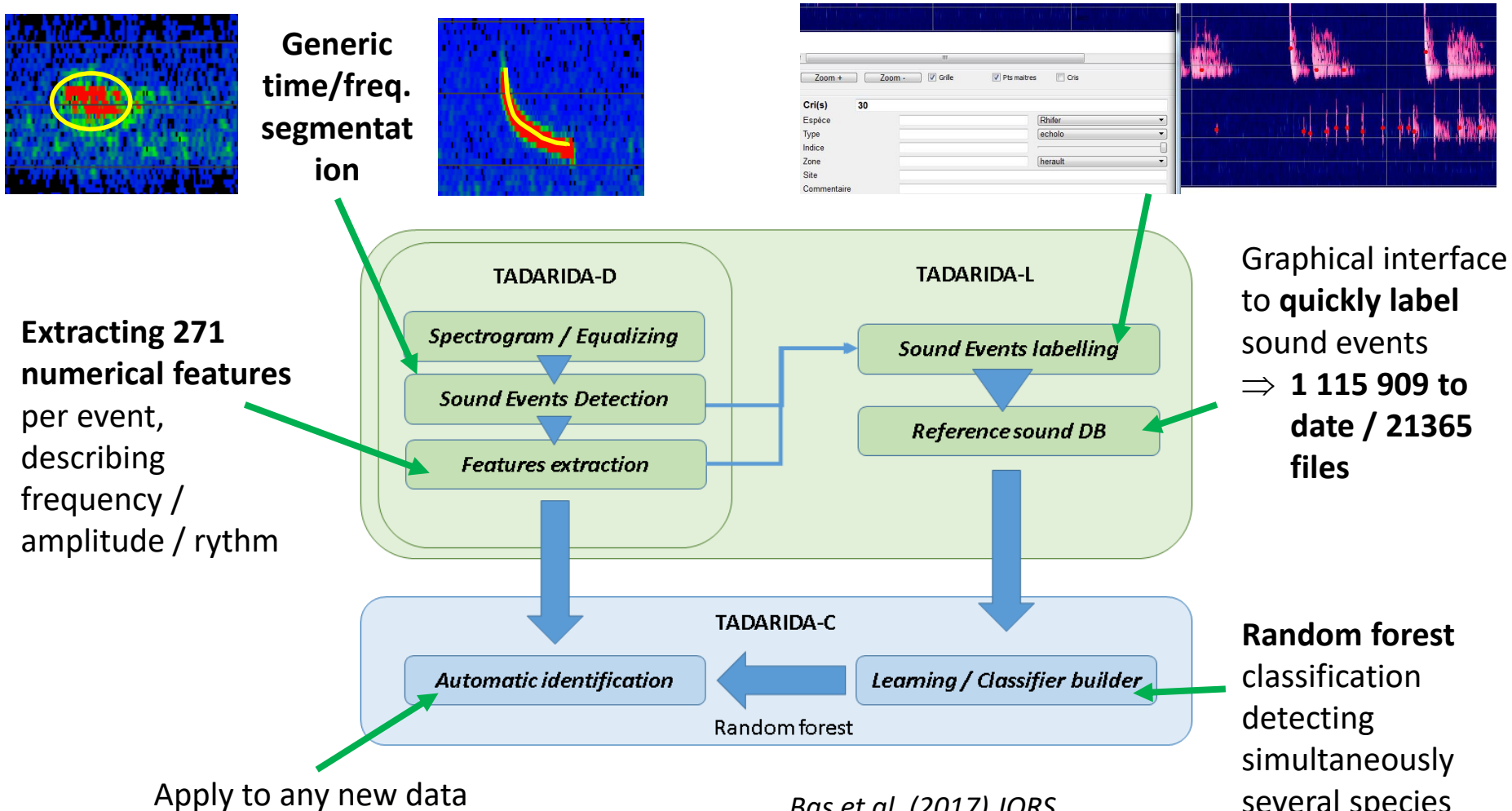
How using auto id?

- Tadarida-C
 - Training « round » :



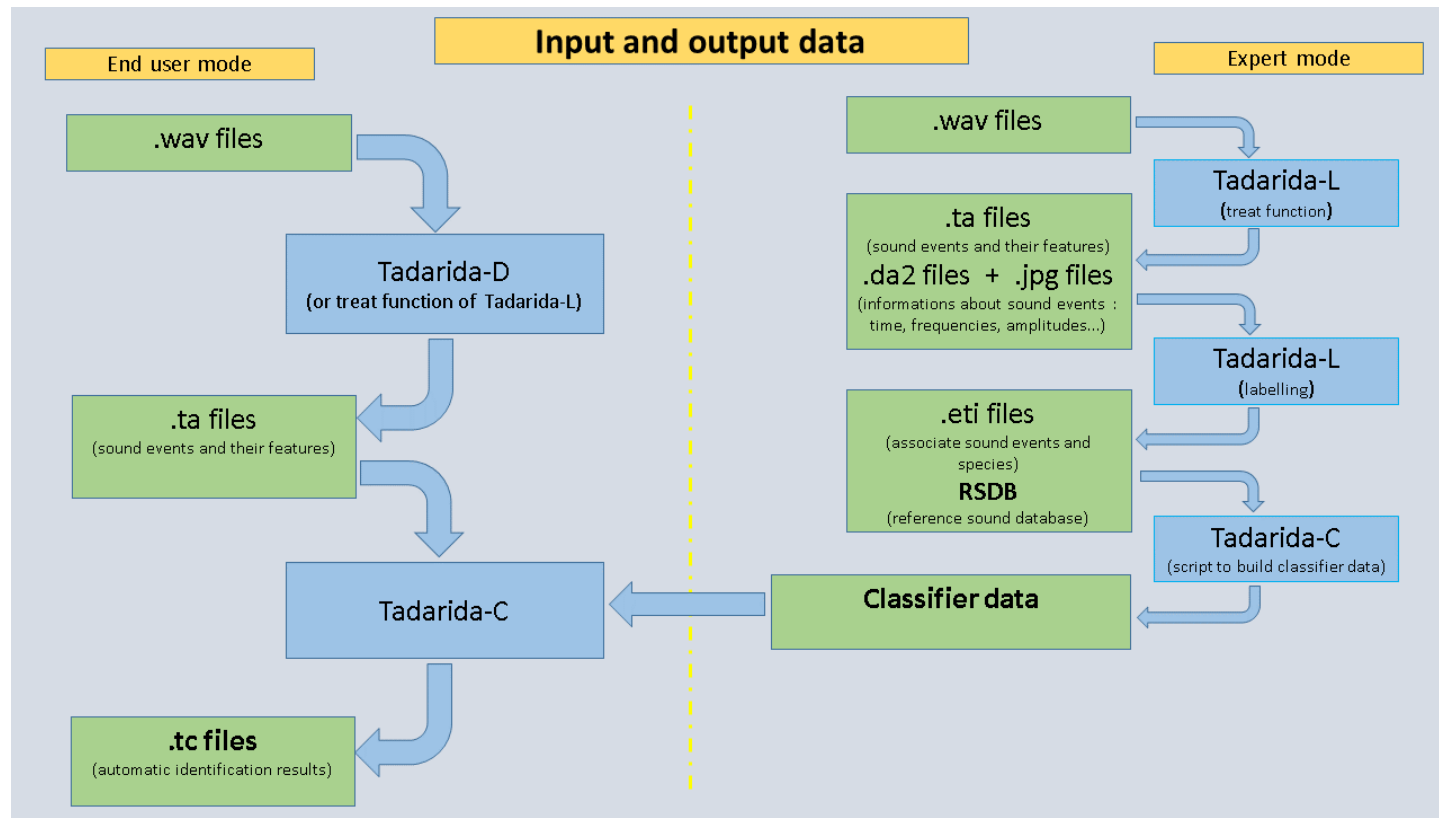
How using auto id?

- The example of Tadarida open software



How using auto id?

The workflow



Improvement perspectives

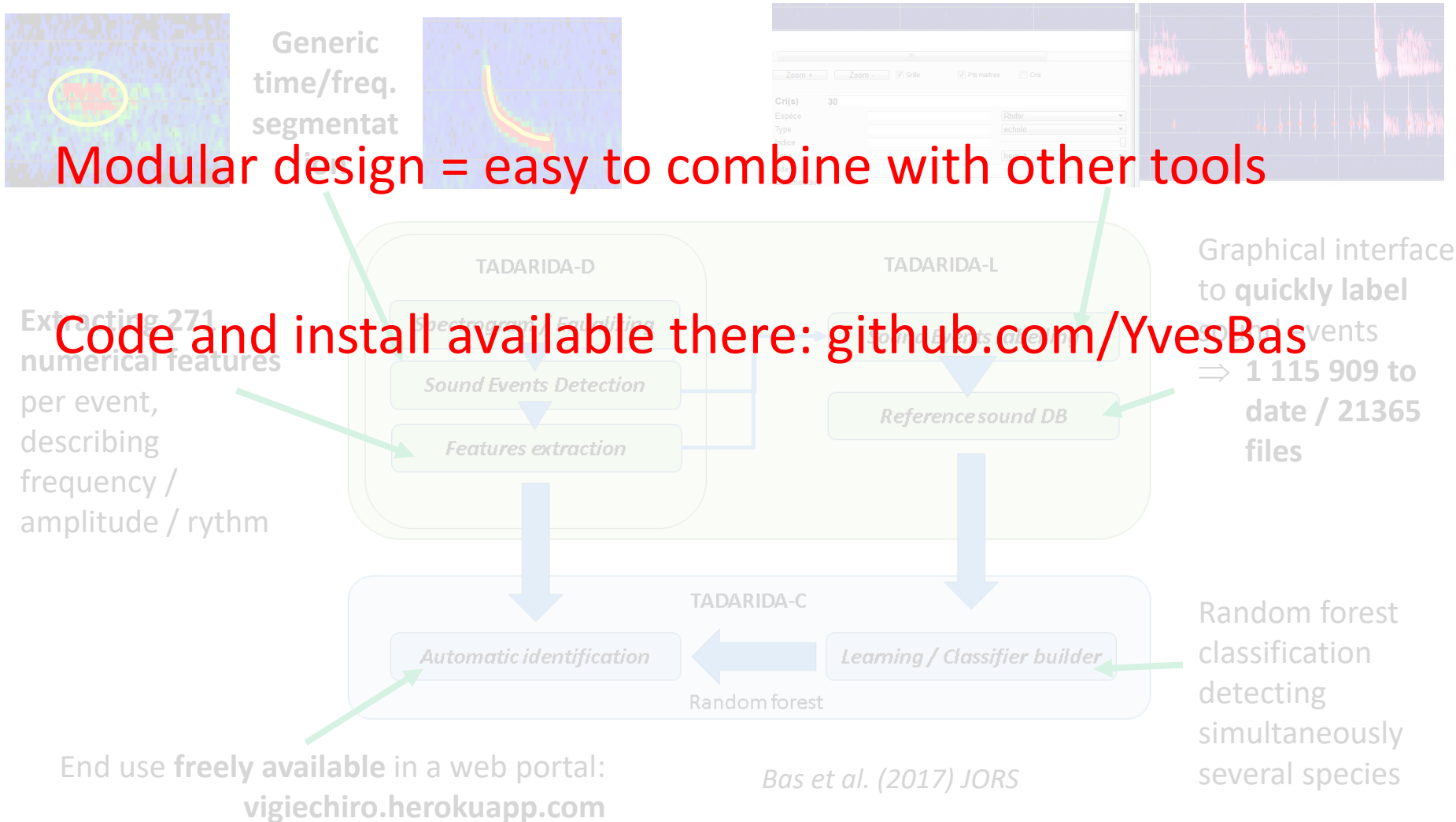
- Segmentation / detection :
 - Use of amplitude distribution within sound events to distinguish close syllables (bird songs)
 - Varying FFT size for narrowband vocalisation (ground-cricket)
 - More features accounting for « echo effects »

Improvement perspectives

- Classification :
 - New methods: deep learning winning BirdClef challenges starting 2016.
 - Improve sound references:
 - Quantity : collaborative > integrating Tadarida-L in a web site
 - Quality: minimising error risk in reference by recording close to known roosts
 - More predictors (acoustic features, context features)
 - Hierarchical classifiers (more species, behaviours)

How using auto id?

- The example of Tadarida open software



How using auto id?

- The example of Tadarida open software

End use available through a web portal: vigiechiro.herokuapp.com



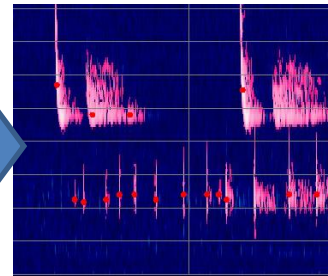
Data collection



Online data transfer



Automatic secure storage (iRODS - IN2P3)



Data processing (Tadarida)

Bilan de la participation

#	Taxon parent	Nom	Nb contact min	Nb de contact
1	Chiroptères	Oreillard gris (<i>Plecotus austriacus</i>)	0	13
2	Chiroptères	Minioptère de Schreibers (<i>Miniopterus schreibersii</i>)	1	2
3	Chiroptères	Pipistrelle soprane (<i>Pipistrellus pygmaeus</i>)	3252	3334
4	Chiroptères	Noctule de Leisler (<i>Nyctalus leisleri</i>)	43	110
5	Chiroptères	Murin de Daubenton (<i>Myotis daubentonii</i>)	0	11
6	Chiroptères	Murin à oreilles échancrées (<i>Myotis emarginatus</i>)	0	1
7	Chiroptères	Pipistrelle de Kuhl (<i>Pipistrellus kuhlii</i>)	642	702
8	Chiroptères	Sérotine commune (<i>Eptesicus serotinus</i>)	0	28
9	Chiroptères	Murin de Capaccini (<i>Myotis capaccinii</i>)	0	1
10	Chiroptères	Pipistrelle de Nathusius (<i>Pipistrellus nathusii</i>)	3	5

Automatic feedback



Car340527-2015-Pass1-Z1-9388_0_20150606_012004_460

Taxon	Confiance	Taxon observateur	Confiance observateur	Taxon validateur	Confiance validateur	FreqM	TDeb	TFl
Pipistrelle soprane (<i>Pipistrellus pygmaeus</i>)	0.994	Pipistrelle soprane (<i>Pipistrellus pygmaeus</i>)	OK	OK	OK	54	0	3.4
Pipistrelle commune (<i>Pipistrellus pipistrellus</i>)	0.996	Pipistrelle commune (<i>Pipistrellus pipistrellus</i>)	OK	OK	OK	45	0.9	2.6
Oreillard gris (<i>Plecotus sp.</i>)	0.65	bruit	OK	OK	OK	8	0	0.3

Online manual checking

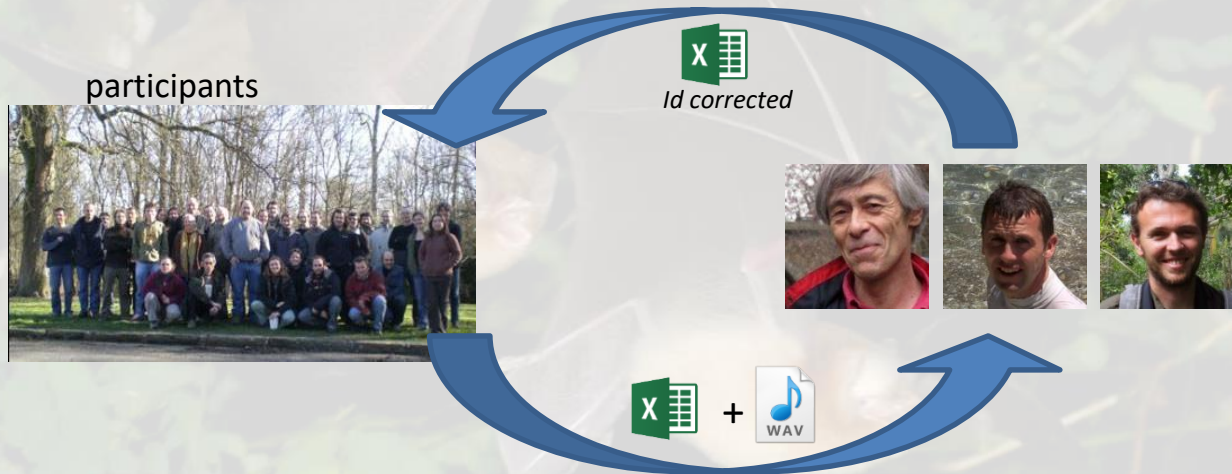


<https://github.com/Scille/vigiechiro-front>

<https://github.com/Scille/vigiechiro-api>

Back to history (2006-2010)

- A virtuous cycle



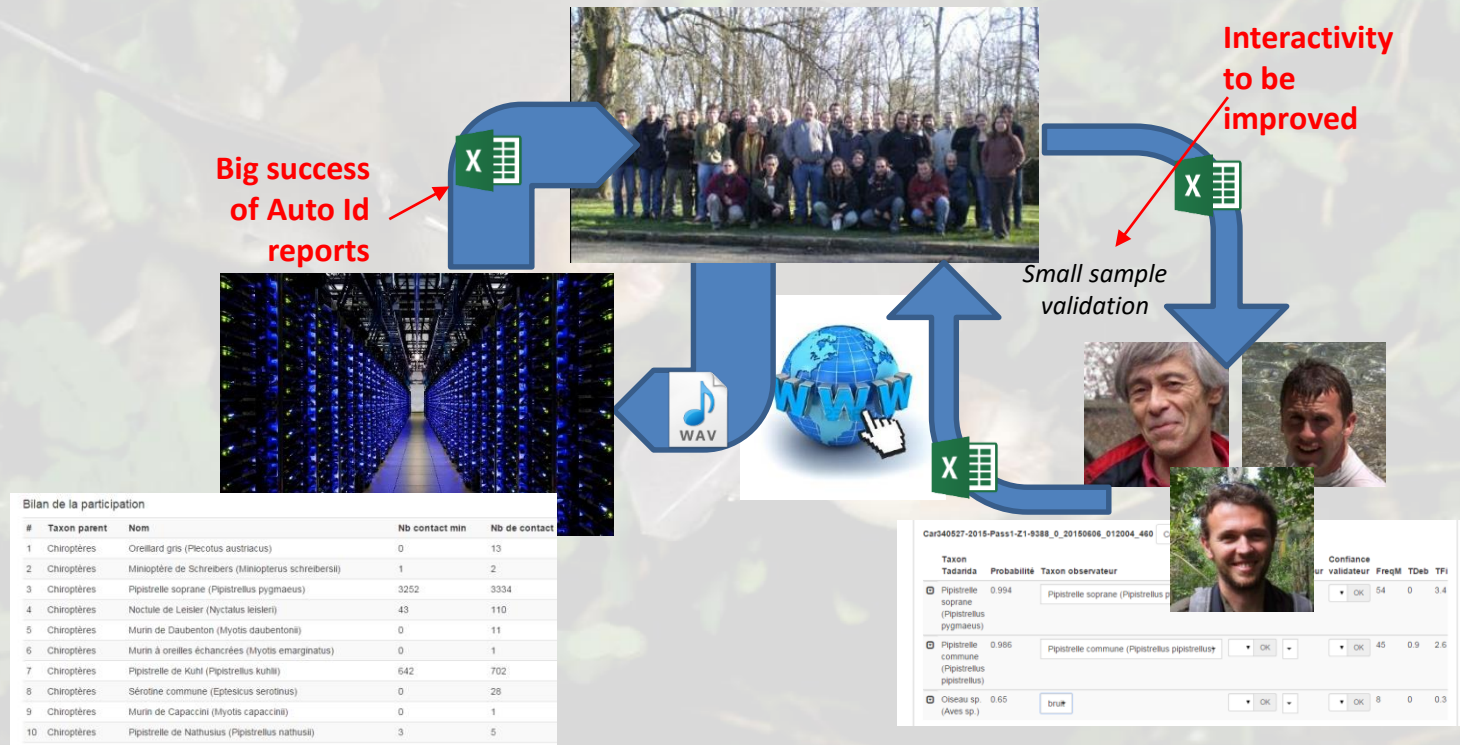
2011-2014: hard to follow...

- Increasing participation slowed the cycle...



2015-2016: Web portal and Tadarida implementation

- Towards a quicker cycle



Vigie-Chiro

- 1) Introduction aux chiroptères (chauves-souris) et leur étude par l'acoustique
- 2) Les protocoles standardisés et participatifs
- 3) L'activité acoustique comme mesure d'abondance relative
- 4) L'identification automatique
- 5) La gestion des erreurs et la qualification des données

How's automatic acoustic monitoring doing?

- Norfolk Bat Survey

Table 2
Results showing the process involved in analysing and validating acoustic bat data collected through this project, and the number of recordings

Identity	Step 1. Initial analysis	Step 2. Secondary recoding		Step 3. Manual analysis of recordings: re	
	Initially assigned to species	Recordings removed (confidence index < 3 and/or < 3 calls)	Confidence index at end of Step 2 (median, range)	Recordings manually checked	% unchanged
Mdau	5403	3635	3 (3–6)	1768	90%
Mmys/Mbra	3471	1950	3 (3–6)	1521	28%
Mnat	1793	491	4 (3–10)	1302	85%
Nnoc	8032	785	8 (3–10)	7247	88%
Nlei	673	357	5 (3–10)	316	87%
Eser	4053	1623	4 (3–9)	2430	99%
Ppip	338,260	8969	10 (3–10)	1000 (sample)	99%
Ppyg	179,482	6826	10 (3–10)	1000 (sample)	99%
Pnat	1740	733	5 (3–10)	1007	91%
Paur	4224	2264	5 (3–10)	1960	99%
Bbar	2732	744	8 (3–10)	1988	100%

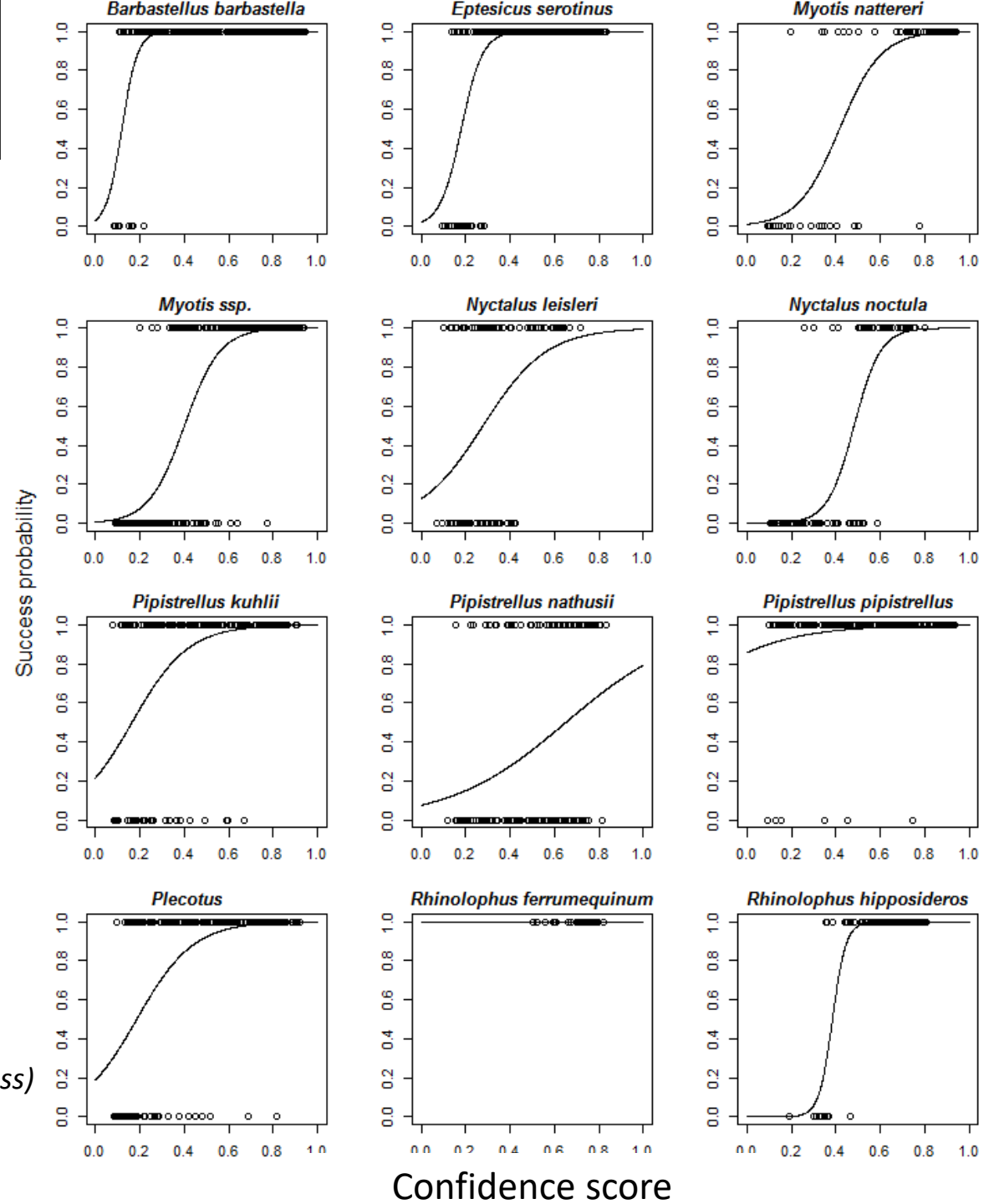
Low error rate for many species

Auto id: Score reliability

- Correlate error risk / confidence score
 - identifying selection thresholds

Confirmed id ~ software confidence

(Barré et al. 2019 MEE, in press)



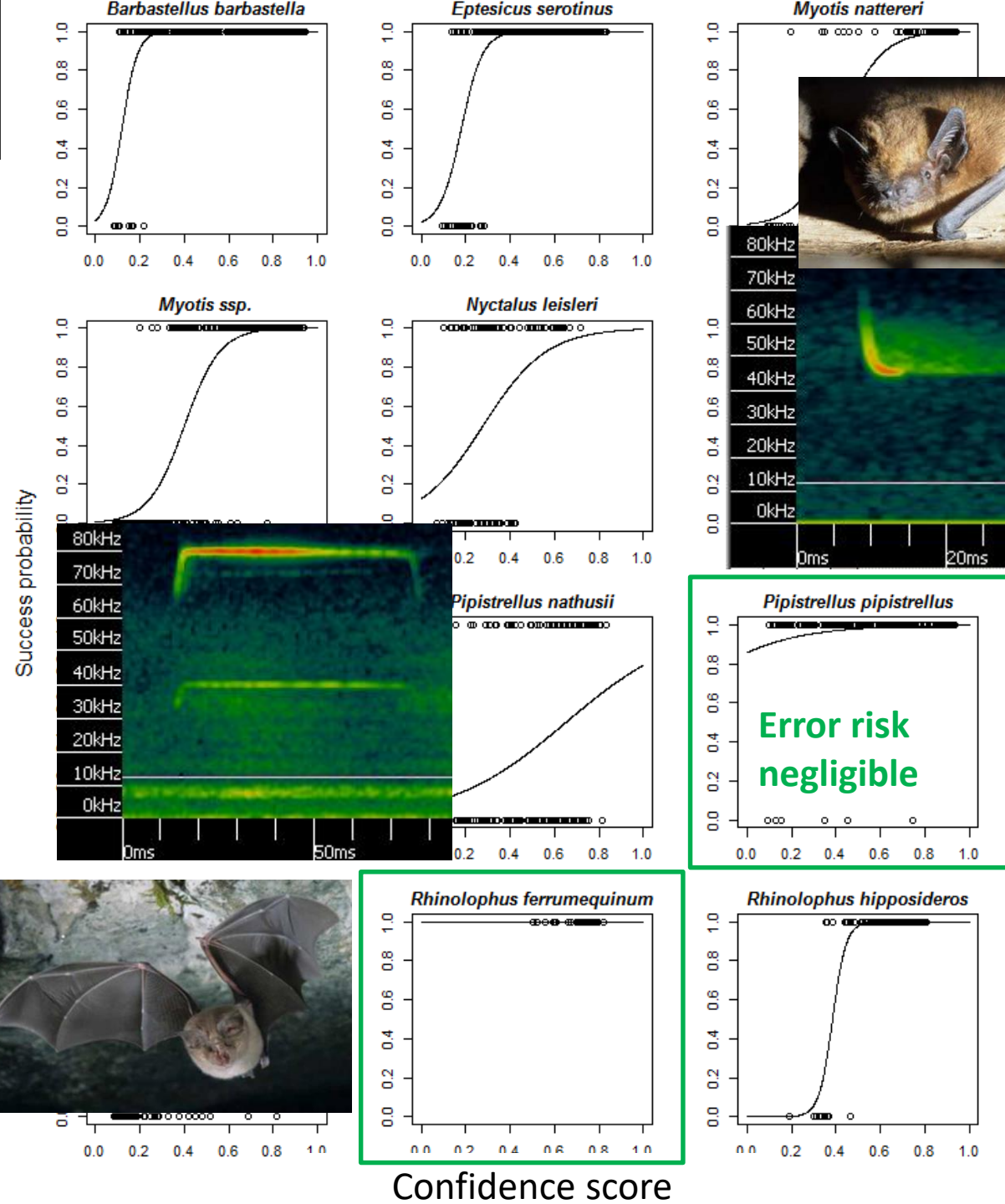
Confidence score

Error modelling

- Correlate error risk / confidence score
 - identifying selection thresholds

Confirmed id ~ software confidence

(Barré et al. 2019 MEE, in p)



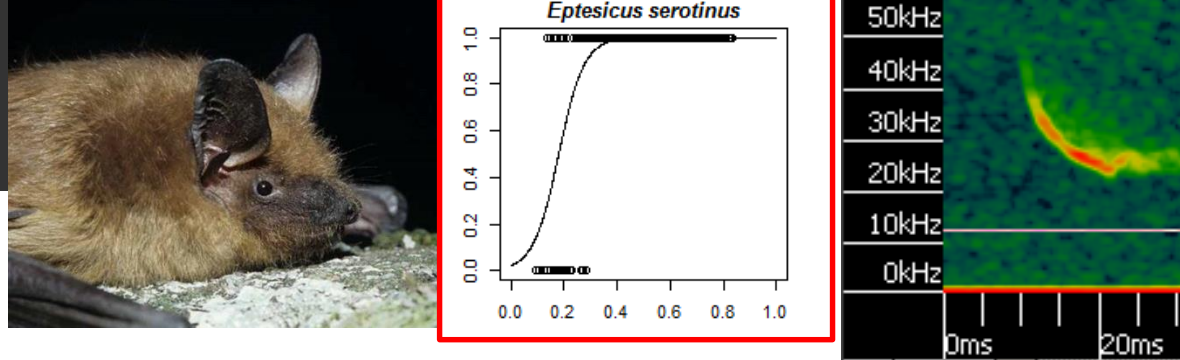
Confidence score

Error modelling

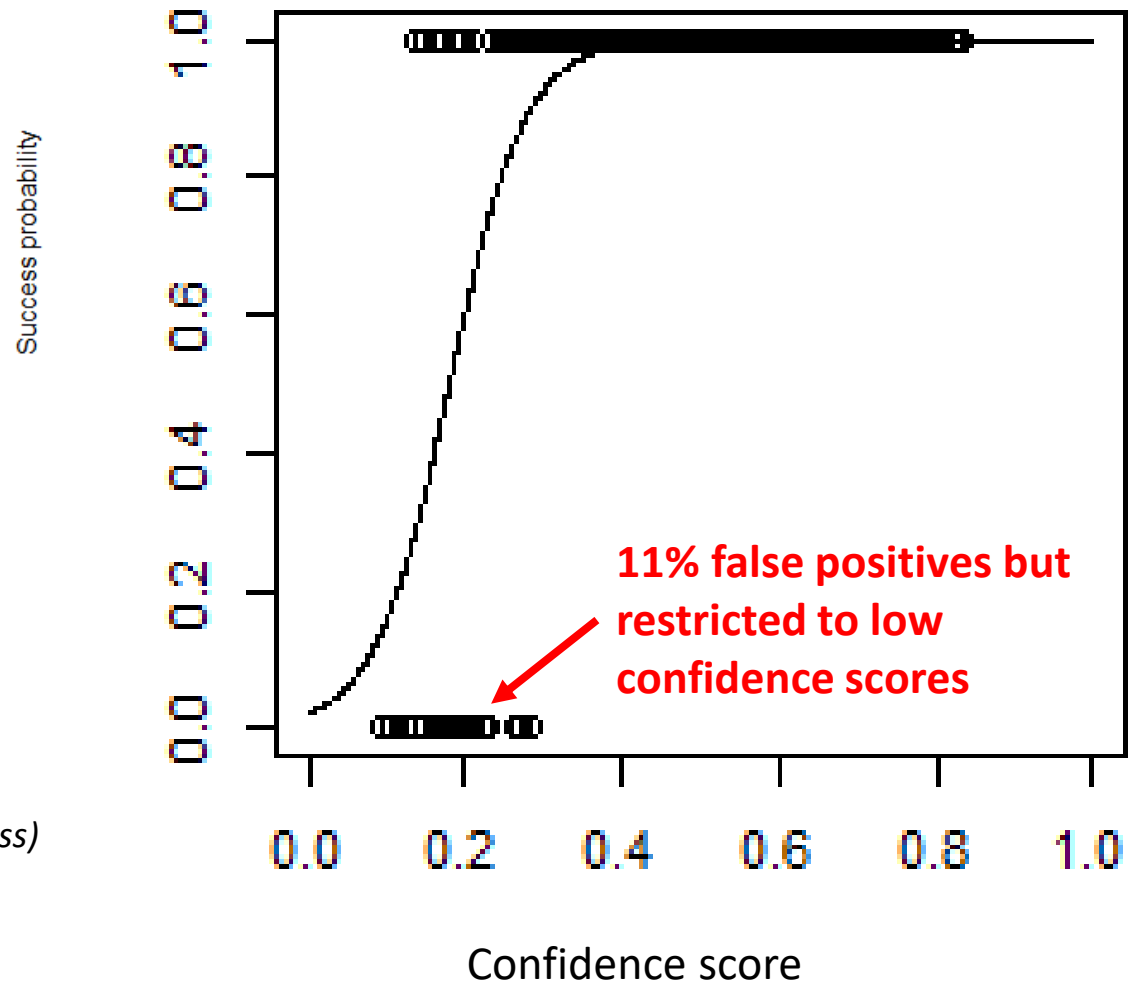
- Correlate error risk / confidence score
 - identifying selection thresholds

Confirmed id ~ software confidence

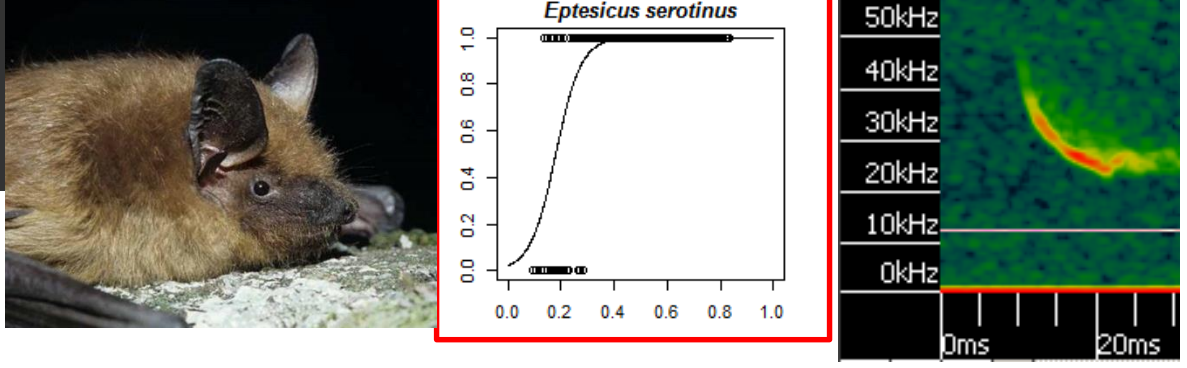
(Barré et al. 2019 MEE, in press)



Eptesicus serotinus



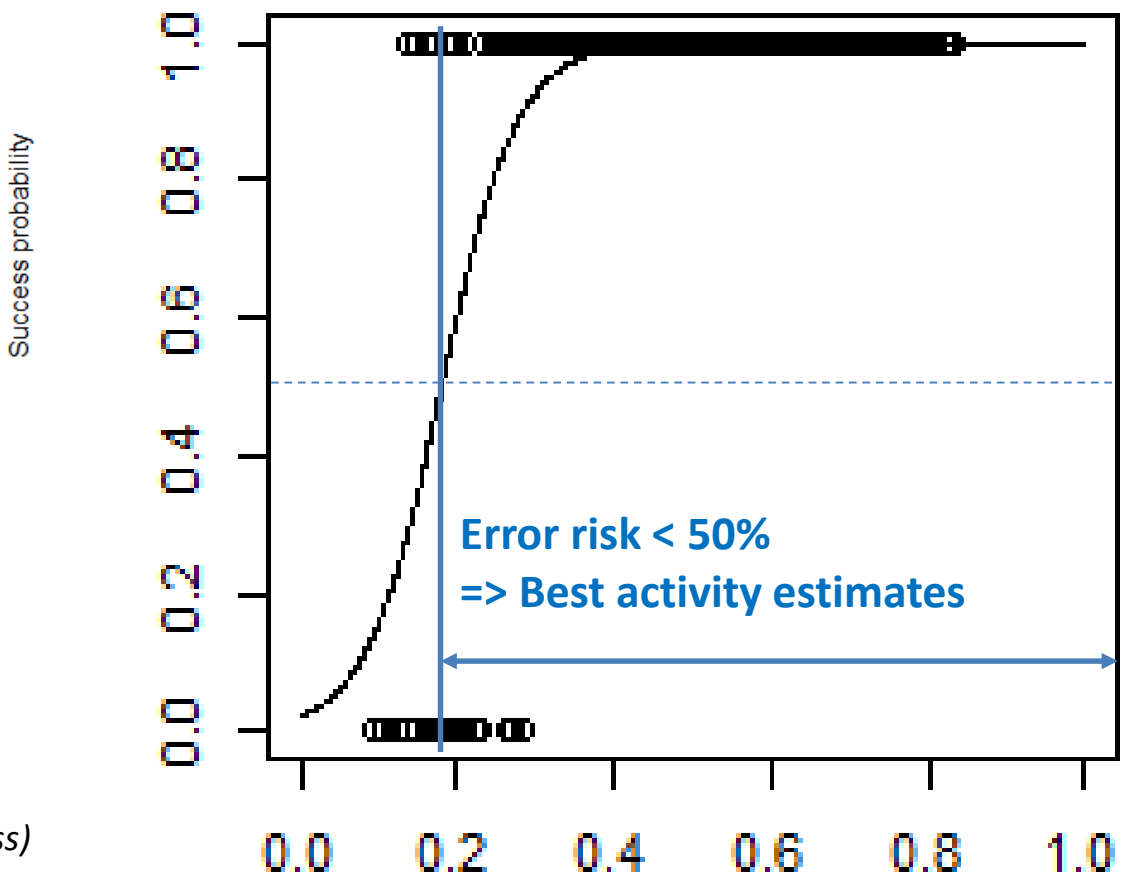
Error modelling



- Correlate error risk / confidence score
 - identifying selection thresholds

Confirmed id ~ software confidence

Eptesicus serotinus



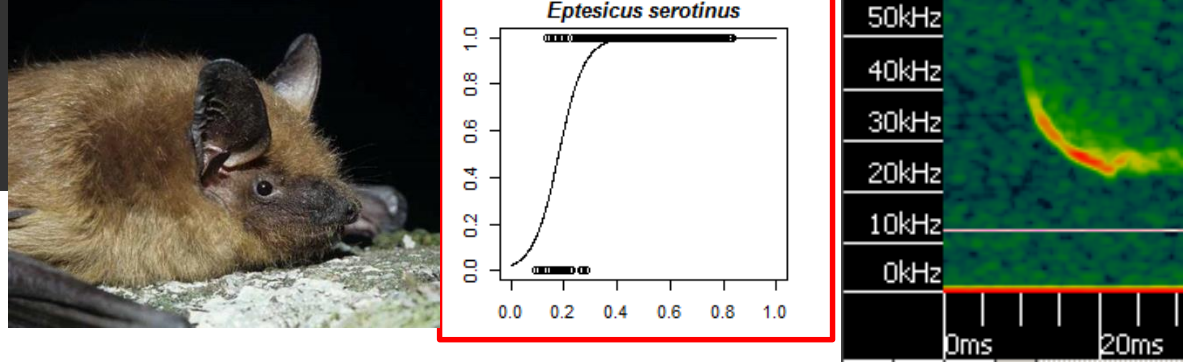
(Barré et al. 2019 MEE, in press)

Error modelling

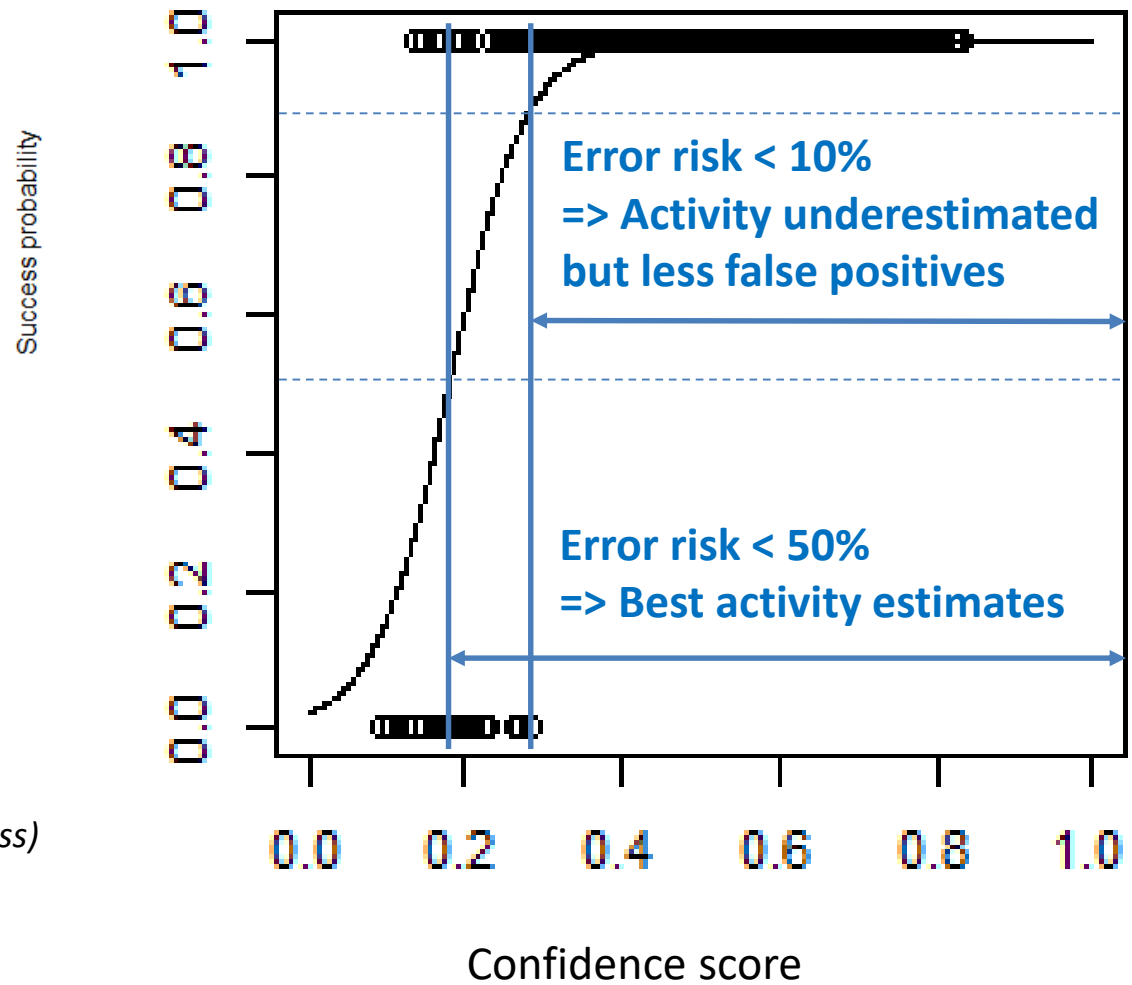
- Correlate error risk / confidence score
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(Barré et al. 2019 MEE, in press)



Eptesicus serotinus



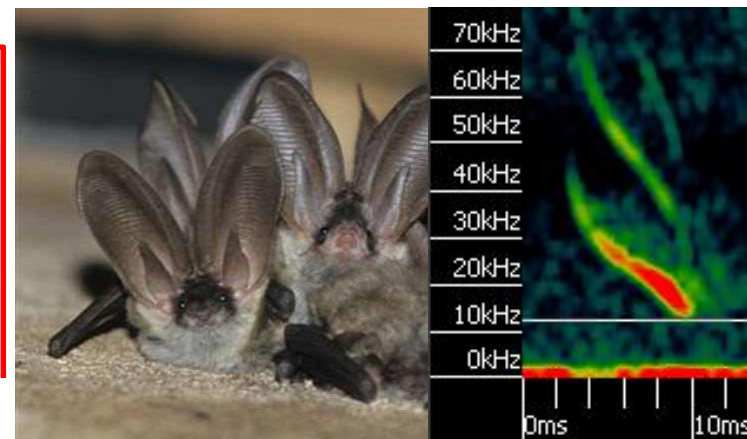
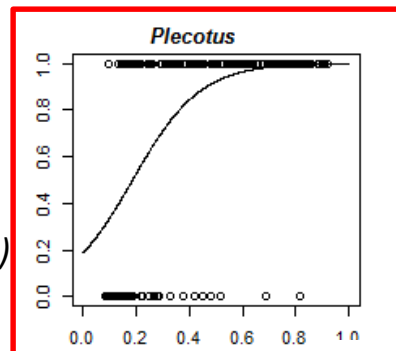
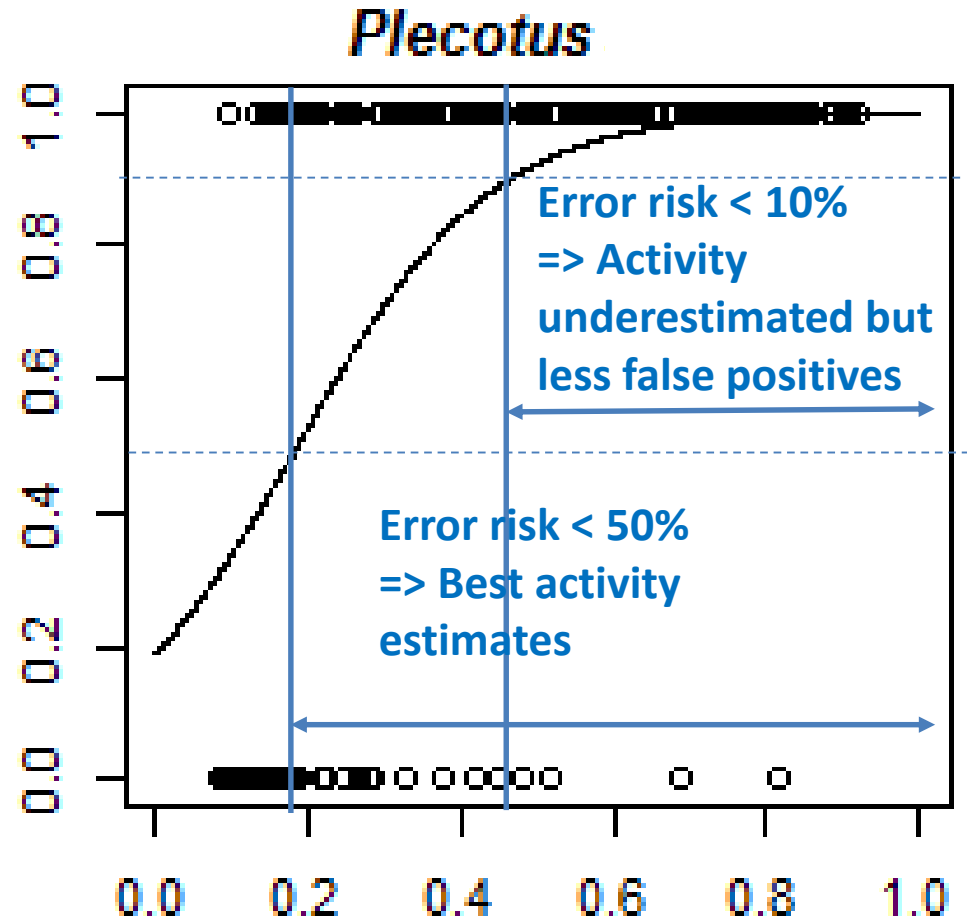
Error modelling

- Correlate error risk / confidence score
 - identifying selection thresholds

Confirmed id ~ software confidence

(Barré et al. 2019 MEE, in press)

Success probability

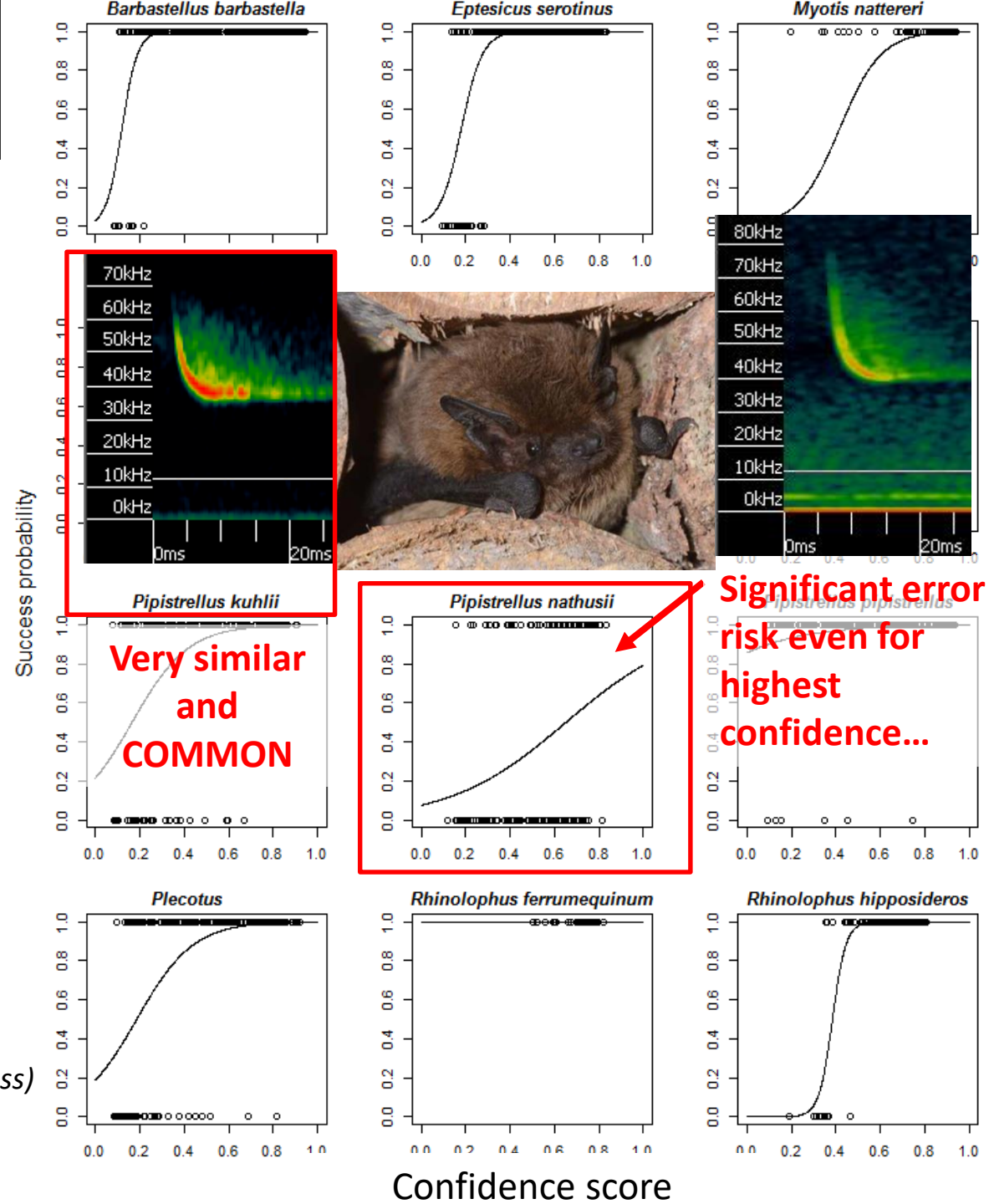


Error modelling

- Correlate error risk / confidence score
 - identifying selection thresholds

Confirmed id ~ software confidence

(Barré et al. 2019 MEE, in press)



How's automatic acoustic monitoring doing?

- Are errors biased / ecological patterns?

Error type	False negative rate	False positive rate
Causes		

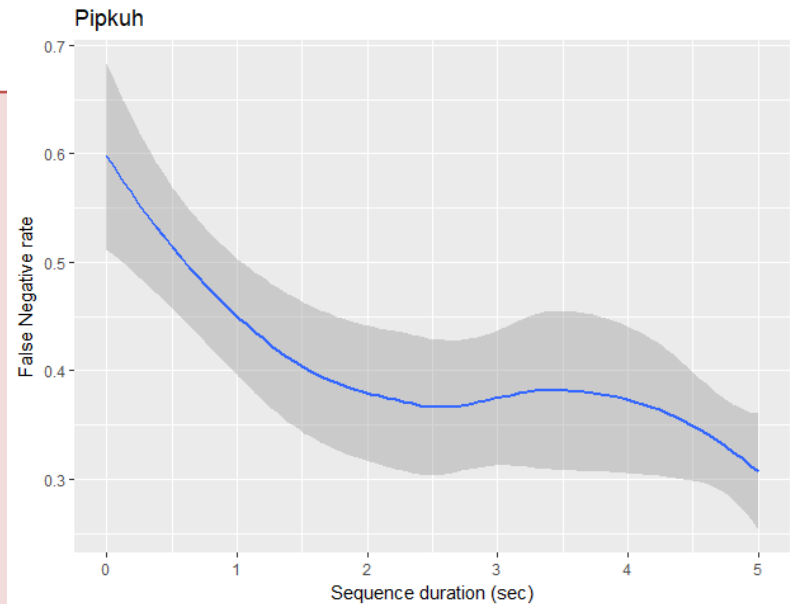
How's automatic acoustic monitoring doing?

- Are errors biased / ecological patterns?

Error type False negative rate

Causes

Mostly influenced by **sound quality**
⇒ Distance to receiver
⇒ Alteration by vegetation density
(=> habitat-dependent)



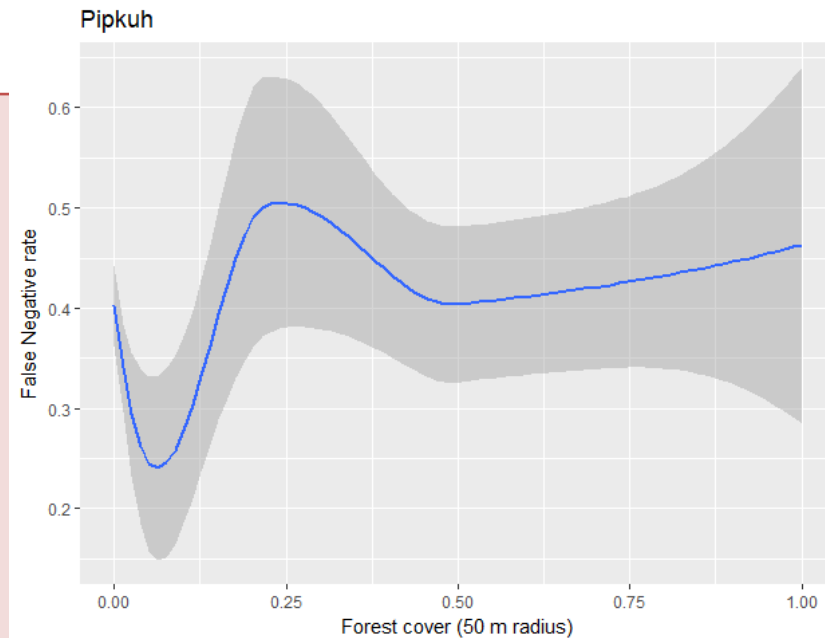
How's automatic acoustic monitoring doing?

- Are errors biased / ecological patterns?

Error type False negative rate

Causes

Mostly influenced by **sound quality**
⇒ Distance to receiver
⇒ Alteration by vegetation density
(=> habitat-dependent)



How's automatic acoustic monitoring doing?

- Are errors biased / ecological patterns?

Error type	False negative rate	False positive rate
Causes	Mostly influenced by sound quality ⇒ Distance to receiver ⇒ Alteration by vegetation density (=> habitat-dependent)	

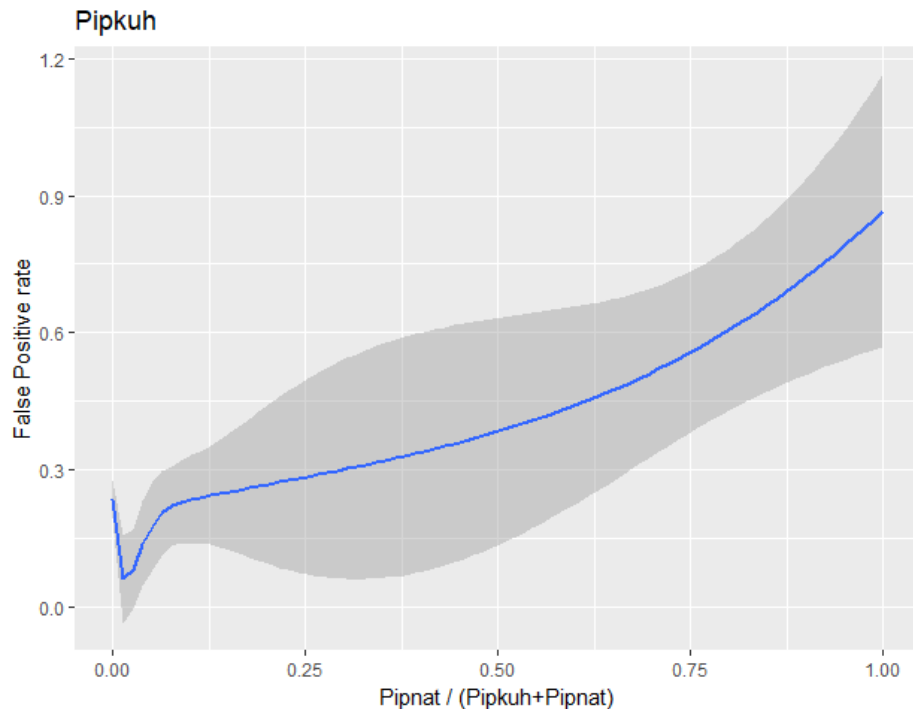
How's automatic acoustic monitoring doing?

- Are errors biased / ecological patterns?

Error type	False negative rate	False positive rate
Causes	Mostly influenced by sound quality ⇒ Distance to receiver ⇒ Alteration by vegetation density (=> habitat-dependent)	Mostly influenced by species relative abundance

How's automatic acoustic monitoring doing?

- Are errors biased / ecological patterns?

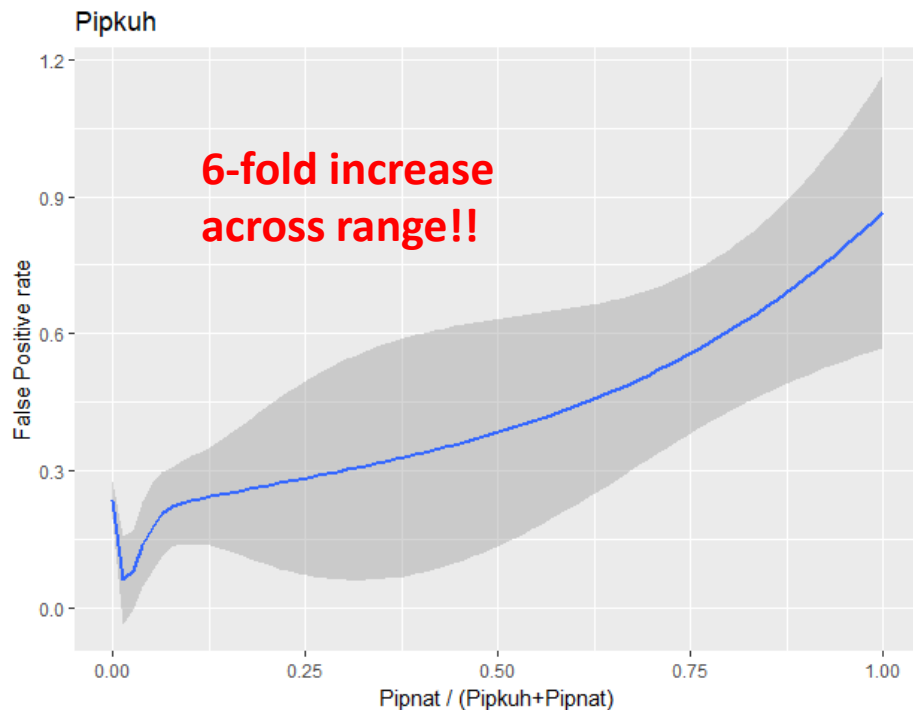


False positive rate

Mostly influenced by
**species relative
abundance**

How's automatic acoustic monitoring doing?

- Are errors biased / ecological patterns?

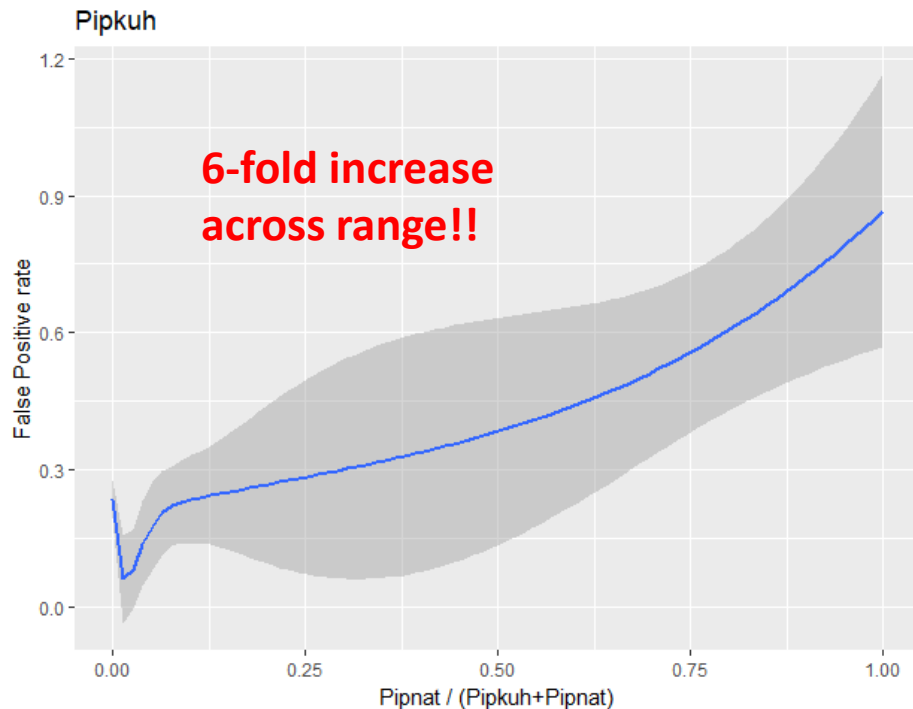


False positive rate

Mostly influenced by species relative abundance

How's automatic acoustic monitoring doing?

- Are errors biased / ecological patterns?



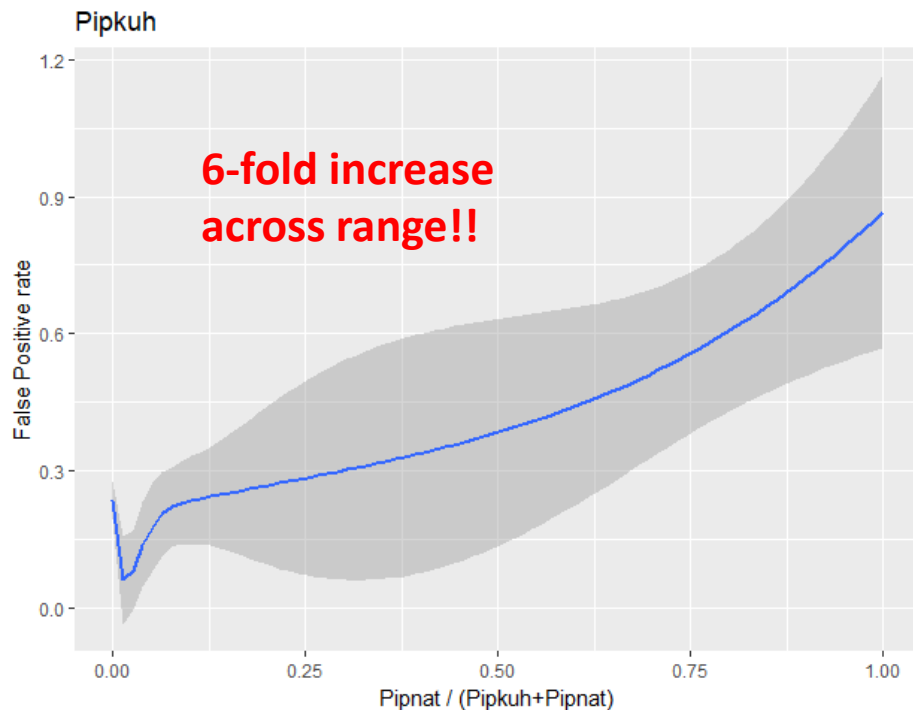
False positive rate

Mostly influenced by species relative abundance

⇒ Strongly spatially structured

How's automatic acoustic monitoring doing?

- Are errors biased / ecological patterns?



False positive rate

Mostly influenced by species relative abundance

⇒ Strongly spatially structured

⇒ Pretty bad for rare species...

How's automatic acoustic monitoring doing?

- Are errors biased / ecological patterns?

Error type	False negative rate	False positive rate
Causes	Mostly influenced by sound quality ⇒ Distance to receiver ⇒ Alteration by vegetation density (=> habitat-dependent)	Mostly influenced by species relative abundance ⇒ Strongly spatially structured ⇒ Pretty bad for rare species...

Biased?

How's automatic acoustic monitoring doing?

- Are errors biased / ecological patterns?

Error type	False negative rate	False positive rate
Causes	Mostly influenced by sound quality ⇒ Distance to receiver ⇒ Alteration by vegetation density (=> habitat-dependent)	Mostly influenced by species relative abundance ⇒ Strongly spatially structured ⇒ Pretty bad for rare species...
Biased?	may be a little	

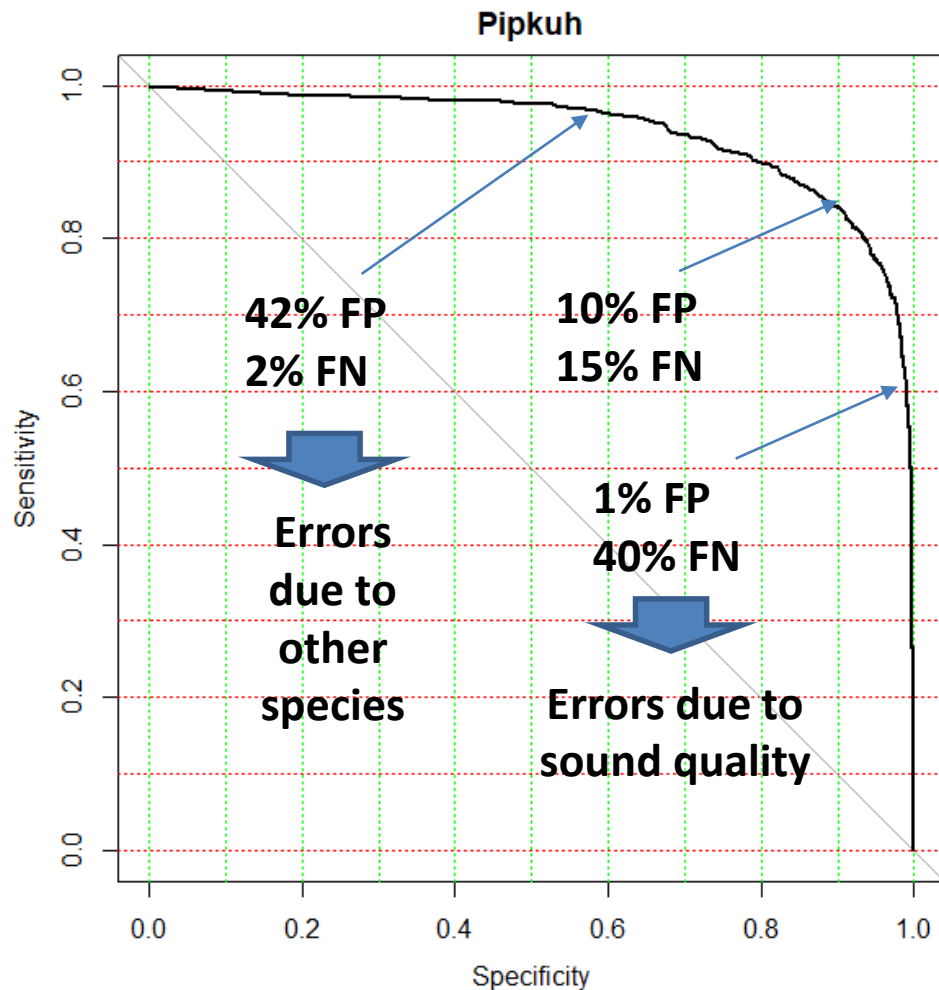
How's automatic acoustic monitoring doing?

- Are errors biased / ecological patterns?

Error type	False negative rate	False positive rate
Causes	Mostly influenced by sound quality ⇒ Distance to receiver ⇒ Alteration by vegetation density (=> habitat-dependent)	Mostly influenced by species relative abundance ⇒ Strongly spatially structured ⇒ Pretty bad for rare species...
Biased?	may be a little	Often heavily!

How's automatic acoustic monitoring doing?

- A threshold to minimise bias?

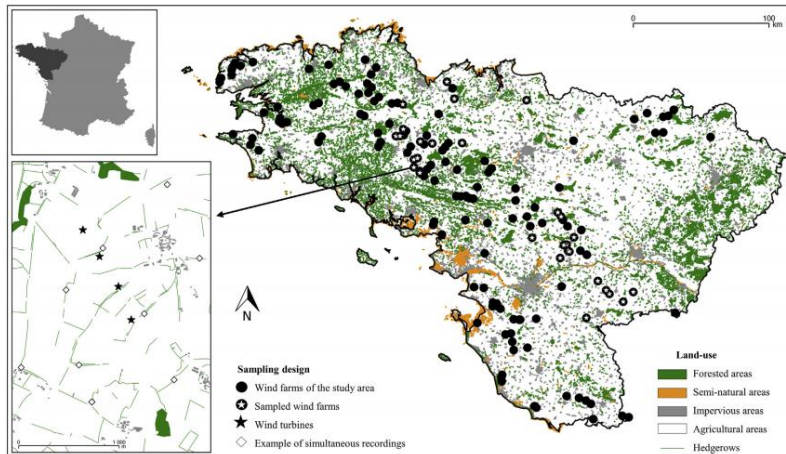


All thresholds will lead to potentially biased measures but sources of bias differ

Solution: checking consistency of ecological inference with varying thresholds (FP/FN rates)

Varying thresholds

- A study of the effect of distance to wind turbine on bats (*Barré et al. 2018 Biol Cons*)

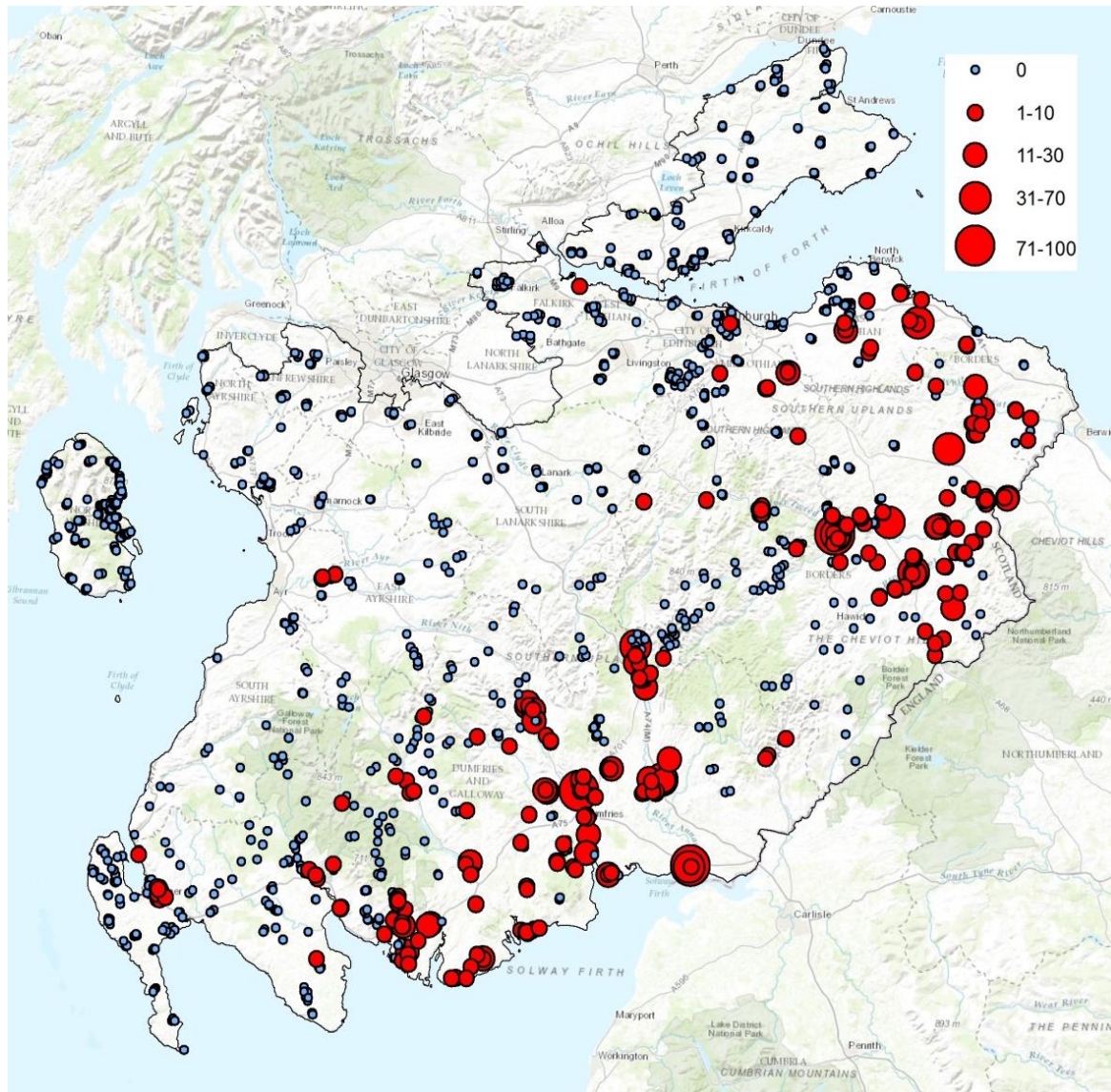


- Estimates vary little!
- ⇒ Inferences are robust against id errors!

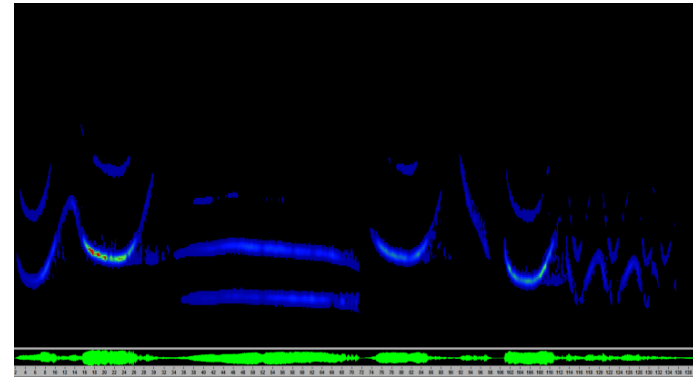
Species	Threshold type	
	FP = FN	FP << FN
<i>Barbastella barbastellus</i>	0.237 ± 0.107	0.237 ± 0.107
<i>Eptesicus serotinus</i>	0.132 ± 0.169	0.141 ± 0.179
<i>Myotis nattereri</i>	0.132 ± 0.106	0.038 ± 0.044
<i>Myotis spp.</i>	0.260 ± 0.091	0.245 ± 0.096
<i>Pipistrellus kuhlii</i>	-0.004 ± 0.100	-0.005 ± 0.103
<i>Pipistrellus pipistrellus</i>	0.413 ± 0.100	0.413 ± 0.100
<i>Plecotus spp.</i>	0.309 ± 0.096	0.233 ± 0.102

Method replicated for artificial light (*Pauwels et al. in review*), motorways (*Claireau et al. in review*), etc

Accurate data: spatial

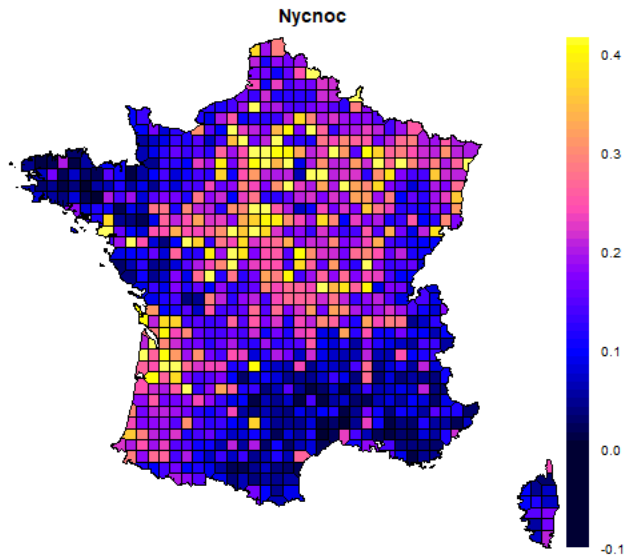


Southern Scotland Bat Survey

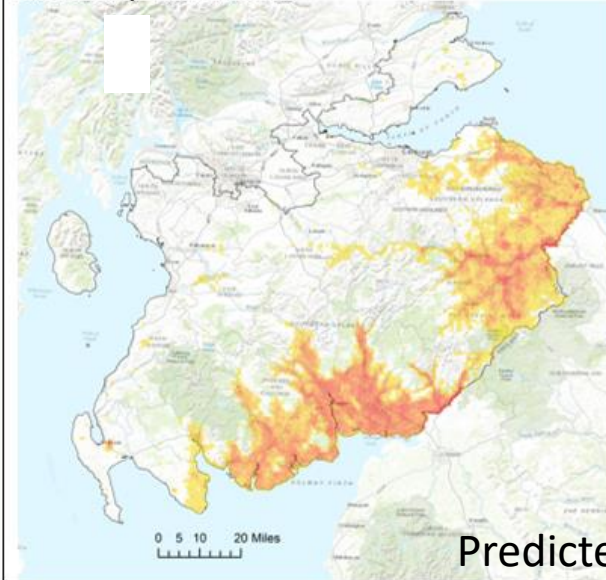


Raw data
Nyctalus noctula

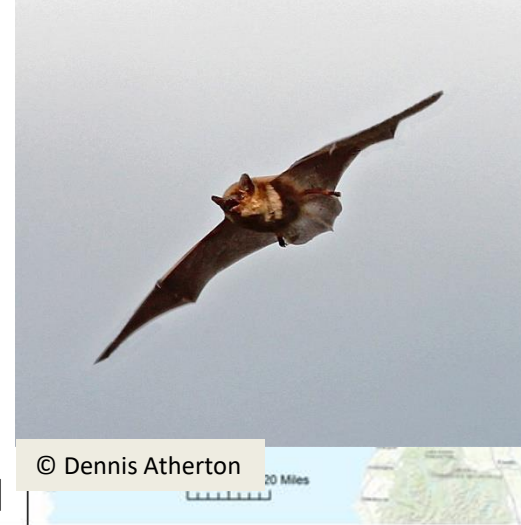
Accurate data: spatial



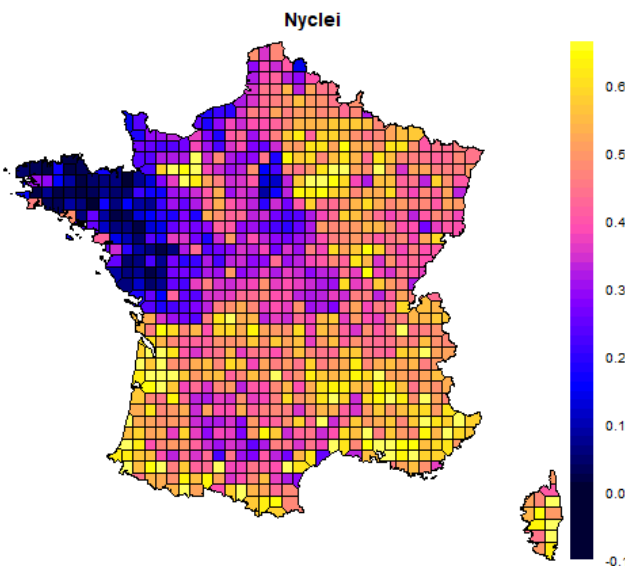
Noctule, *Nyctalus noctula*



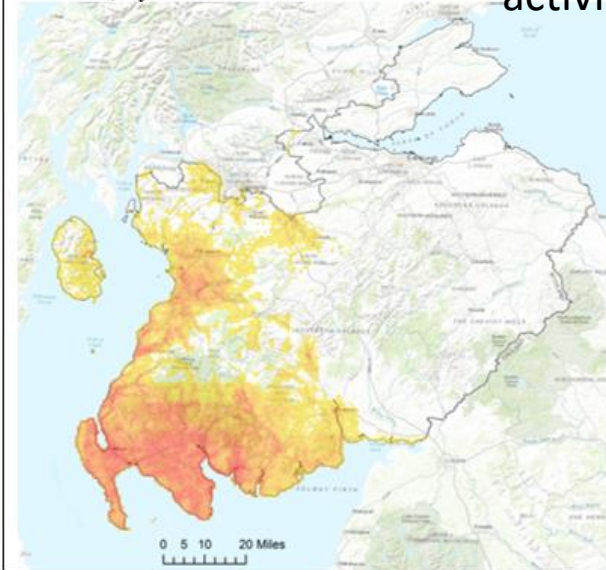
Newson et al. 2017
Biological Conservation



© Dennis Atherton



Leisler's, *Nyctalus leisleri*



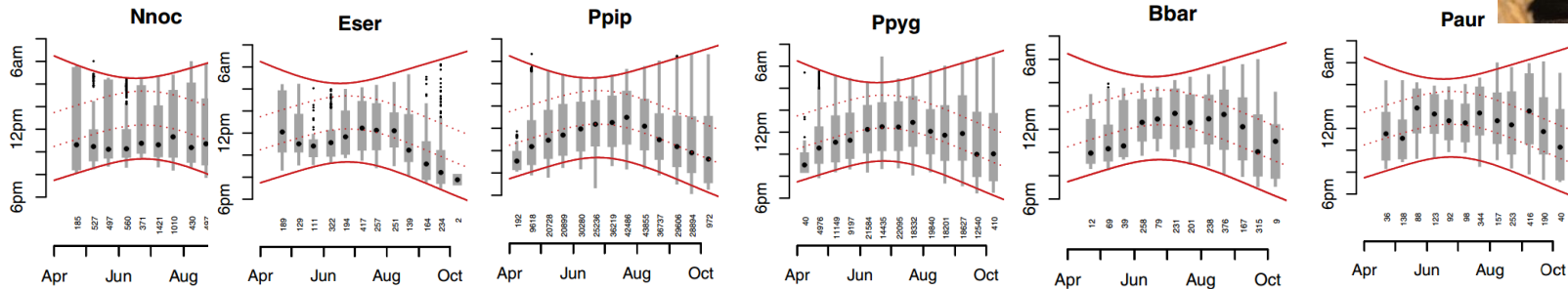
Predicted
activity



© Mark Carmody

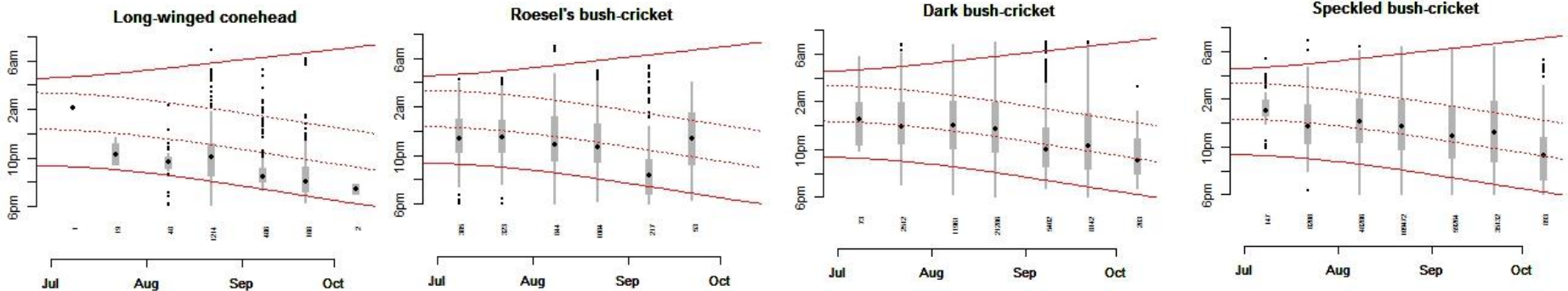
Accurate data: phenology

Newson et al. (2015)
Biological Conservation



← Crepuscular activity

→ Whole-night activity



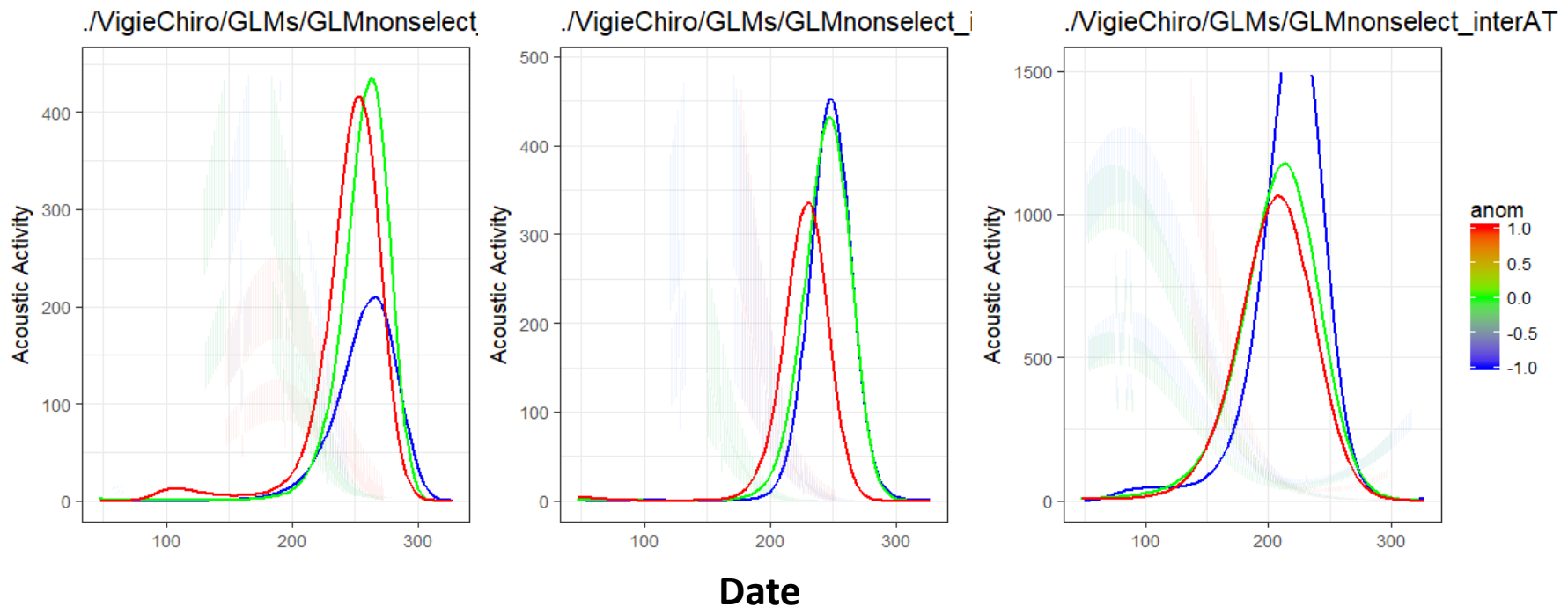
Newson et al. (2017) *Methods in Ecology and Evolution*



Accurate data: phenology

Bas et al. (in prep)

- Detecting seasonal phenological shifts



And already some species trends

Strongly declining in France



© Dennis Atherton



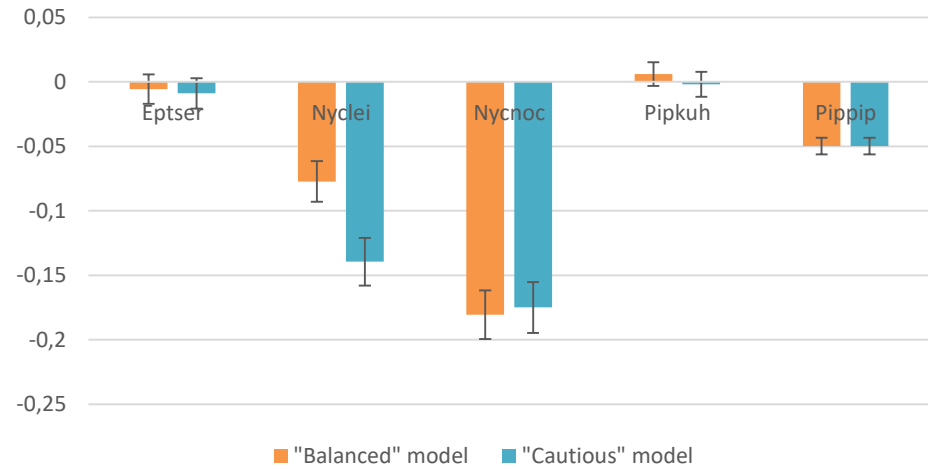
© Mark Carmody



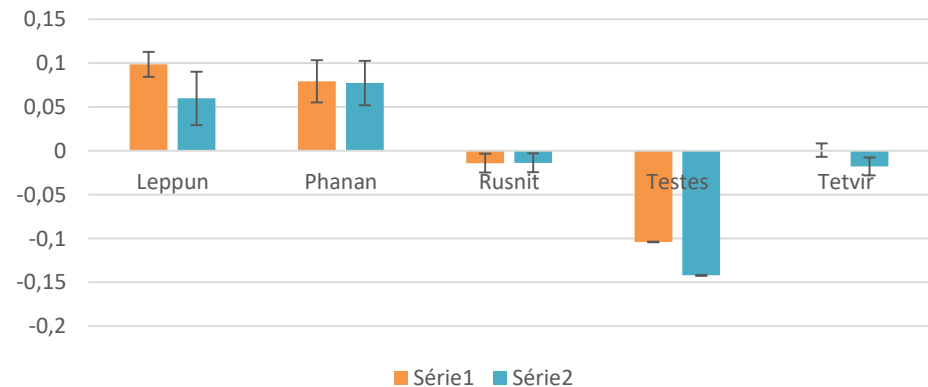
© Requira'n'h

...declines not previously suspected...

Estimated trends (2006-2016)



Estimated trends (2006-2016)



Conclusion

1) Large-scale ecoacoustic monitoring works!

- Citizen science + auto id + free web services = unprecedented data accuracy in space and time!
- Generic sound event detection = multi-taxonomic targets = sharing costs and benefits!

2) Prospects:

- Further investigating biases in acoustic data
- Improving auto id
 - More reference data => collaborative work needed
 - Adding contextual information in training (i.e. species relative abundance)

Thank you for your attention! And many thanks to participants of Vigie-Chiro, Norfolk Bat Survey and South Scotland Bat Survey!!

But, is it dangerous??



Ecological Indicators

journal homepage: www.elsevier.com/locate/ecolind

The use of automated identification of bat echolocation calls in acoustic monitoring: A cautionary note for a sound analysis

Danilo Russo^{a,b,*}, Christian C. Voigt^{c,d}

^a Wildlife Research Unit, Laboratorio di Ecologia Applicata, Sezione di Biologia e Protezione dei Sistemi
Università degli Studi di Napoli Federico II, Via Università 100, I-80055 Portici, Napoli, Italy
^b School of Biological Sciences, Life Sciences Building, University of Bristol, 24 Tyndall Avenue, Bristol 1
^c Department of Evolutionary Ecology, Leibniz Institute for Zoo and Wildlife Research, Alfred-Kowalke
^d Department of Animal Behaviour, Institute of Zoology, Freie Universität Berlin, Takustr. 6, 14195 Bei



Ecological Indicators

journal homepage: www.elsevier.com/locate/ecolind



Testing the performances of automated identification of bat echolocation calls: A request for prudence



Jens Rydell^{a,*}, Stefan Nyman^b, Johan Eklöf^c, Gareth Jones^d, Danilo Russo^{d,e}

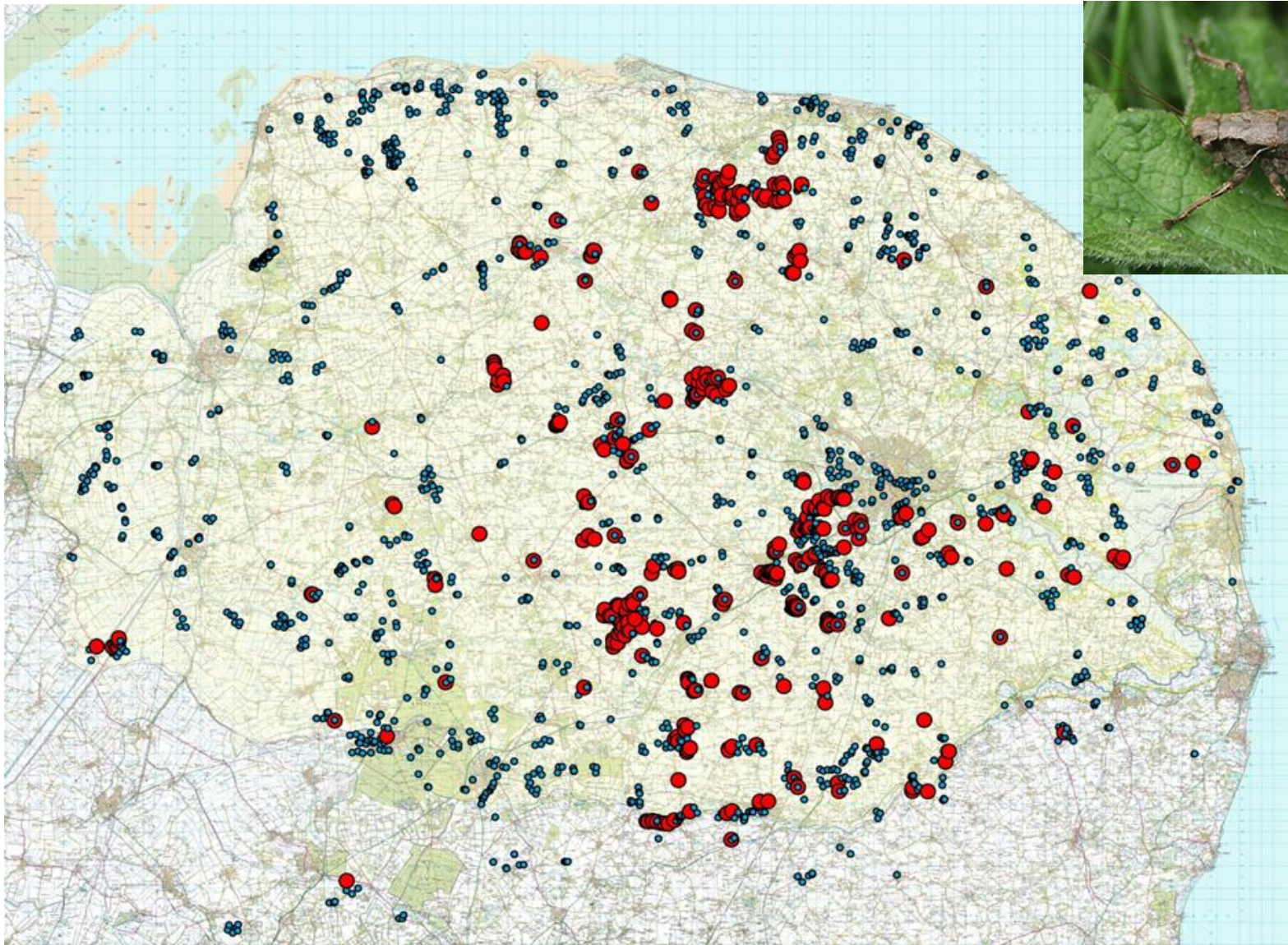
^a Biology Department, Lund University, SE-223 62 Lund, Sweden
^b Skarpskyttrevägen 30D, SE-226 42 Lund, Sweden
^c Krokalsvägen 88, SE-51734 Bollebygd, Sweden
^d School of Biological Sciences, Life Sciences Building, University of Bristol, 24 Tyndall Avenue, Bristol BS8 1TQ, UK
^e Wildlife Research Unit, Laboratorio di Ecologia Applicata, Sezione di Biologia e Protezione dei Sistemi Agrari e Forestali, Dipartimento di Agraria,
Università degli Studi di Napoli Federico II, Via Università 100, Portici (Napoli), Italy

- Well, it's obviously not perfect, so you cannot neglect error rates! You still NEED to:

- 1) Estimate error rates
- 2) Account for it in your analysis (w or w/o covariates)

=> That's what we call « semi-automatic id »

Also works for bush-crickets: Dark Bush-cricket (42,132 recordings)



Norfolk Bat Survey

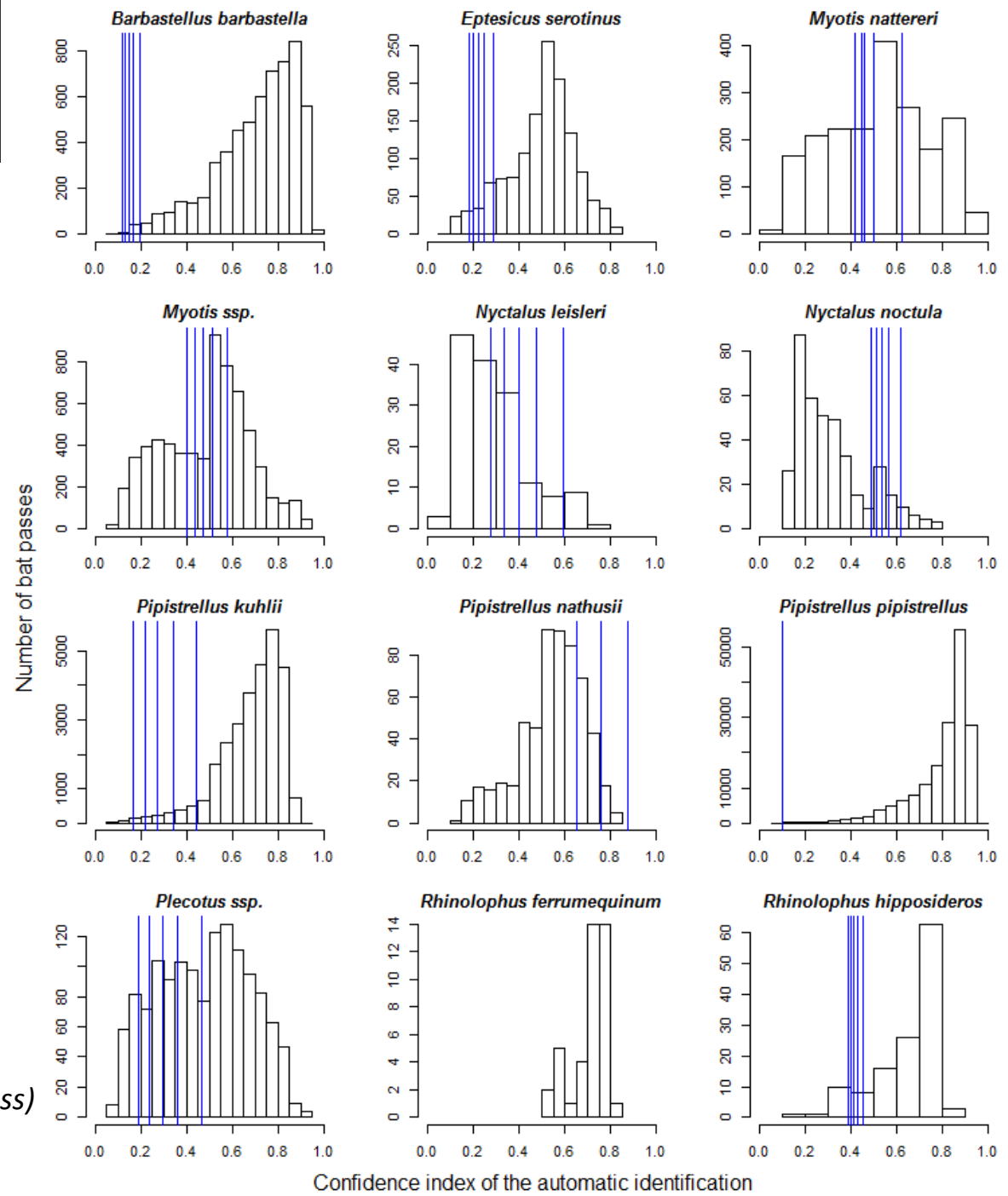
Auto id: Score reliability

- Correlate error risk / confidence score
 - identify selection thresholds

Confirmed id ~ software confidence



(Barré et al. 2019 MEE, in press)

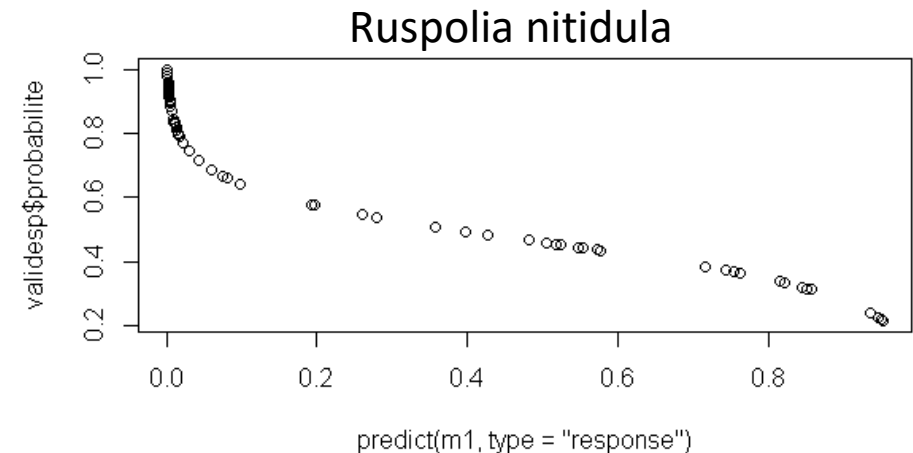
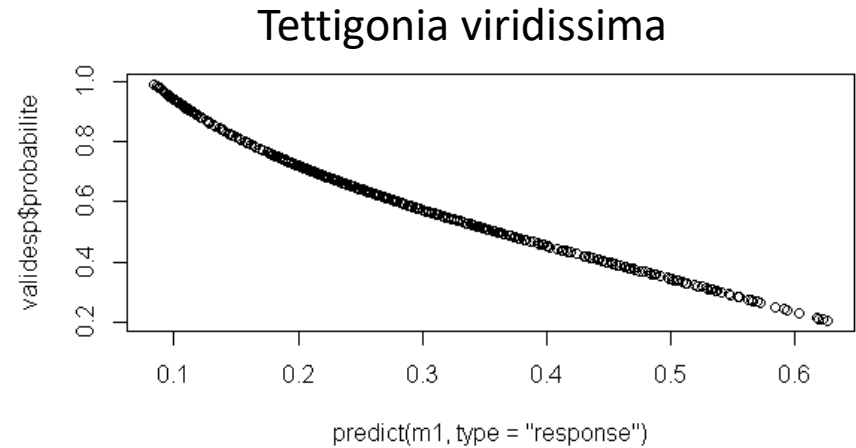


Auto id: Score reliability

- Correlate error risk / confidence score
 - identify selection thresholds

***Confirmed id ~
software
confidence***

(Barré et al. 2019 MEE, in press)



Auto id: Score reliability

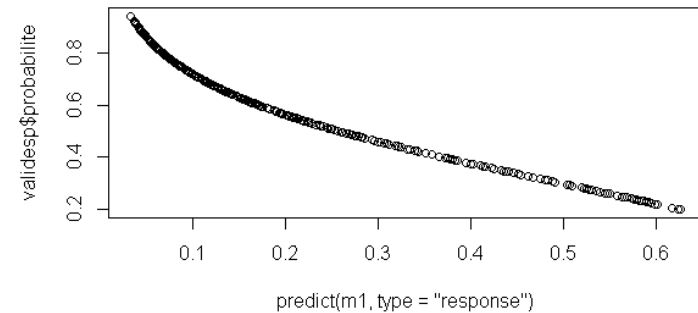
- Correlate error risk / confidence score
 - identify selection thresholds

***Confirmed id ~
software
confidence***

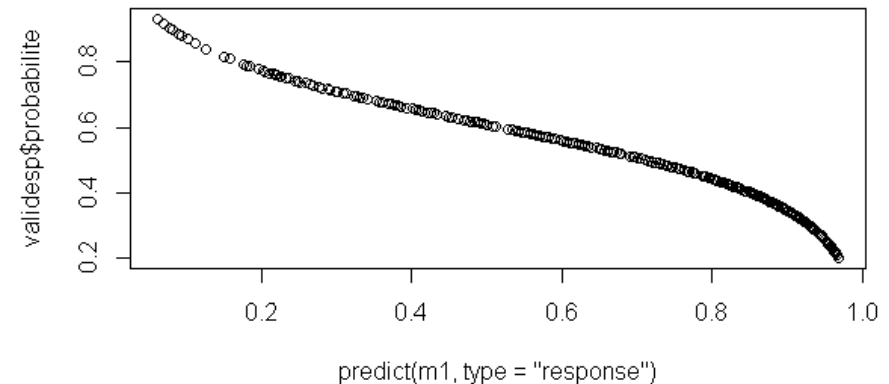
(Barré et al. 2019 MEE, in press)



Leptophyes punctatissima



Pholidoptera griseoptera



Auto id: Score reliability

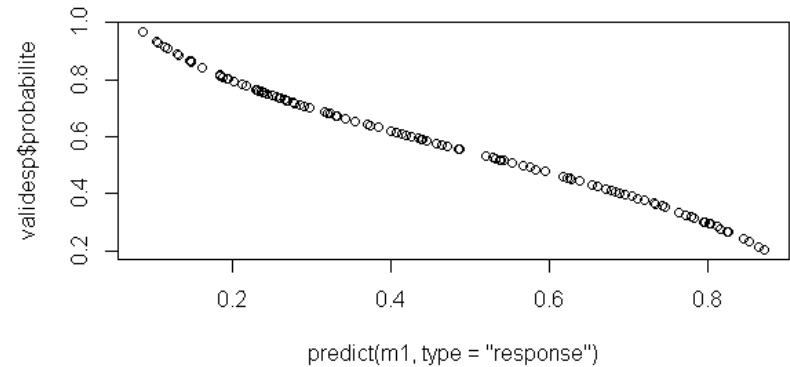
- Correlate error risk / confidence score
 - identify selection thresholds

***Confirmed id ~
software
confidence***

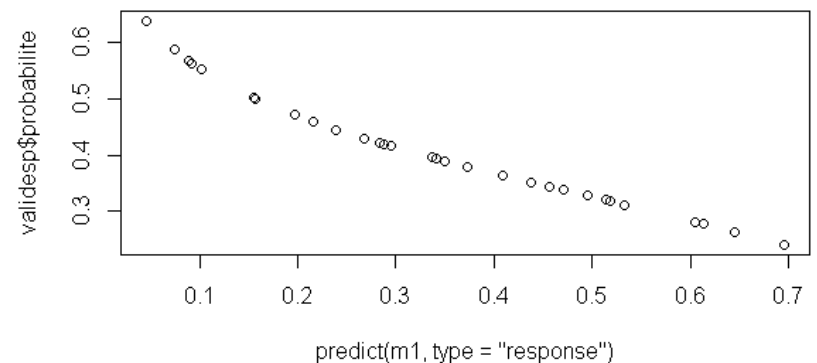
(Barré et al. 2019 MEE, in press)



Tessellana tessellata



Platycleis albopunctata



Auto id: Score reliability

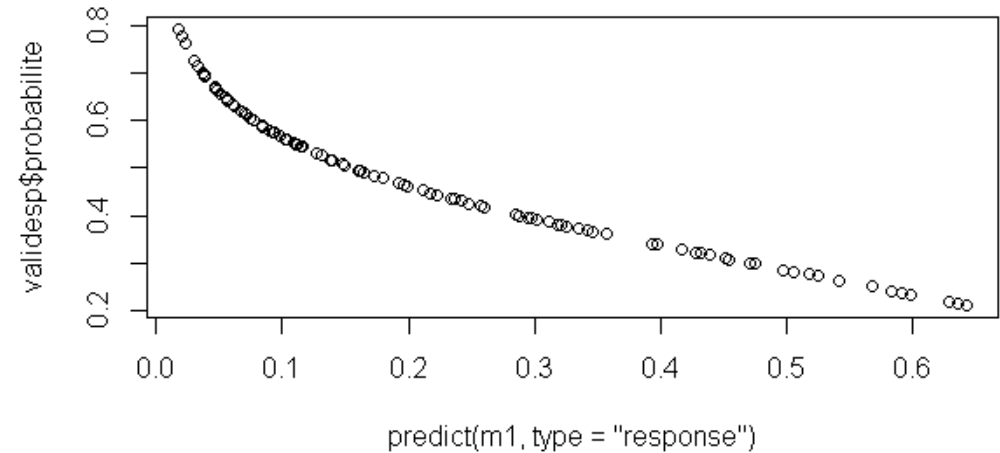
- Correlate error risk / confidence score
 - identify selection thresholds

***Confirmed id ~
software
confidence***

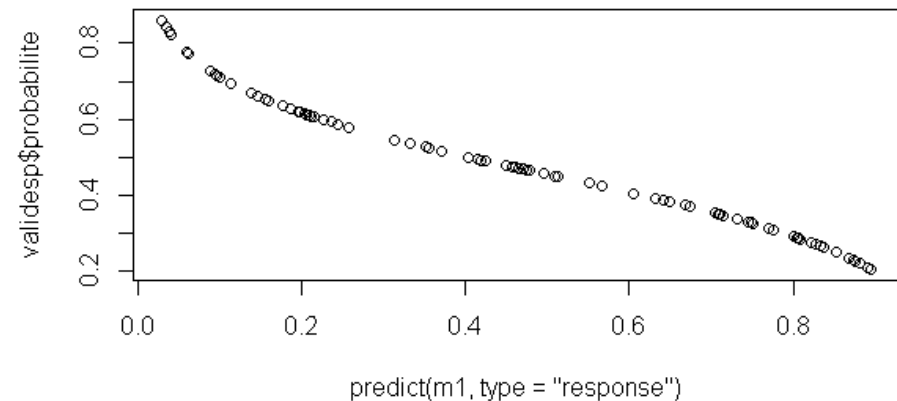
(Barré et al. 2019 MEE, in press)



Phaneroptera nana



Ephippiger ephippiger



Varying thresholds

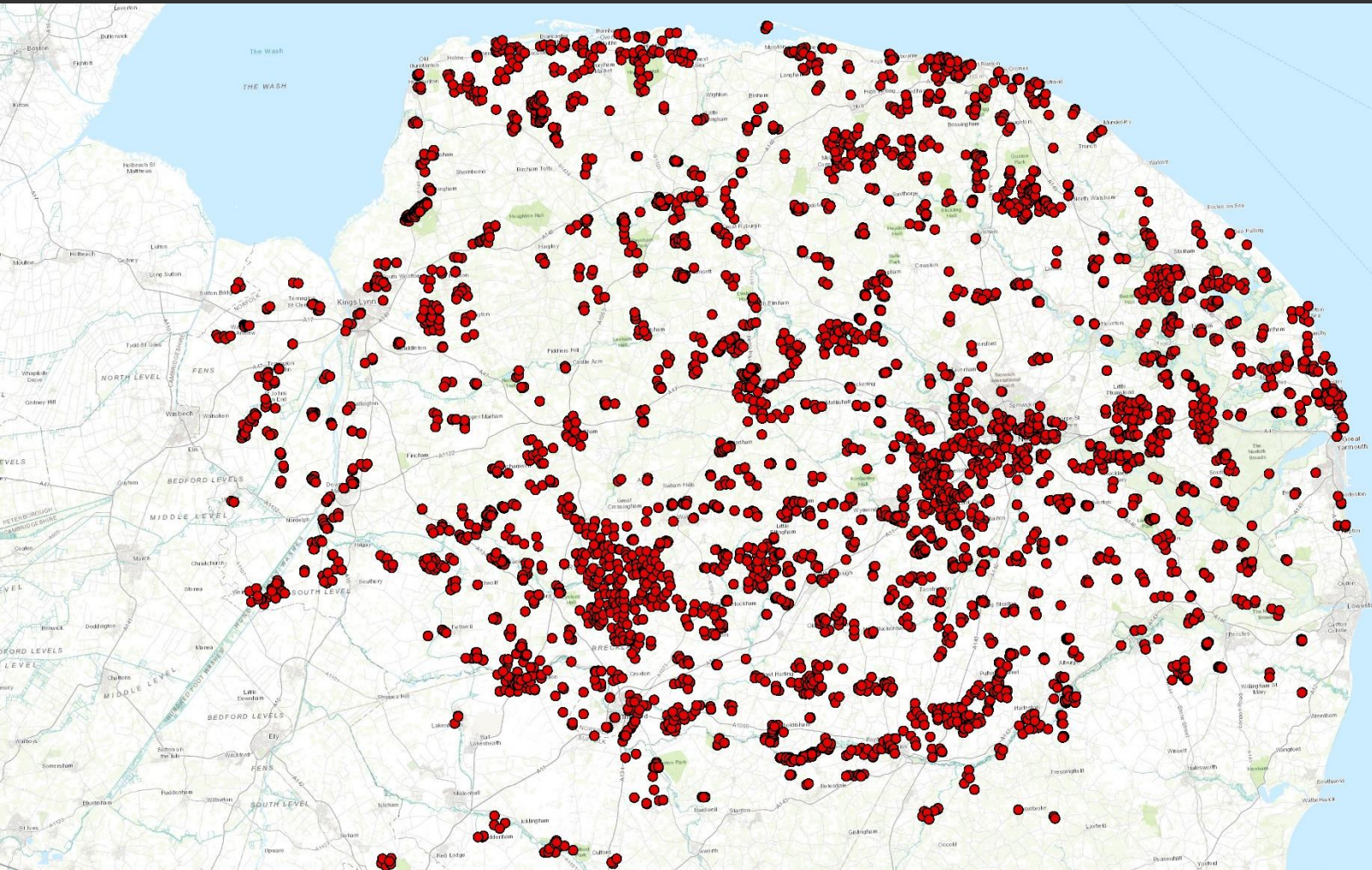
- Hedgerow effect (Barré et al. In prep)

⇒ Estimates vary little!

⇒ Inferences are robust against id errors!

Species	Environmental variables	Error risk tolerance	
		0.5	0.1
Barbastellus barbastella	Open vs. edge habitat	-2.91±0.23 ***	-2.94±0.24 ***
Eptesicus serotinus	Open vs. edge habitat	-0.60±0.40	-0.52±0.42
Myotis nattereri	Open vs. edge habitat	-1.20±0.25 ***	-1.08±0.33 ***
Myotis ssp.	Open vs. edge habitat	-1.64±0.20 ***	-1.87±0.27 ***
Nyctalus leislerii	Open vs. edge habitat	-0.41±0.29	0.92±0.66
Nyctalus noctula	Open vs. edge habitat	1.27±0.28 ***	1.27±0.50 *
Pipistrellus kuhlii	Open vs. edge habitat	-2.08±0.26 ***	-2.17±0.27 ***
Pipistrellus nathusii	Open vs. edge habitat	0.68±0.32 *	/
Pipistrellus pipistrellus	Open vs. edge habitat	-2.93±0.19 ***	-2.93±0.19 ***
Plecotus ssp.	Open vs. edge habitat	-0.89±0.19 ***	-0.81±0.20 ***
Rhinolophus ferrumequinum	Open vs. edge habitat	0.23±0.99	0.23±0.99
Rhinolophus hipposideros	Open vs. edge habitat	-3.01±0.72 ***	-2.98±0.73 ***

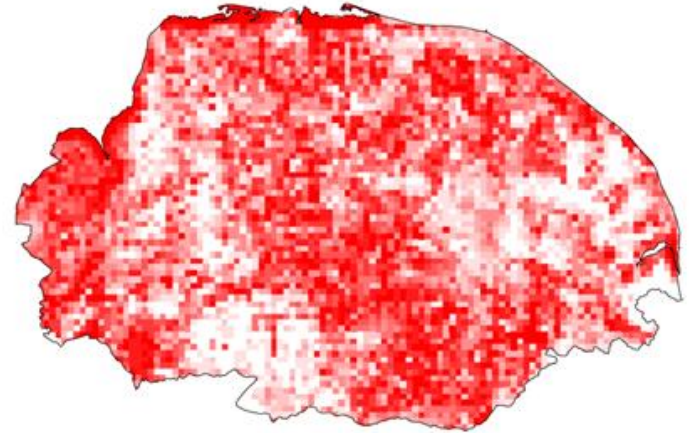
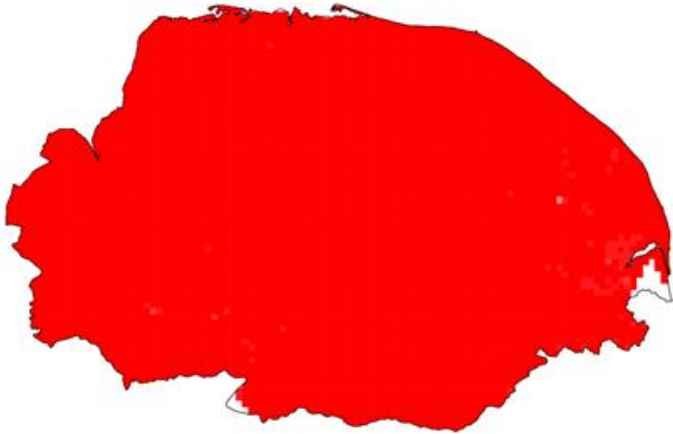
Survey coverage



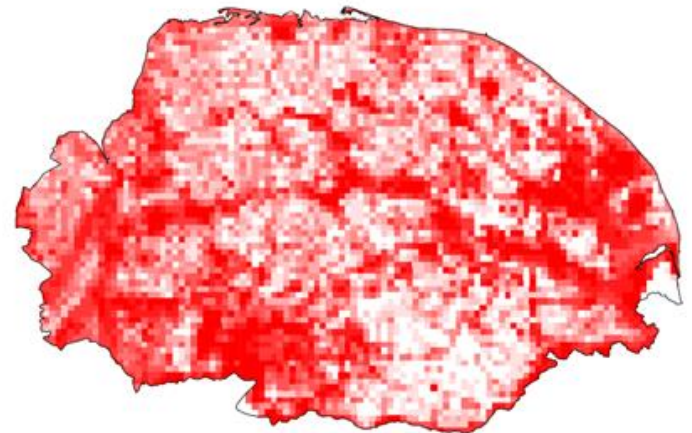
- 1,445 1-km squares surveyed (27% of Norfolk) 2013-2016
- 6,246 complete nights of recording
- > 1.4 million bat recordings

Predicted occurrence (left) and activity (right)

Common pipistrelle, *Pipistrellus pipistrellus*

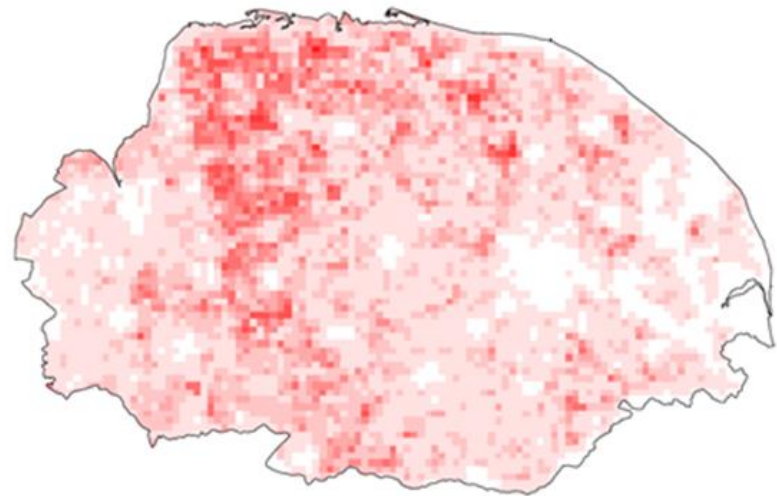
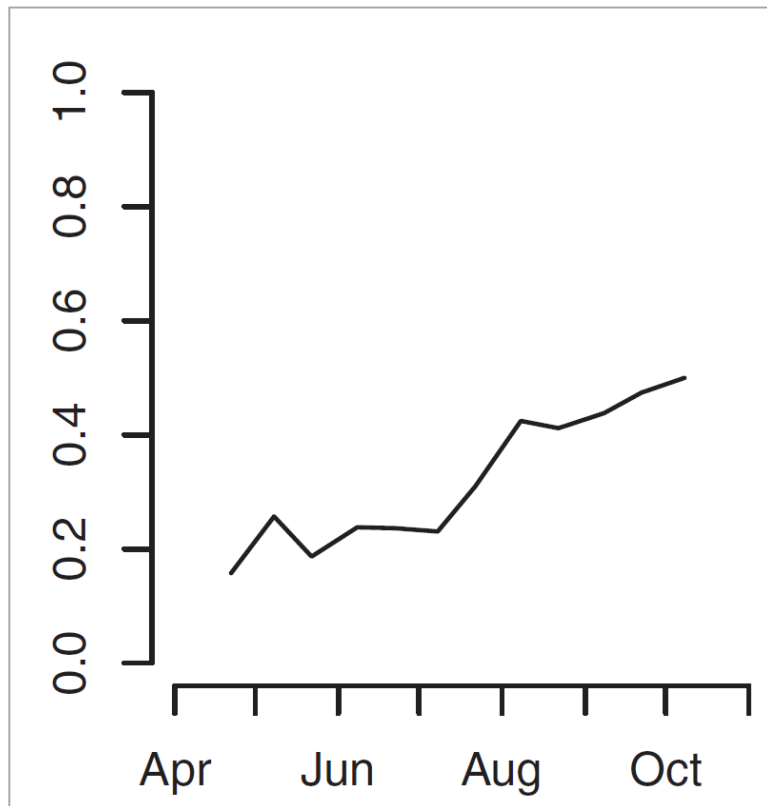


Soprano pipistrelle, *Pipistrellus pygmaeus*



Insight into seasonal movements

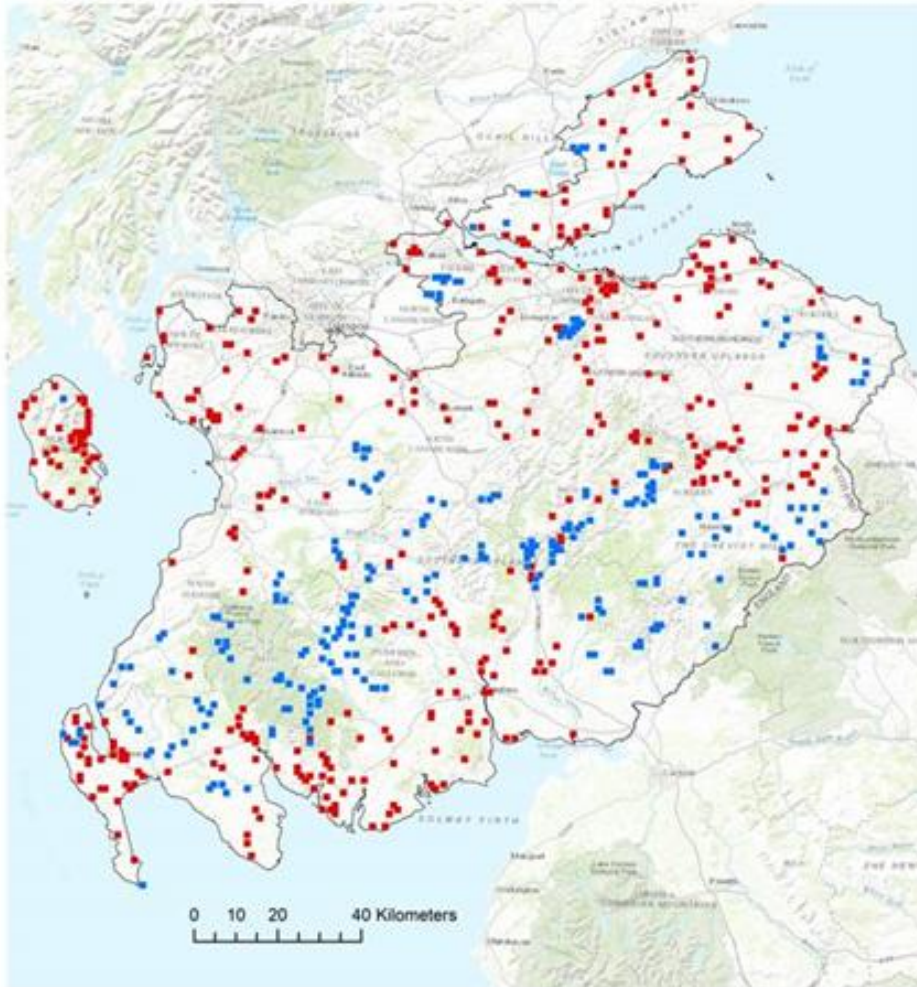
Barbastelle - *Barbastella barbastellus*



Survey coverage

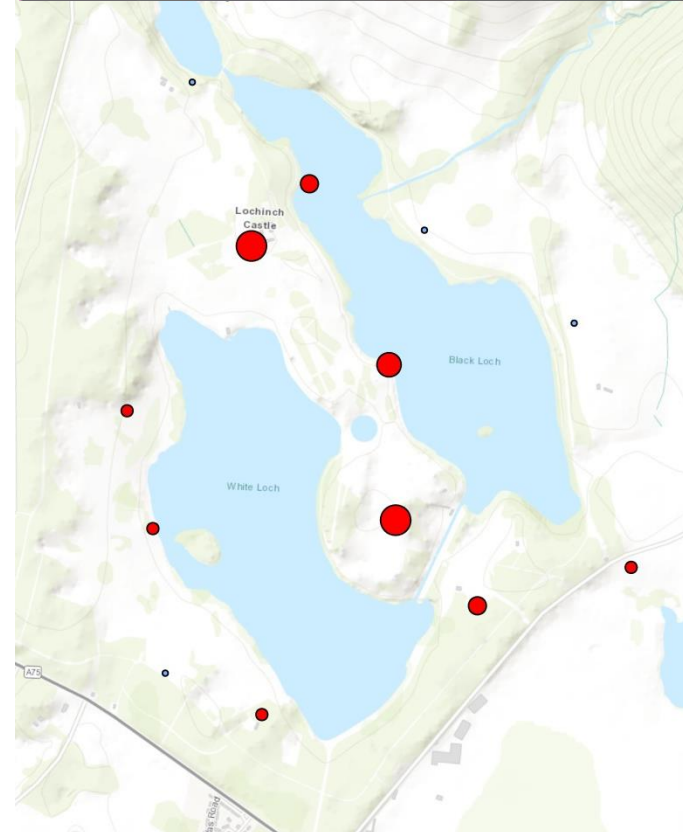
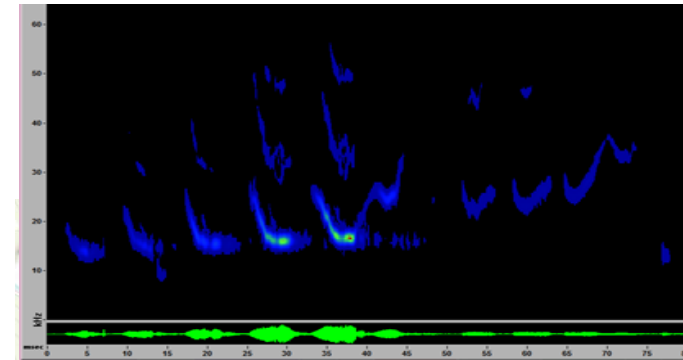
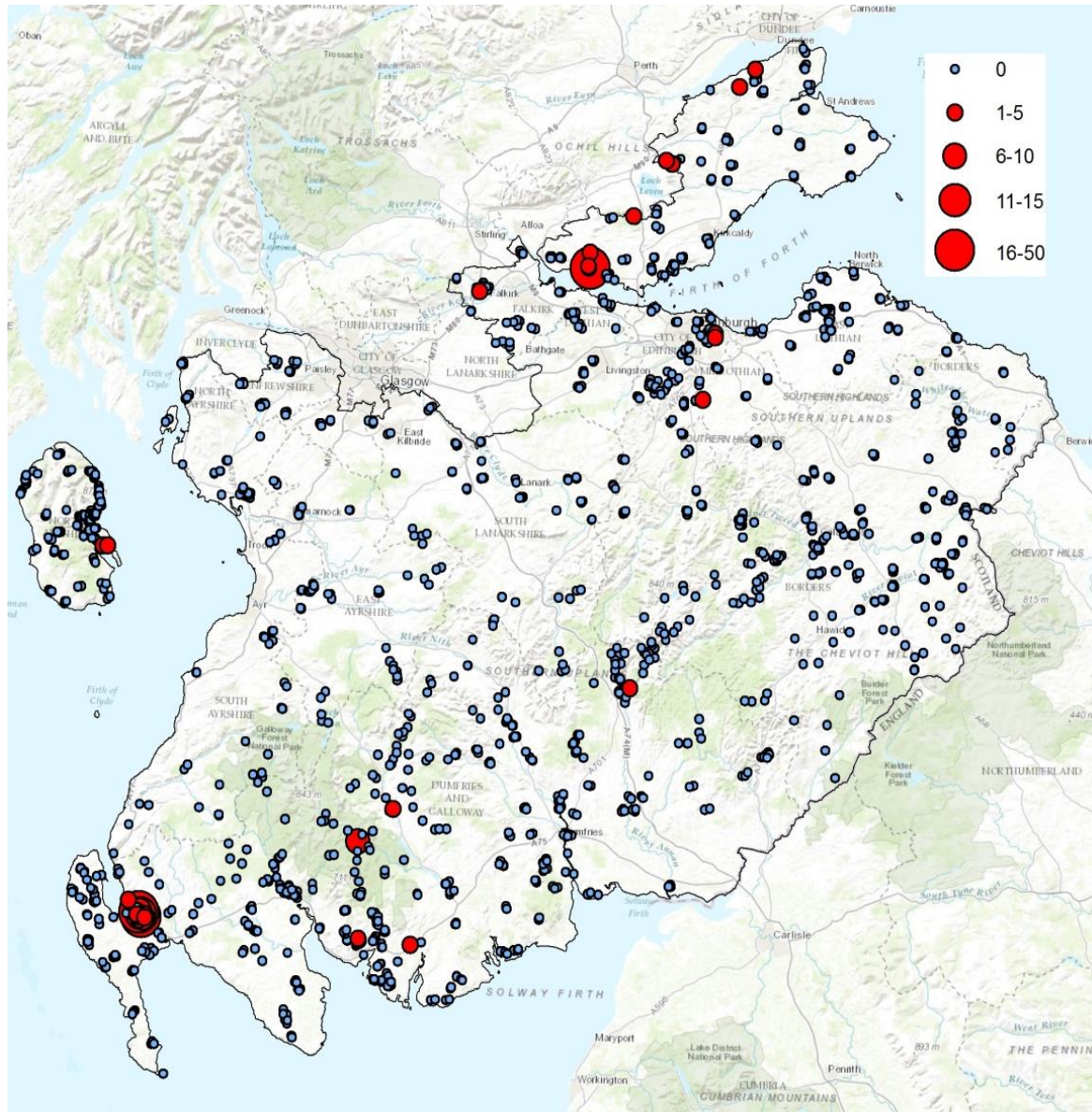
Red = Volunteers

Blue = BTO fieldworkers



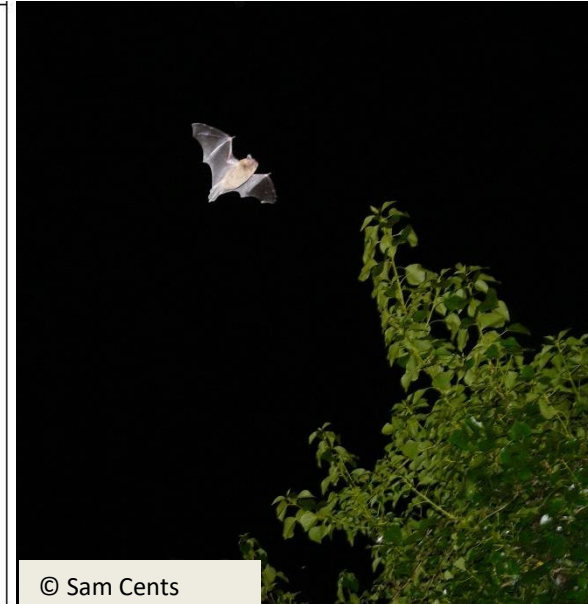
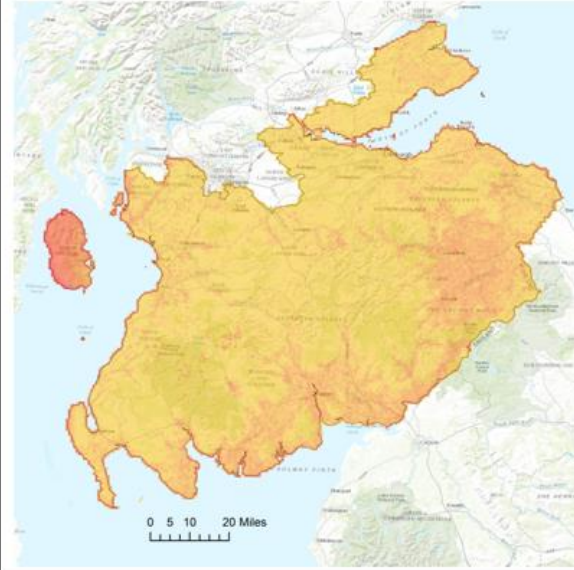
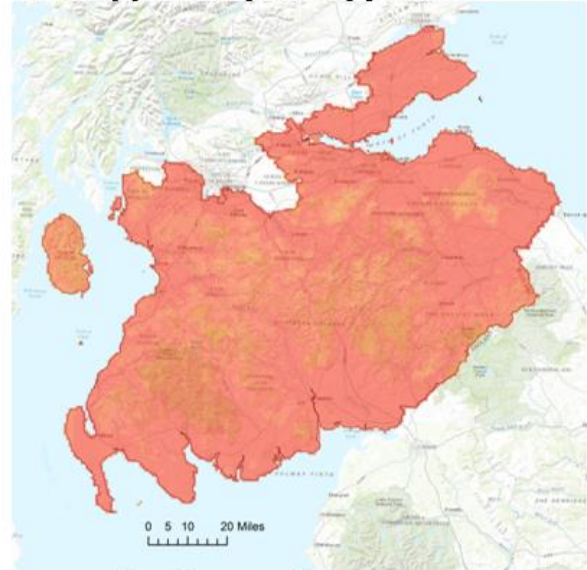
- 715 1-km's
- 1,537 nights of recording
- 399,242 bat recordings
- 275 volunteers - 375 squares
- Two BTO fieldworkers - 339 squares

Nathusius' pipistrelle (0.05% of recordings)



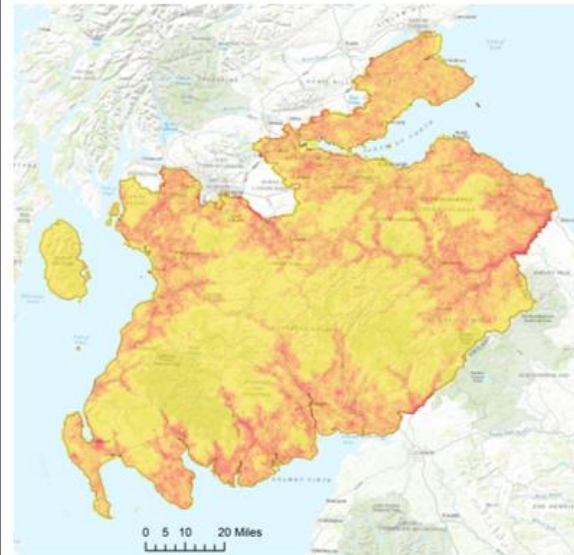
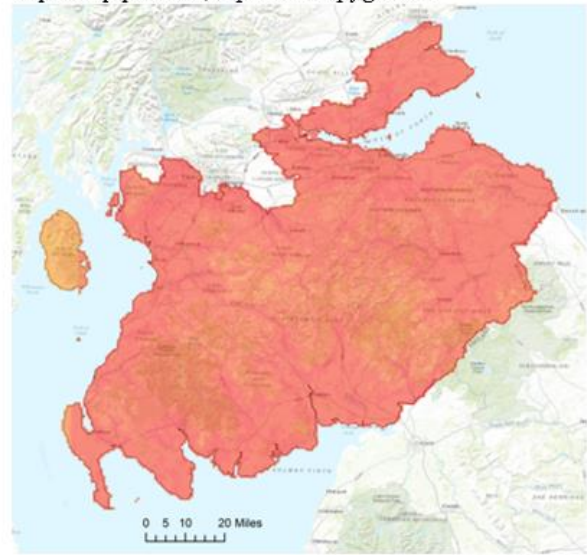
Predicted occurrence (left) and activity (right)

Common pipistrelle, *Pipistrellus pipistrellus*



© Sam Cents

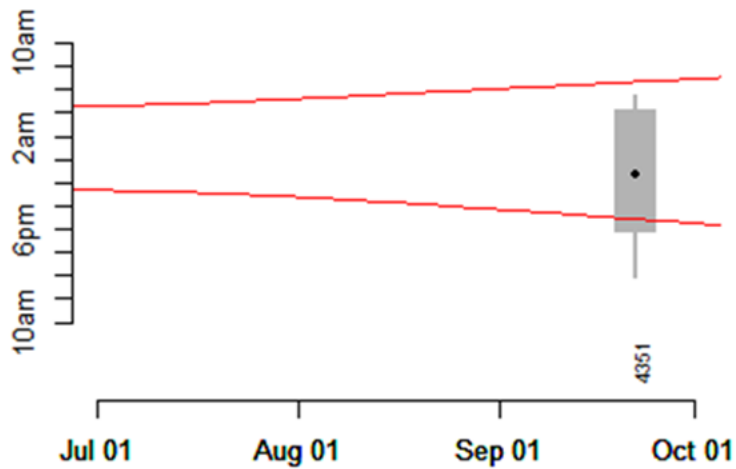
Soprano pipistrelle, *Pipistrellus pygmaeus*



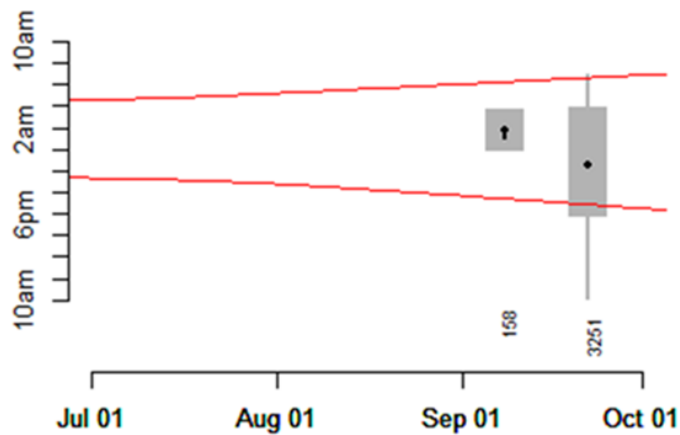
© John Dixon

Detectors recording over the day and night

Dark bush-cricket

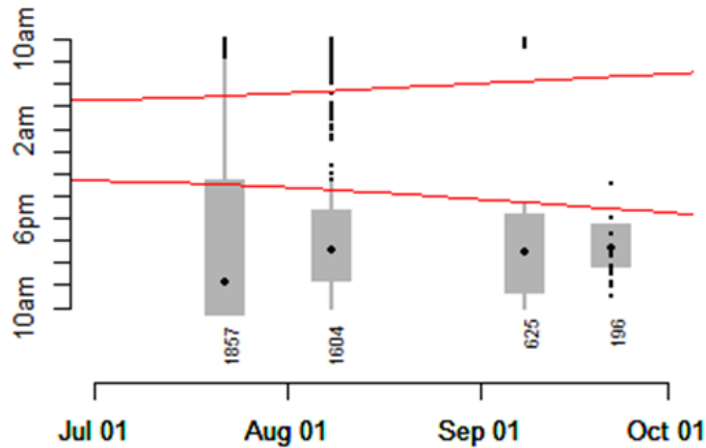


Speckled bush-cricket

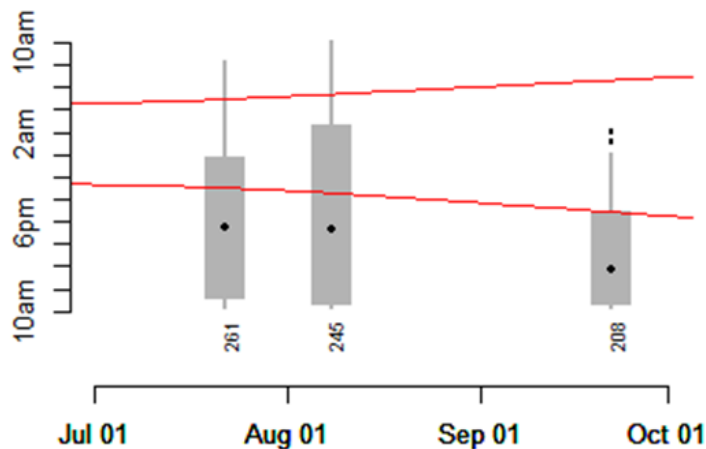


Detectors recording over the day and night

Short-winged Conehead

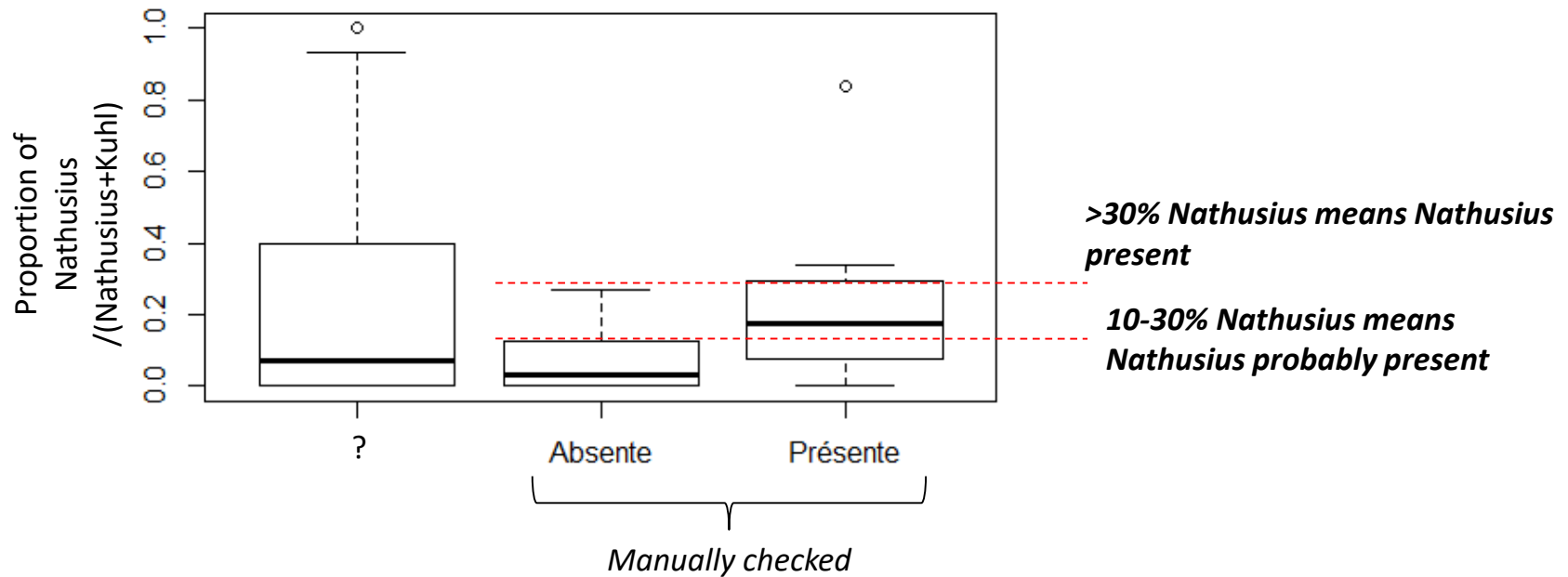


Roesel's bush-cricket



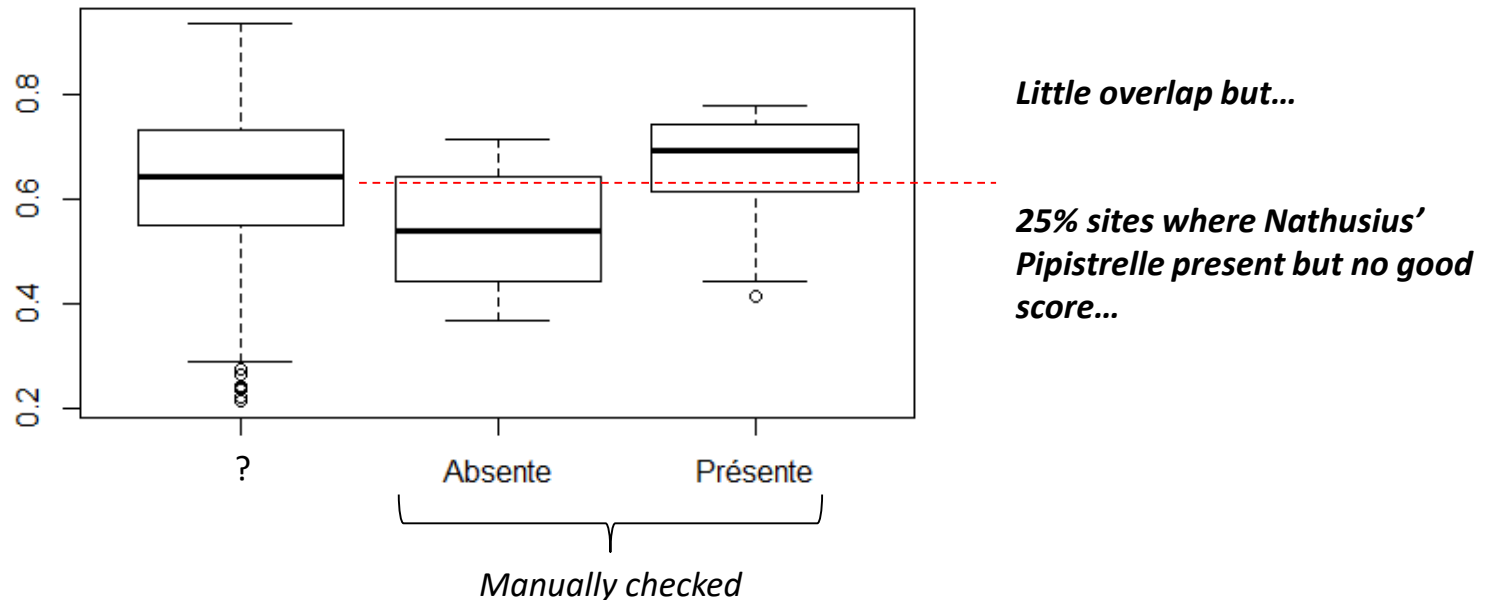
Limits of the "auto id"

- Solution: looking at other « auto id » results on the same location
 - 1) Rate of Nathusius' positive id among « Kuhl's + Nathusius' »



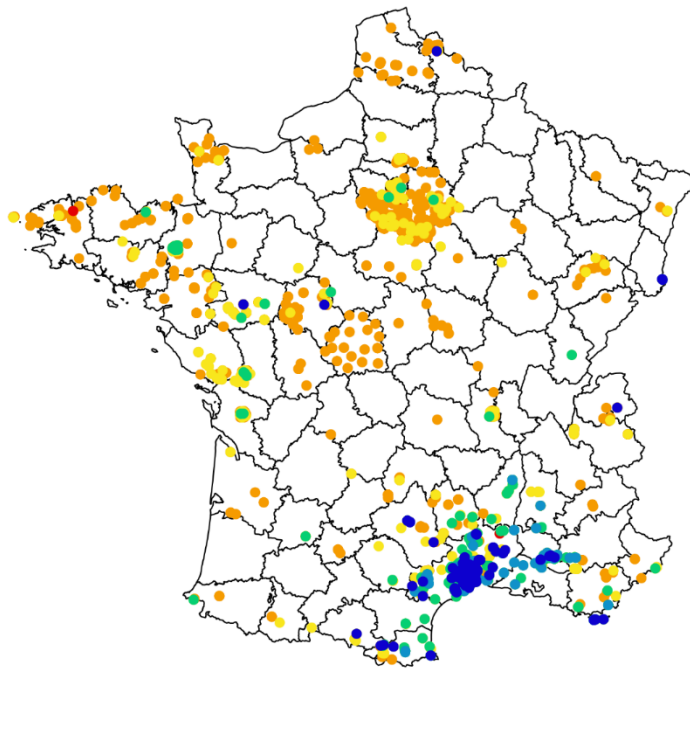
Limits of the "auto id"

- Solution: looking at other « auto id » results on the same location
2) Maximum random forest score among Nathusius' positive id



More complex modelling in progress: integrating features measured at several temporal scale (call sequence, minute, hour, night, etc) = 2nd layer of classification

Pipistrelle soprane : présence-absence

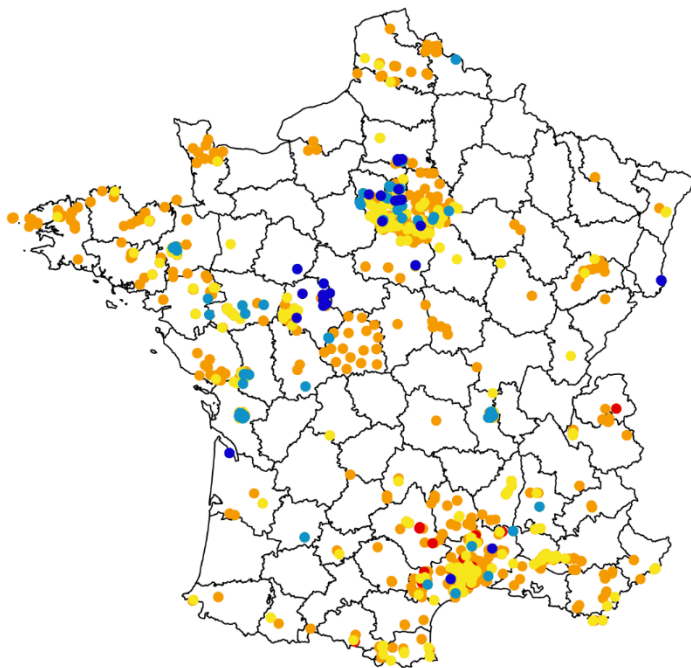


**Nombreux faux positifs dans le Nord,
mais la plupart peuvent être
discriminés par un score faible**

Légende

- Présence vérifiée manuellement
- Présence très probable ($p > 0.97$)
- Présence probable ($p > 0.9$)
- Présence à vérifier ($0.55 < p < 0.8$)
- Absence probable (non détectée ou $p < 0.55$)
- Absence vérifiée manuellement

Noctule commune : présence-absence



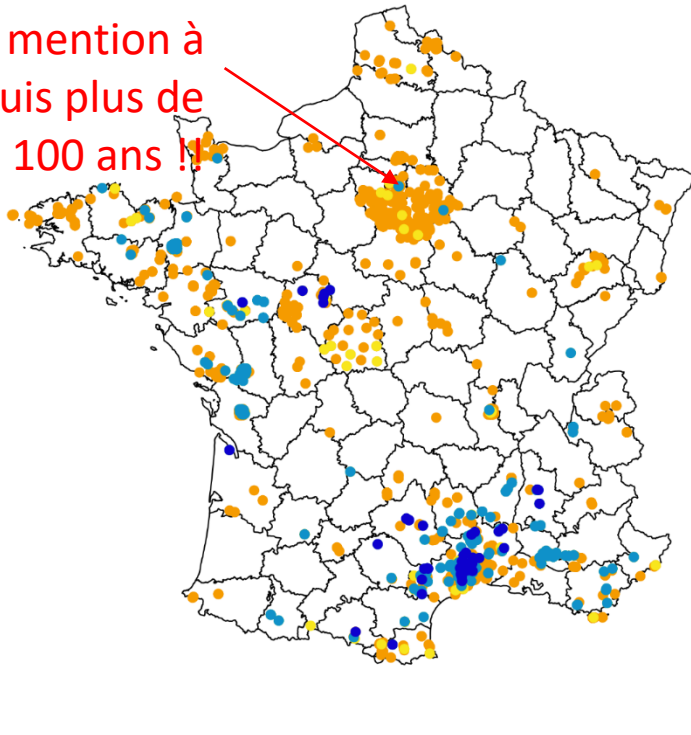
La plupart des faux positifs peuvent être discriminés par un score faible

Légende

- Présence vérifiée manuellement
- Présence très probable ($p > 0.75$)
- Présence à vérifier ($0.4 < p < 0.75$)
- Absence probable (non détectée ou $p < 0.4$)
- Absence vérifiée manuellement

Petit Rhinolophe : présence-absence

1ere mention à
Paris depuis plus de
100 ans !!



**La plupart des faux positifs peuvent
être discriminés par un score faible**

Légende

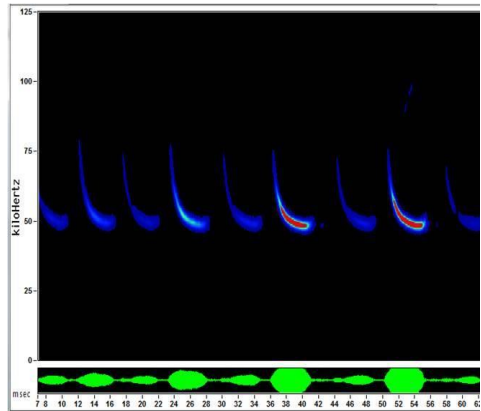
- Présence vérifiée manuellement
- Présence très probable ($p > 0.5$)
- Présence à vérifier ($p < 0.5$)
- Absence probable (non détectée)
- Absence vérifiée manuellement

Data collection, analysis, feedback pipeline



Data collection

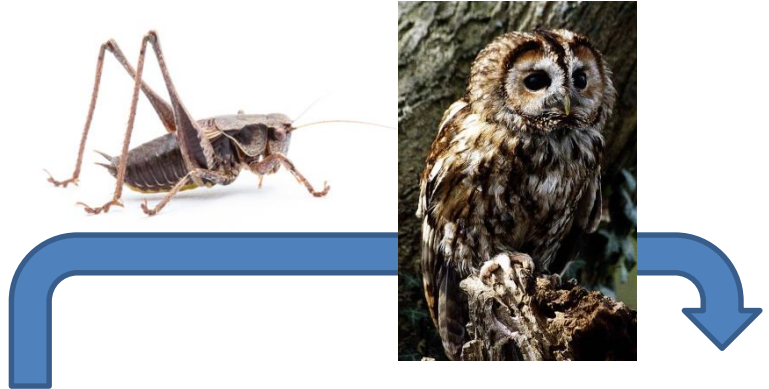
Data transfer



Data processing



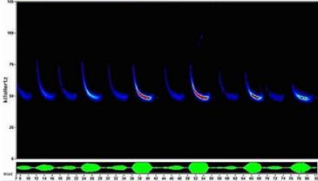
Data storage



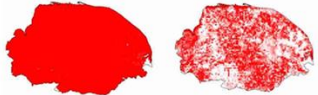
Common pipitrelle—peak frequency about 45 kHz

The Common Pipitrelle is the most common and widespread of British bat species. They appear fast and jerky in flight as they dodge about pursuing small insects which the bats catch and eat on the wing. A single Common Pipitrelle can consume up to 3,000 insects in a night.

Photo credit: Amy Lavan



Maps of predicted distribution (left) and activity as a measure of relative abundance (right)



Taken from Newton et al. (2015) and derived from data collected by volunteers through the Norfolk Bat Survey (www.batforums.co.uk)

Feedback